

# Ancient DNA

Next Generation Sequencing Analysis

DTU, 9/1/2026



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University of Copenhagen  
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# 1984 - the birth of ancient DNA

NATURE VOL. 312 15 NOVEMBER 1984

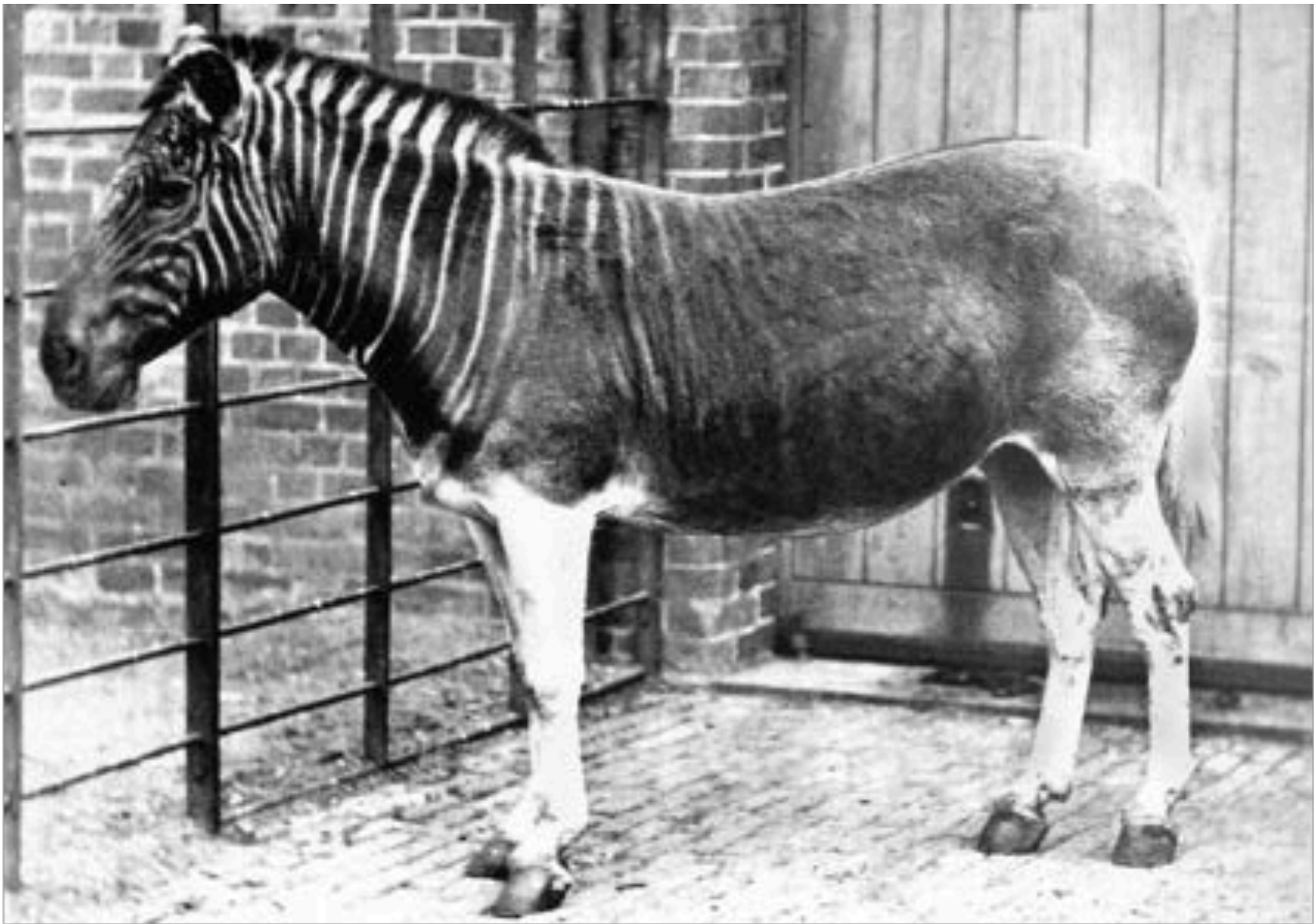
## DNA sequences from the quagga, an extinct member of the horse family

Russell Higuchi\*, Barbara Bowman\*, Mary Freiburger\*,  
Oliver A. Ryder† & Allan C. Wilson\*

\* Department of Biochemistry, University of California, Berkeley,  
California 94720, USA

† Research Department, San Diego Zoo, San Diego,  
California 92103, USA

To determine whether DNA survives and can be recovered from the remains of extinct creatures, we have examined dried muscle from a museum specimen of the quagga, a zebra-like species (*Equus quagga*) that became extinct in 1883 (ref. 1). We report that DNA



### Unidentified reading frame 1

Quagga	C CCA ATC CTG CTC GCC GTA GCA TTC CTC ACA CTA GTT GAA CGA AAA GTC TTA GGC TAC ATA CAA CTT CGT AAA GGA CCC AAC ATC GTA GGC CCC TAT GGC CTA CTA CAA CCC ATT AC
Zebra	. . . . . T . . . . . G . . . . . T . . . . . C . . . . . G*

### Cytochrome oxidase I

Quagga	A GGA GGA TTC GTT CAC TGA TTC CCT CTA TTC TCA GGA TAC ACA CTC AAC CAA ACC TGA GCA AAA ATT CAC TTT ACA ATT ATA TTC GTA GGG GTC AAC ATA ATT TTC TTC CCA
Zebra	G . . . . . T . . . . . G . . . . . C . . . . . A . . . . . T . . . . . C*

**Fig. 1** Sequences of the coding strands determined for two pieces of quagga mtDNA. The sequences are arranged in triplets corresponding to the amino acids that they encode. At 12 positions, the quagga sequences differ from those of mtDNA from a mountain zebra; only for these positions is the nature of the base specified for the zebra. The two asterisks identify triplets at which the zebra and quagga differ by an amino acid replacement.



# Nobel prize in Medicine 2022

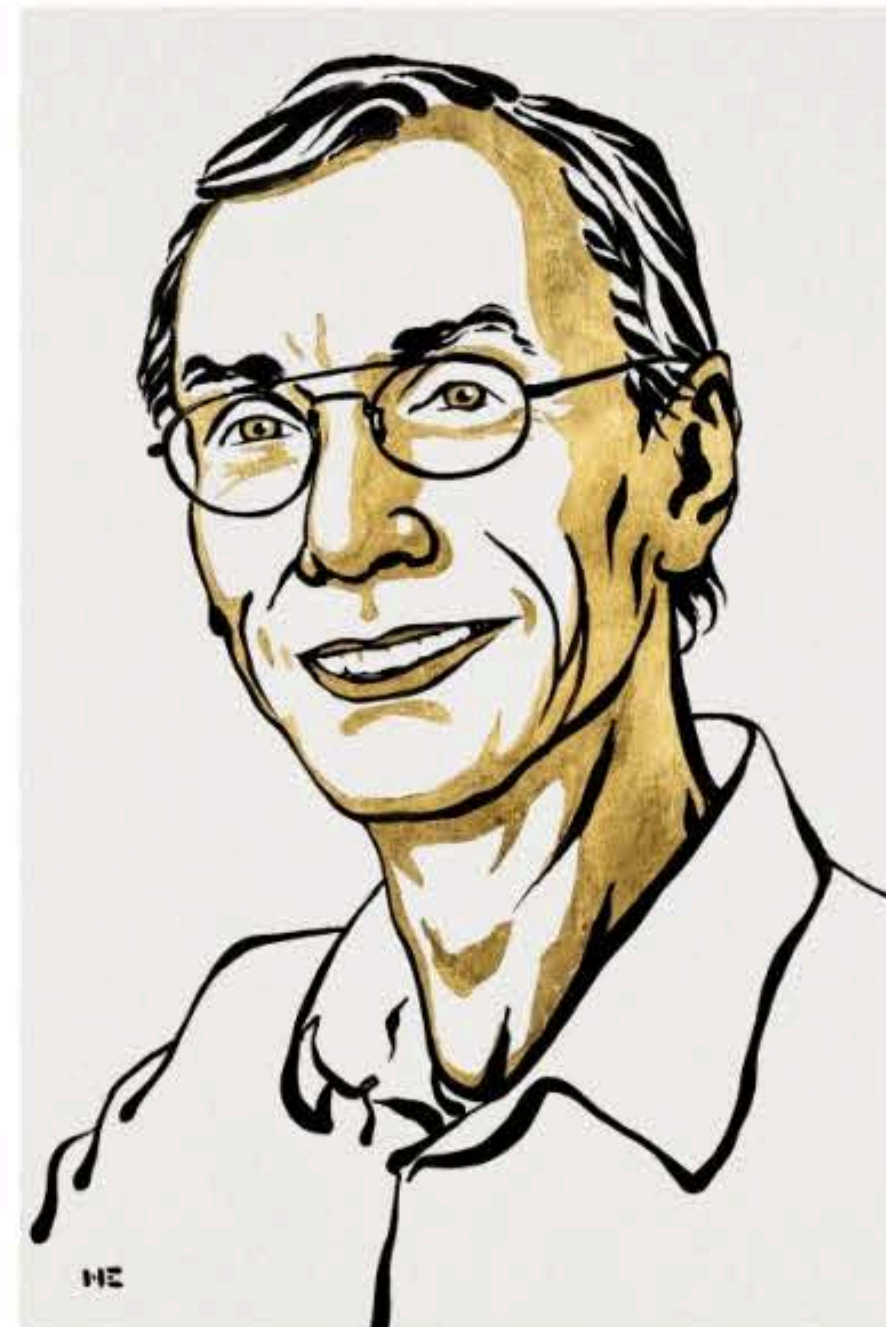
The Nobel Prize in Physiology or  
Medicine 2022

