

DTU Health Technology Bioinformatics

Week 8 Recap

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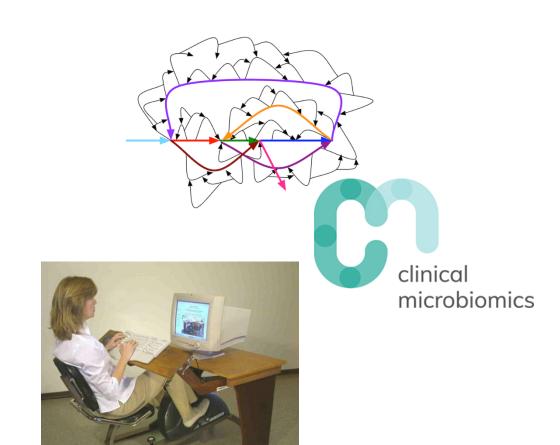
Last Week...last year...





Two weeks ago...

- Metagenomics de novo assembly
- Nonpareil exercises





Q1: What is nonparell and what is it used for?

- Nonpareil is used to estimate the coverage of metagenome datasets.
- It uses the redundancy of the reads in metagenomic datasets to estimate the average coverage and predict the amount of sequences that will be required to achieve "nearly complete coverage".



Q2: Why do we use the R1 reads to ensure higher quality?

• We generally see that the R2 reads will be of lower quality, due to imprecisions on the sequencing machines for R2.



Q3: Briefly explain what the Rscript is used for

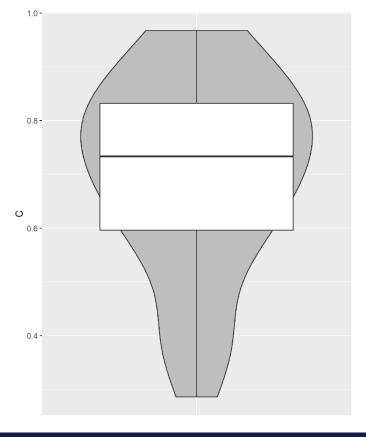
- It collects the nonpareil results for all samples and creates a summary.
- Returns a matrix with the following values for the dataset:
- kappa: "Redundancy" value of the entire dataset.
- C: Average coverage of the entire dataset.
- LRstar: Estimated sequencing effort required to reach the objective average coverage (star, 95)
- LR: Actual sequencing effort of the dataset.
- modelR: Pearson's R coefficient betweeen the rarefied data and the projected model.
- diversity: Nonpareil sequence-diversity index (Nd). This value's units are the natural logarithm of the units of sequencing effort (log-bp), and indicates the inflection point of the fitted model for the Nonpareil curve. If the fit doesn't converge, or the model is not estimated, the value is zero (0).



Q4: What can you tell about the coverage?

• So in general we have pretty good (more than 50% of the metagenome) coverage of most

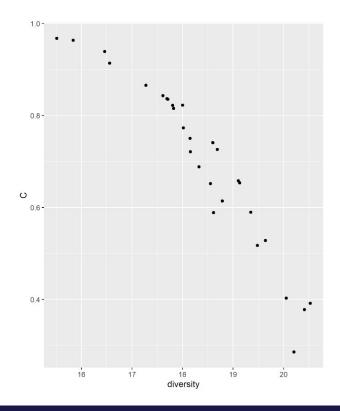
samples





Q5: Do the coverage correlate with the diversity?

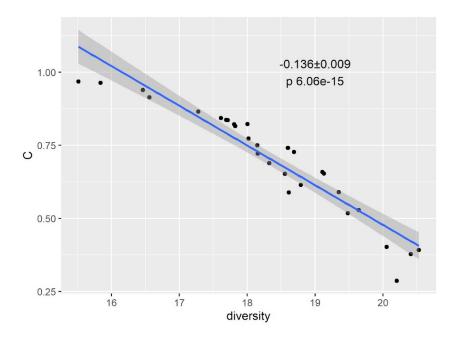
• Not surprisingly there is an obvious negative correlation between diversity of metagenome and coverage since all samples were sequenced with similar number of reads.





Q6: Do the model support your answer from Q5?

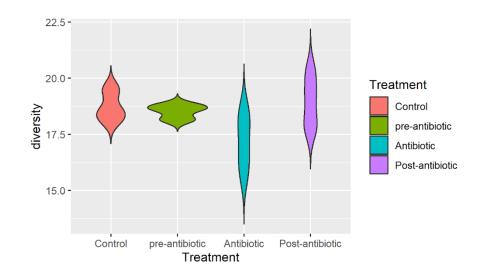
• Yes, we see that the coverage is negatively correlated with the diversity.





Q7: What do we see on the violin plot? How do the treatment affect the diversity?

 So metagenome diversity seems lowest during antibiotic-treatment and significantly lower compared to post-antibiotic treatment samples. I am fairly certain that with more samples, we would have significantly lower diversity in the fish being treated with antibiotics compared with all three other conditions.





De novo assembly

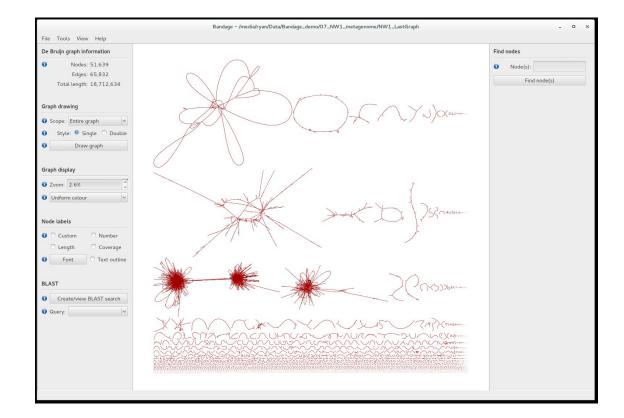
- Metagenomics de novo assembly is even harder
- Currently we use de Bruijn graphs to assemble





De Bruijn graphs

- Really struggles with repetitive regions
 - Horisontally transferred regions
 - Closely related strains
 - Etc.





Today

- Kaiju exercise
- Metagenomic binning
- Start the project work!