Svante Pääbo

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## Svante Pääbo Facts



Ill. Niklas Elmehed © Nobel  
Prize Outreach

Svante Pääbo

The Nobel Prize in Physiology or Medicine 2022

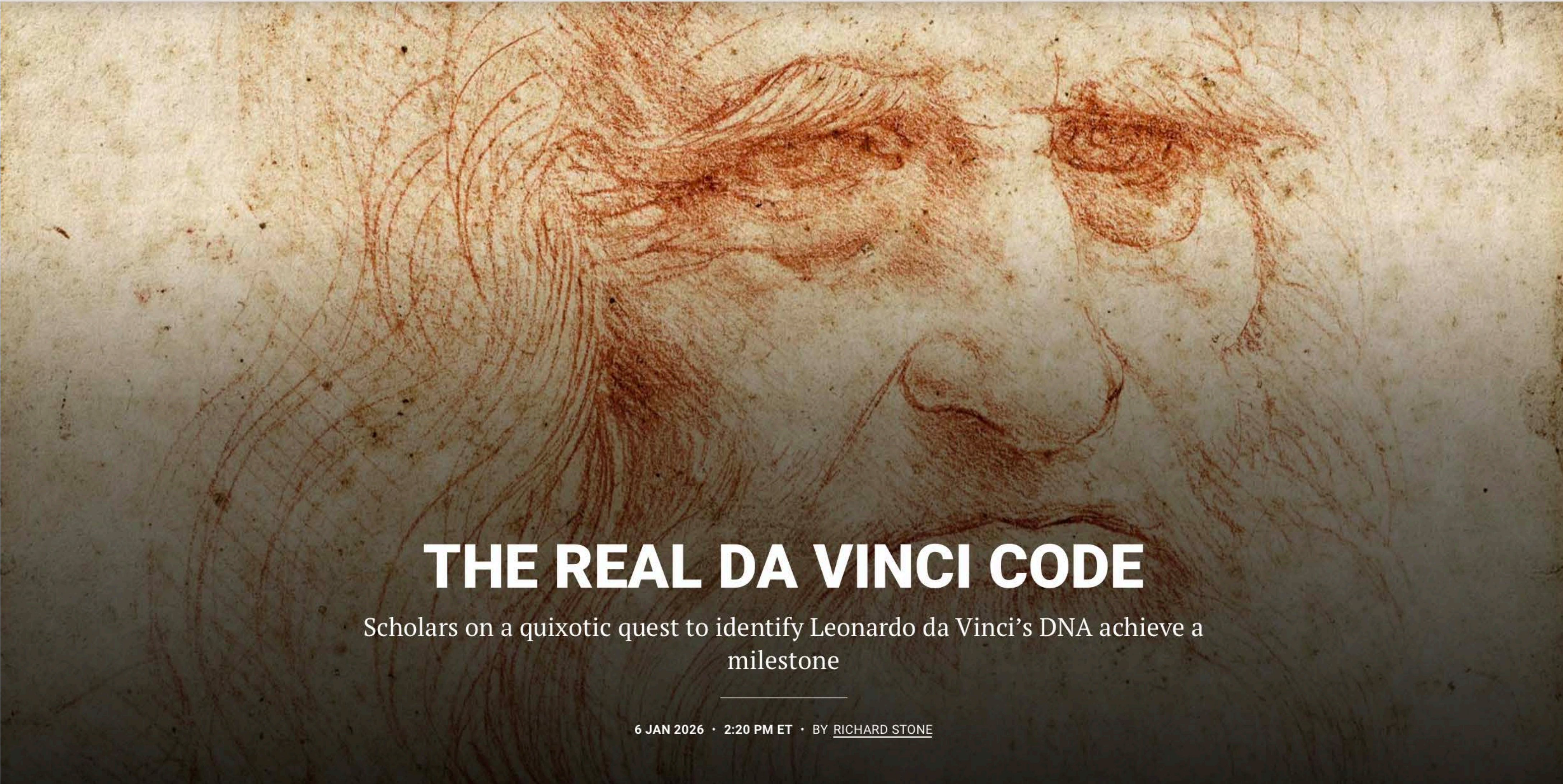
Born: 20 April 1955, Stockholm, Sweden

Affiliation at the time of the award: Max Planck Institute for  
Evolutionary Anthropology, Leipzig, Germany, Okinawa  
Institute of Science and Technology, Okinawa, Japan

Prize motivation: “for his discoveries concerning the  
genomes of extinct hominins and human evolution”

Prize share: 1/1





## THE REAL DA VINCI CODE

Scholars on a quixotic quest to identify Leonardo da Vinci's DNA achieve a milestone

6 JAN 2026 · 2:20 PM ET · BY [RICHARD STONE](#)



# The week in ancient DNA

New Results

Follow this preprint

## Biological signatures of history: Examination of composite biomes and Y chromosome analysis from da Vinci-associated cultural artifacts

Harinder Singh, Seesandra V. Rajagopala, Rebecca Hart, Pille Hallast, Mark Loftus, Rosana Wiscovitch-Russo, Cody R. K. Conrad, David S. Thaler, Guadalupe Piñar, Karina C. Åberg, Rossella Lorenzi, José A Lorente, Jesse H. Ausubel, Thomas P. Sakmar, Rhonda K. Roby, Charles Lee, Norberto Gonzalez-Juarbe

doi: <https://doi.org/10.64898/2026.01.06.697880>

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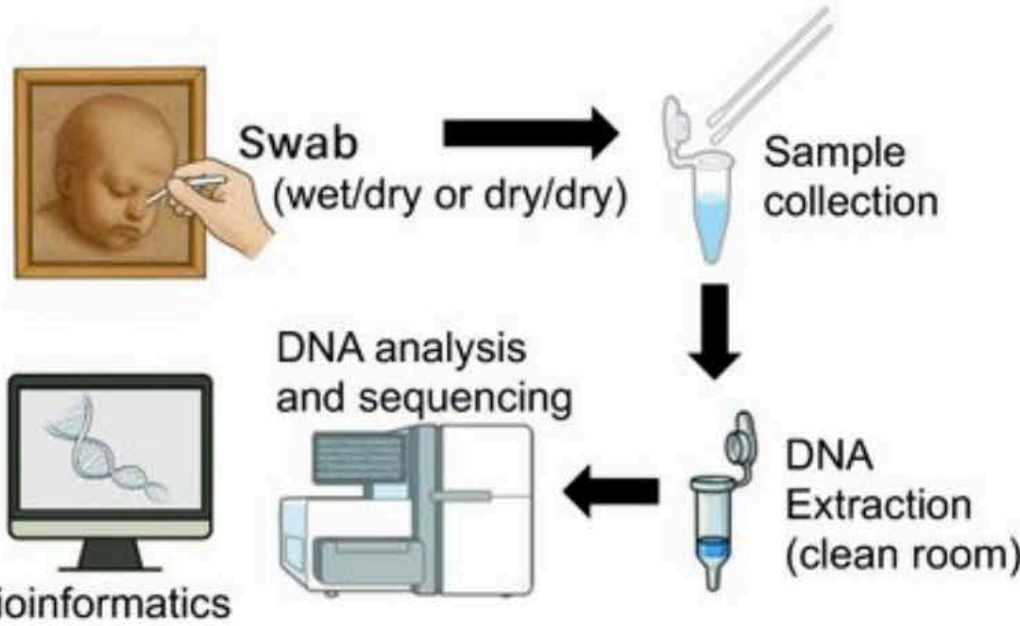


A.

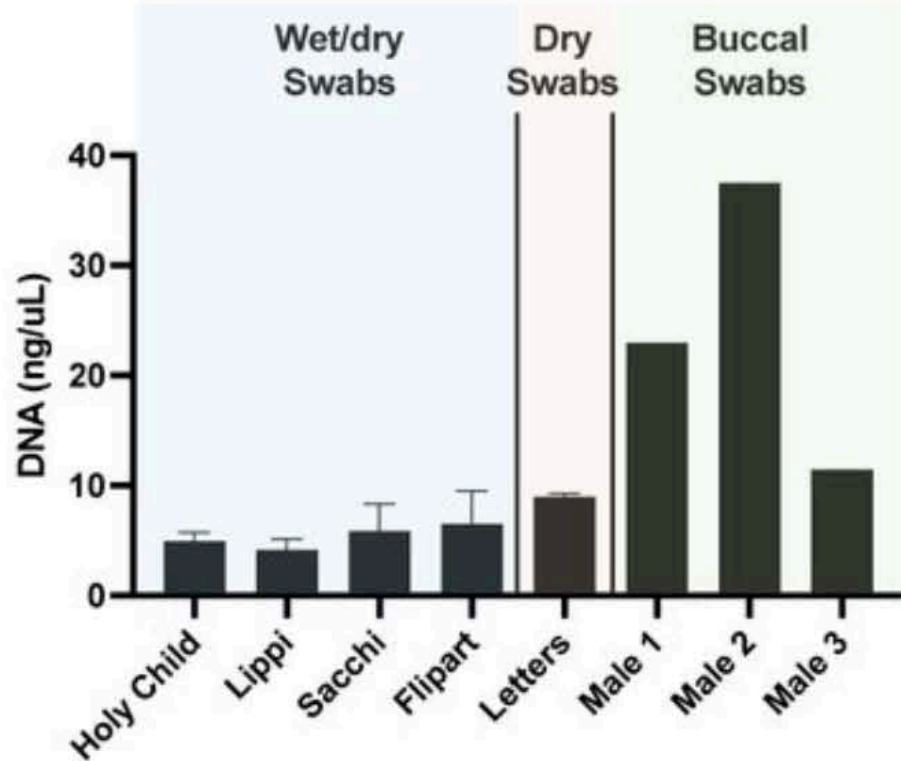


Title or unique ID:	Holy Child	Letters	St. John in a Classical Robe by a Pillar	Man Caught in the Wind	Seated Gentleman at Writing Table
Style:	Red chalk	Handwritten letter	Silver point and ink with blue wash	Red chalk	Red chalk
Paper:	Laid paper	Linen-rag paper	Laid paper	Laid paper	Laid paper
Measurement:	133 x 102 mm	not taken	133 x 102 mm	296 x 212 mm	260 x 172 mm
Sampling Technique:	Double swab (wet/dry)	Dry swab	Double swab (wet/dry)	Double swab (wet/dry)	Double swab (wet/dry)
Country:	Italy	Spain/Italy	Italy	Italy	France
Date or period:	c. 1470 CE	c. 1392-1403 CE	c.1501 CE	c. 1625 CE	c. 1740 CE
Maker:	Leonardo Da Vinci (1452-1519)	Frosino di Ser Giovanni da Vinci (1400s)	Filippino Lippi (1457-1504)	Andrea Sacchi (1599-1661)	Charles J. Flipart (1721-1796)

B.



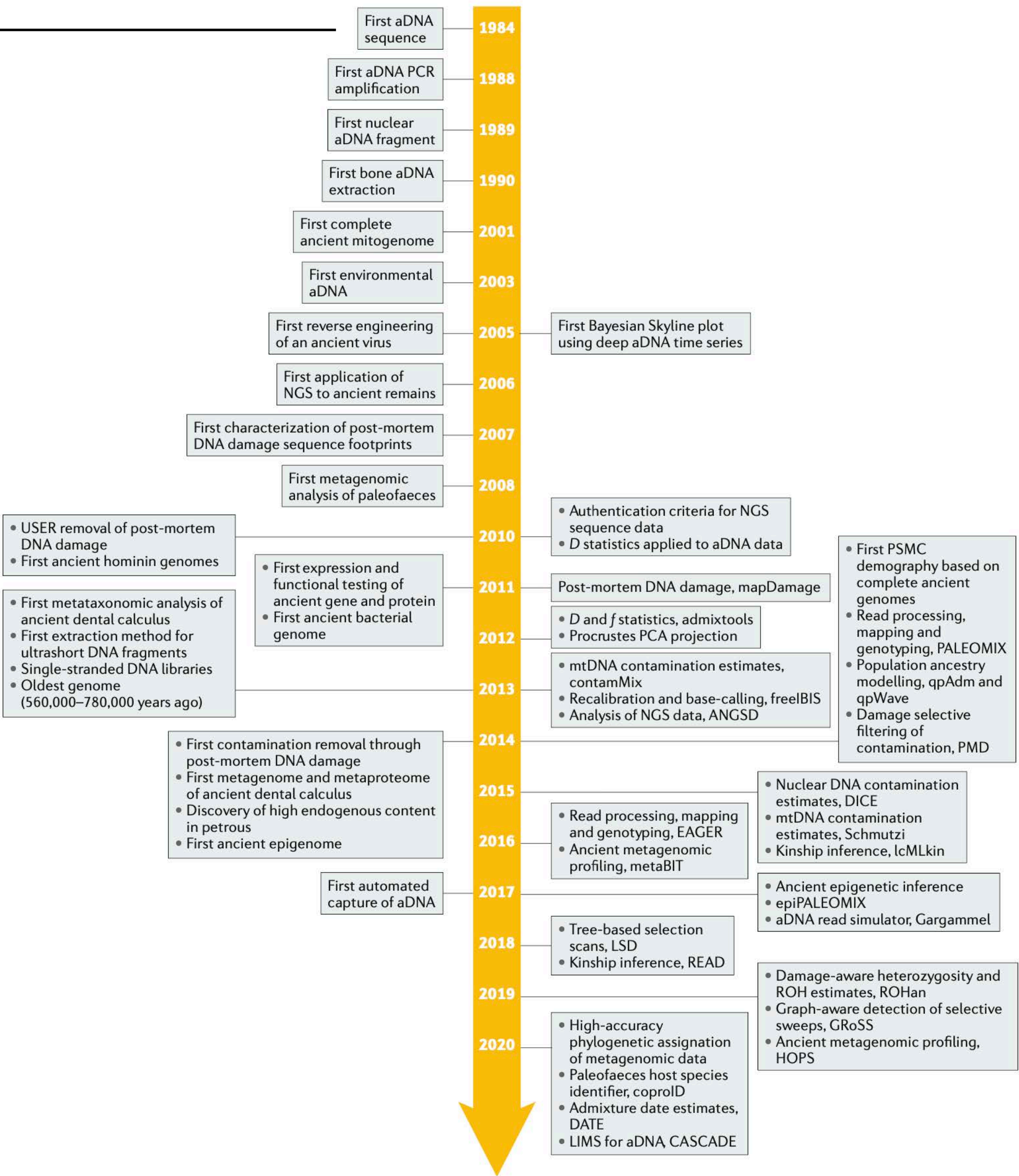
C.





# Milestones in ancient DNA

1984 - First aDNA sequence

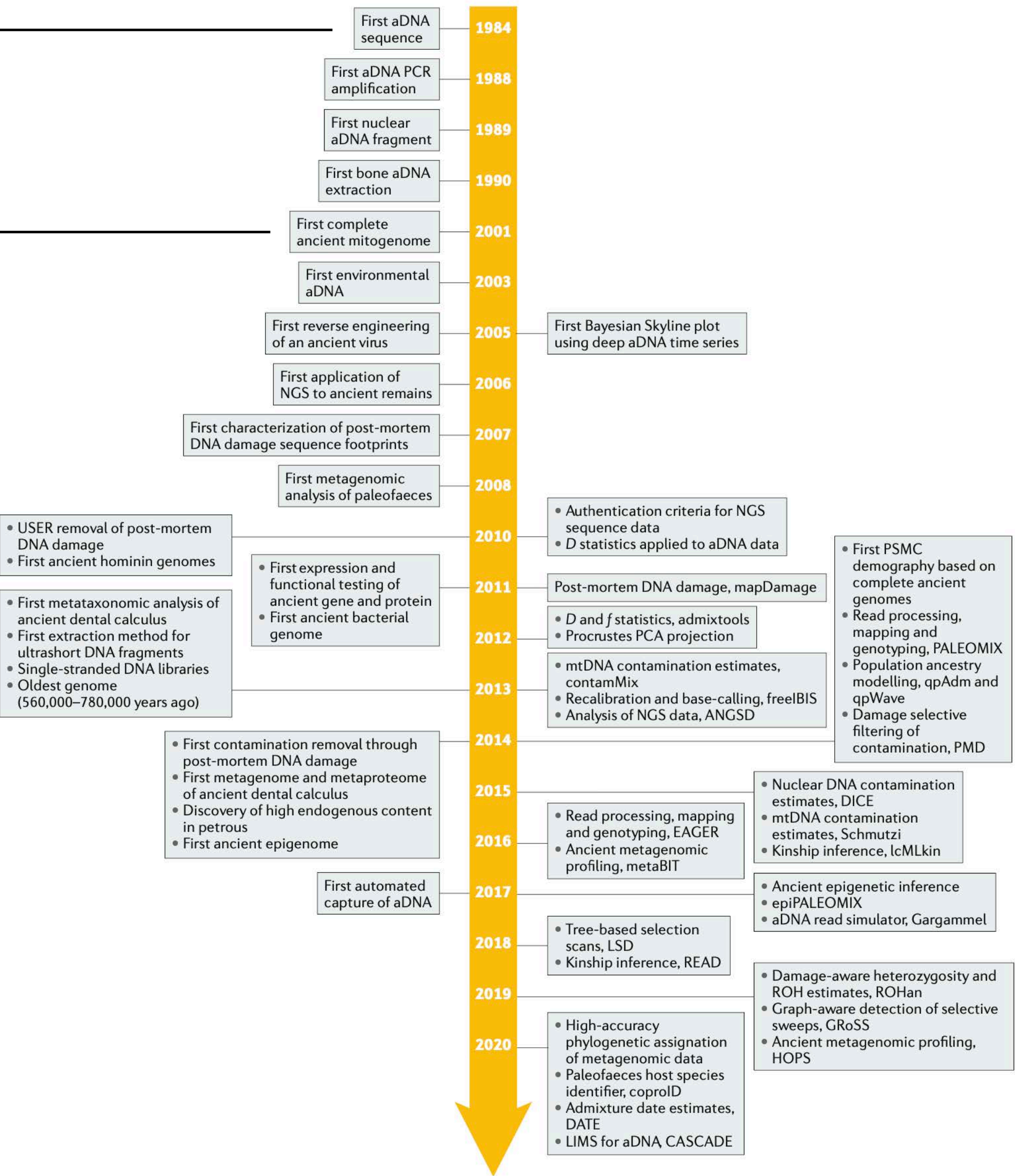




# Milestones in ancient DNA

1984 - First aDNA sequence

2001 - First ancient mitogenome



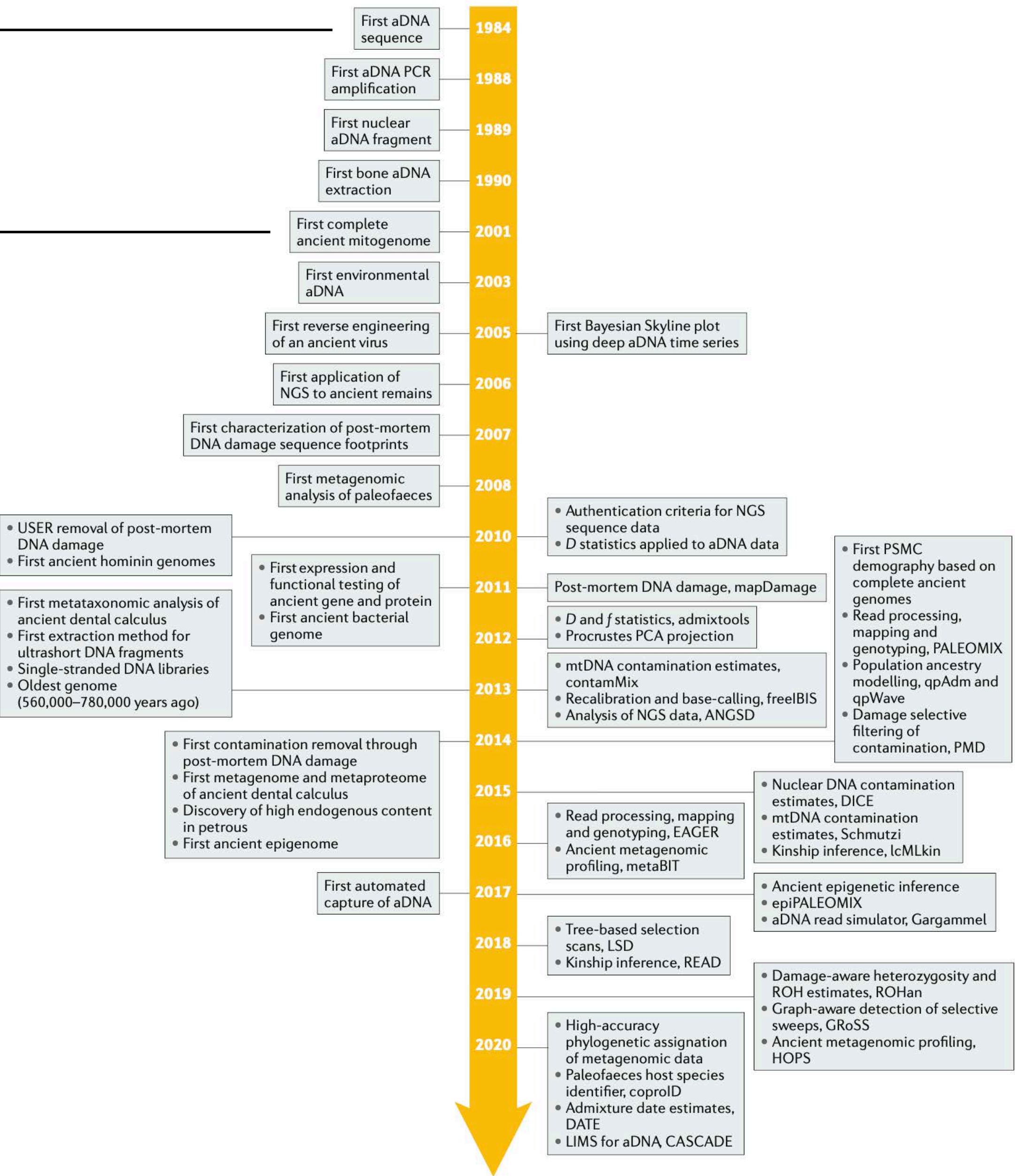


# Milestones in ancient DNA

1984 - First aDNA sequence

2001 - First ancient mitogenome

2010 - First ancient human genome





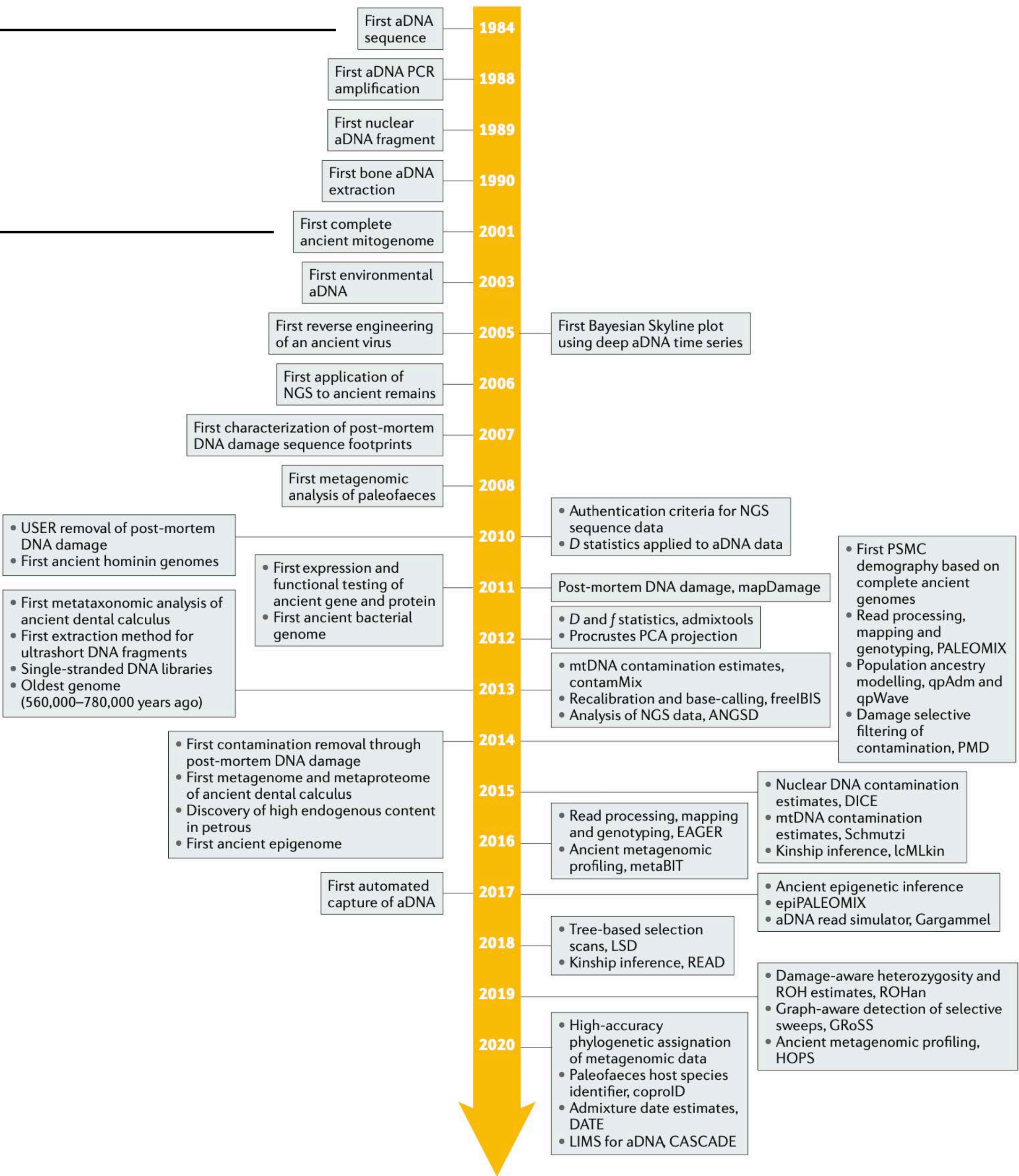
# Milestones in ancient DNA

1984 - First aDNA sequence

2001 - First ancient mitogenome

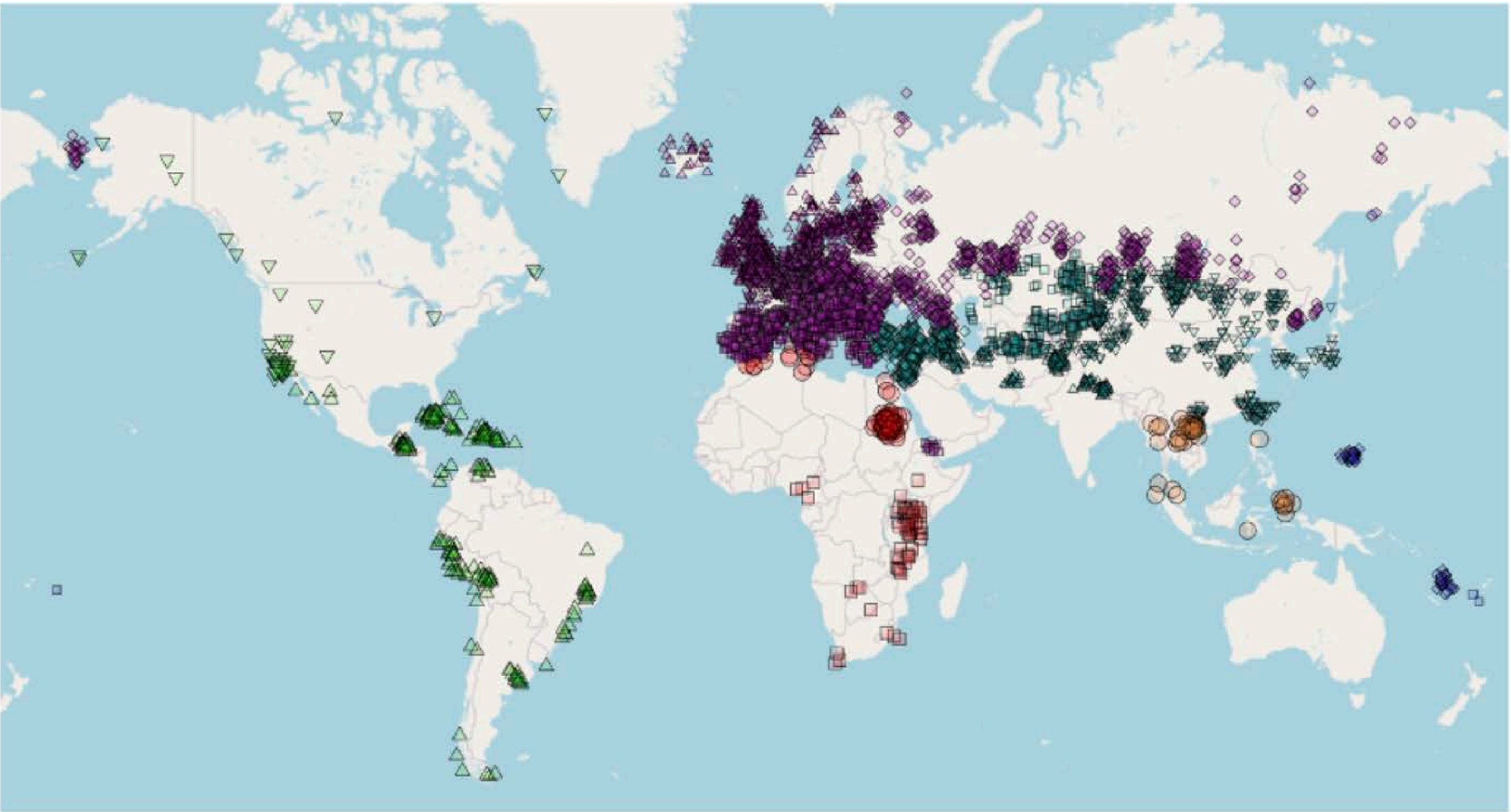
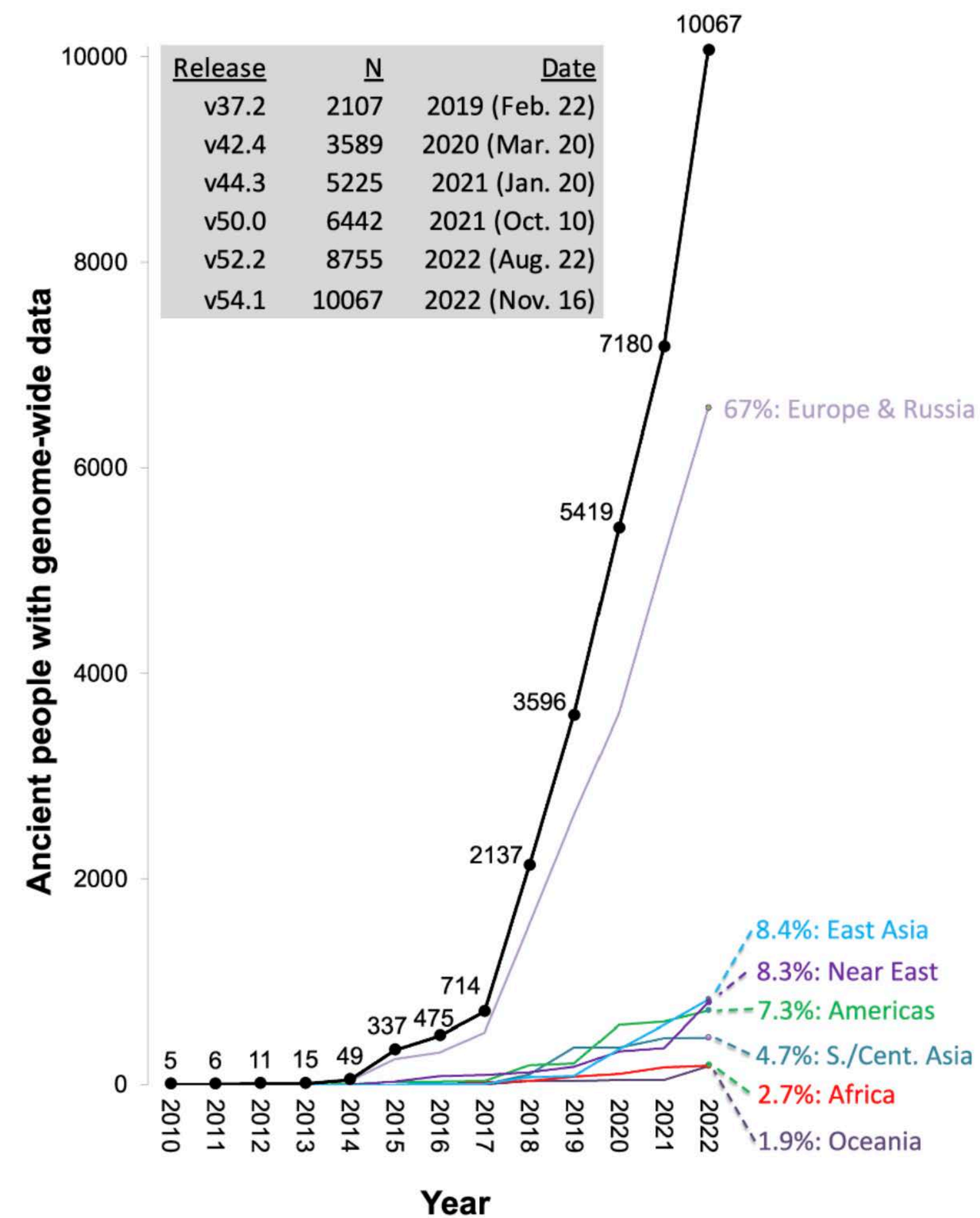
2010 - First ancient human genome

The “big data” era of ancient genomics





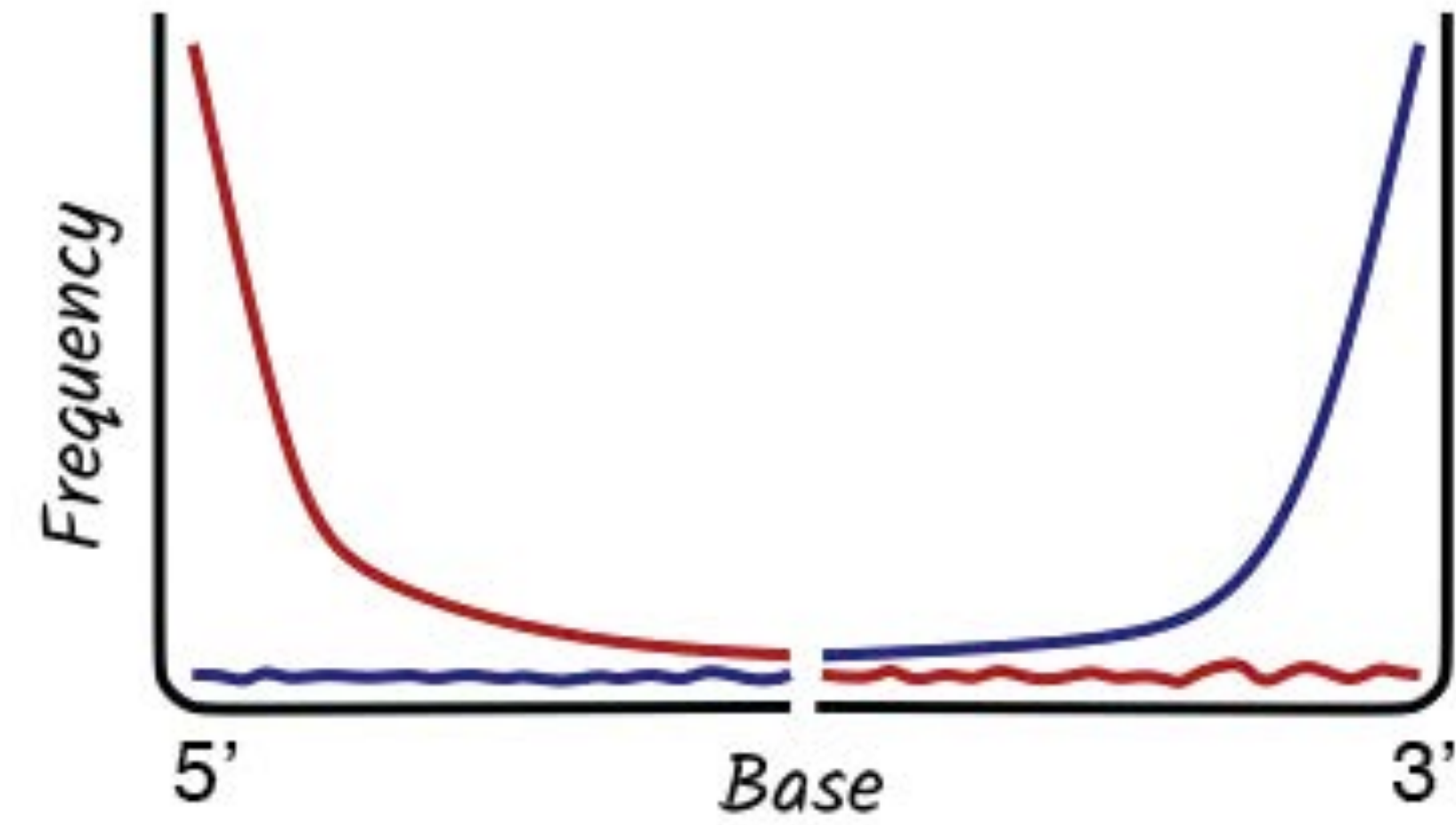
# The ancient human DNA revolution



Over 10,000 ancient humans with genome-wide data since 2010



# Topics for today's lecture

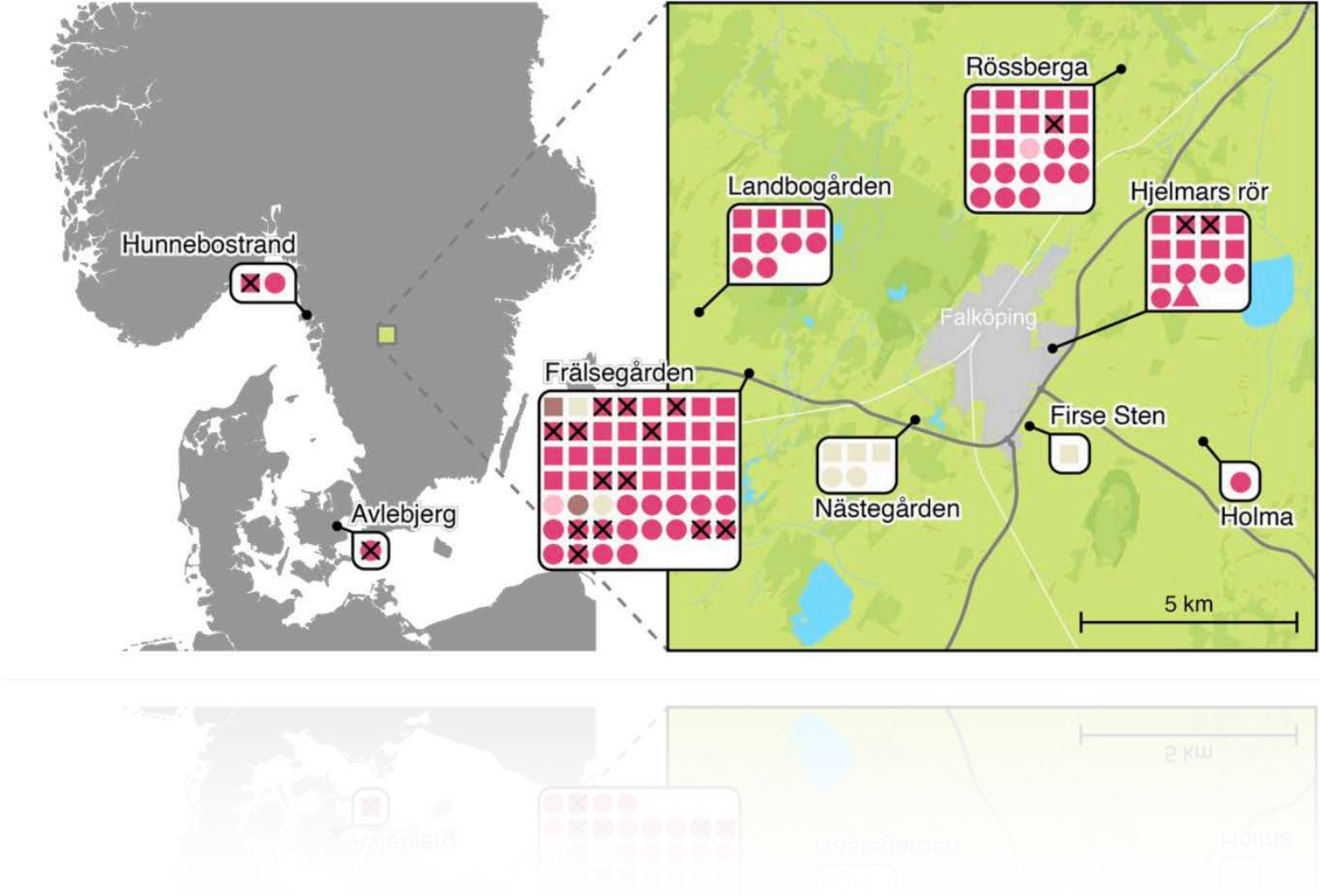
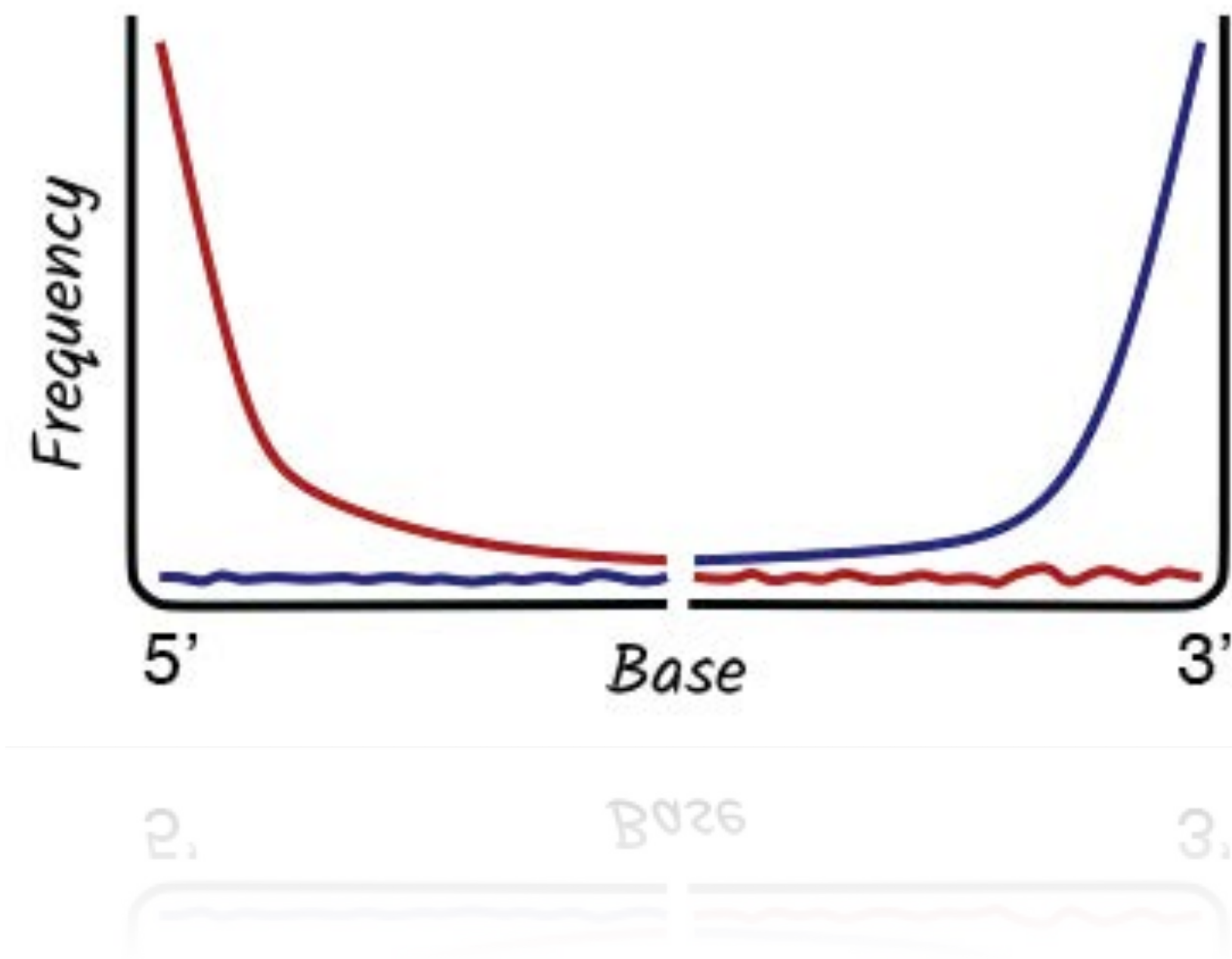


## Ancient DNA primer

Characteristics and challenges



# Topics for today's lecture

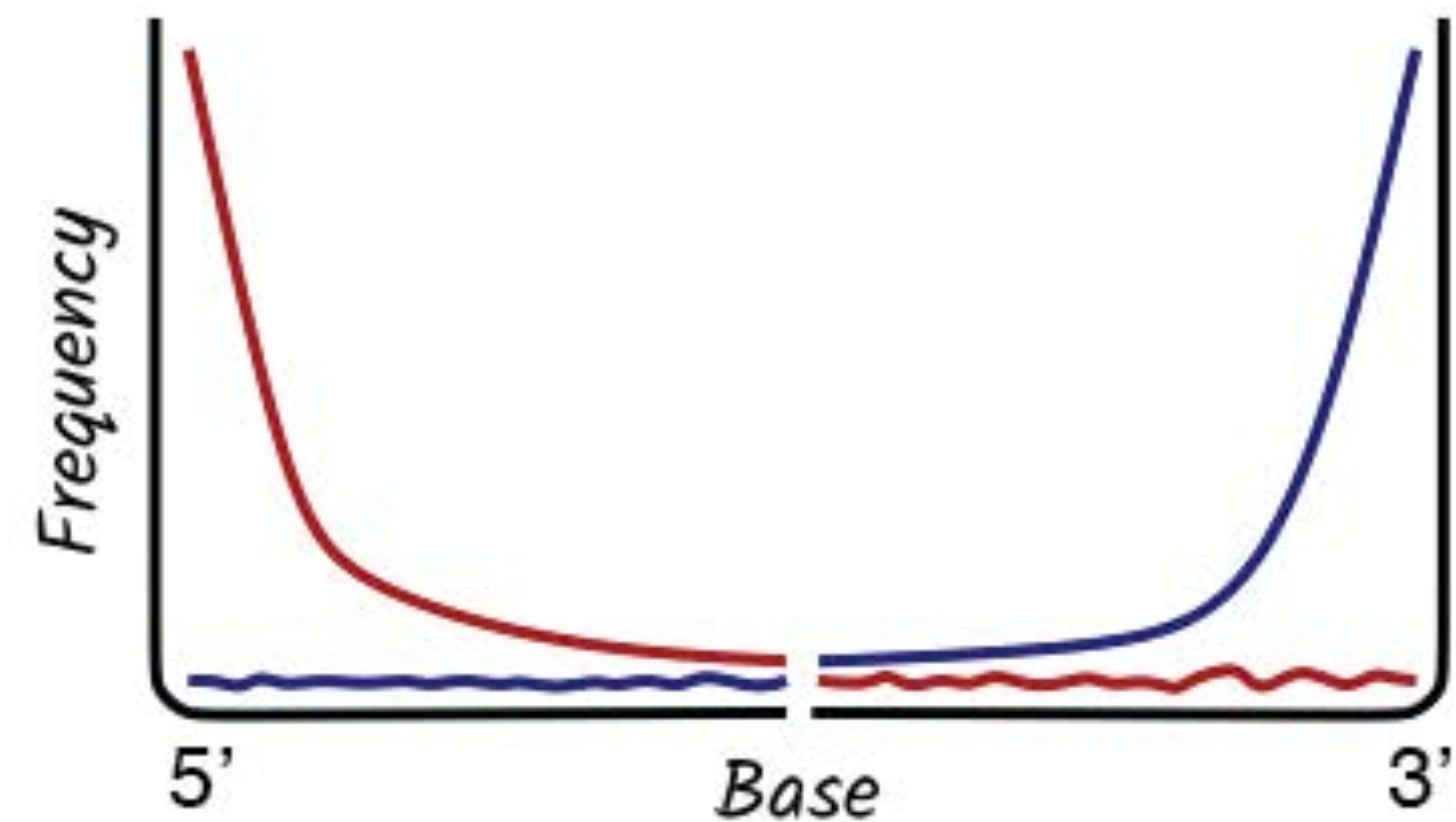


**Ancient DNA primer**  
Characteristics and challenges

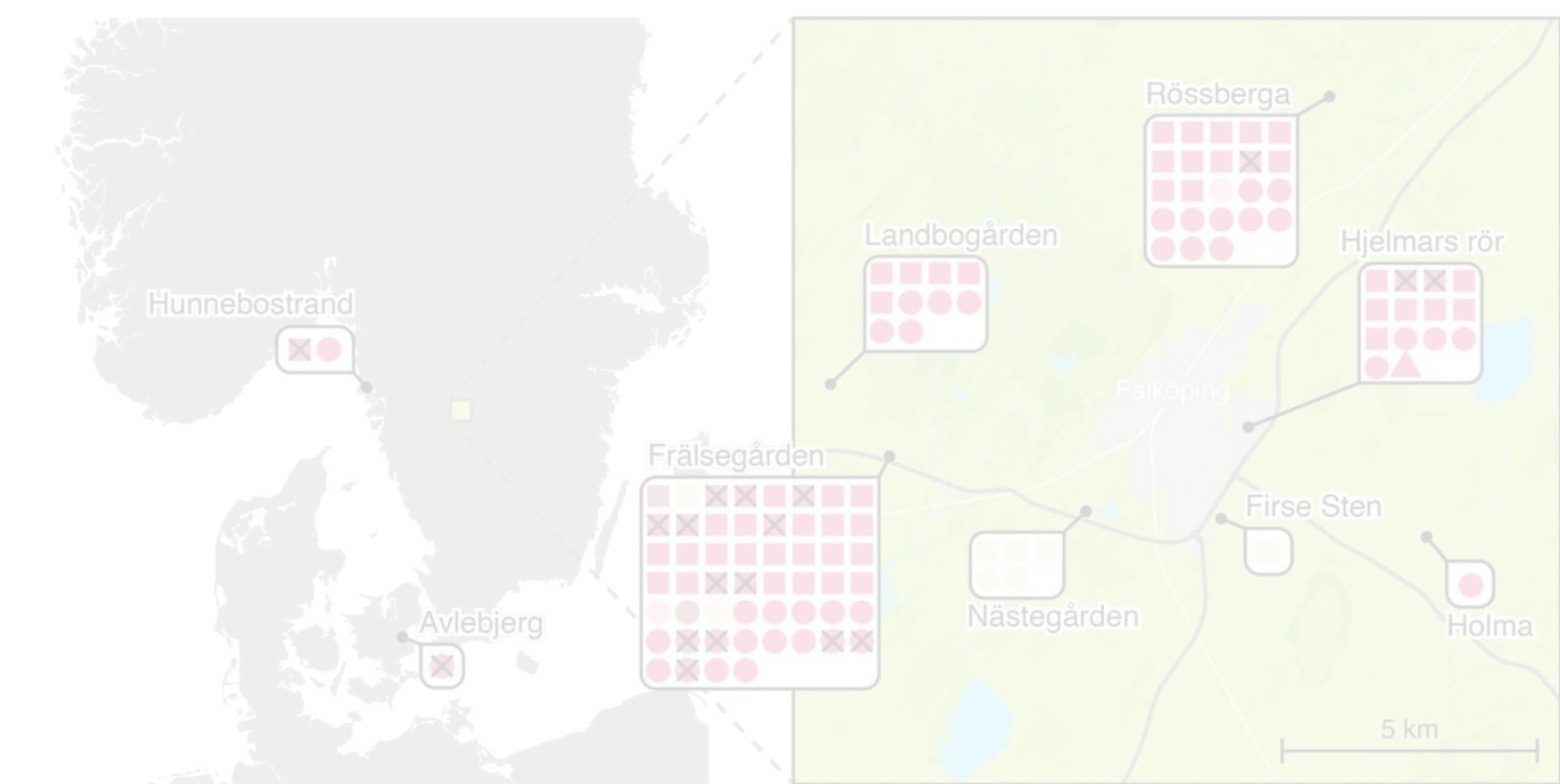
**Ancient DNA showcase**  
Kinship and plague in Stone Age Scandinavia



# Topics for today's lecture



**Ancient DNA primer**  
Characteristics and challenges



**Ancient DNA showcase**  
Kinship and plague in Stone Age Scandinavia



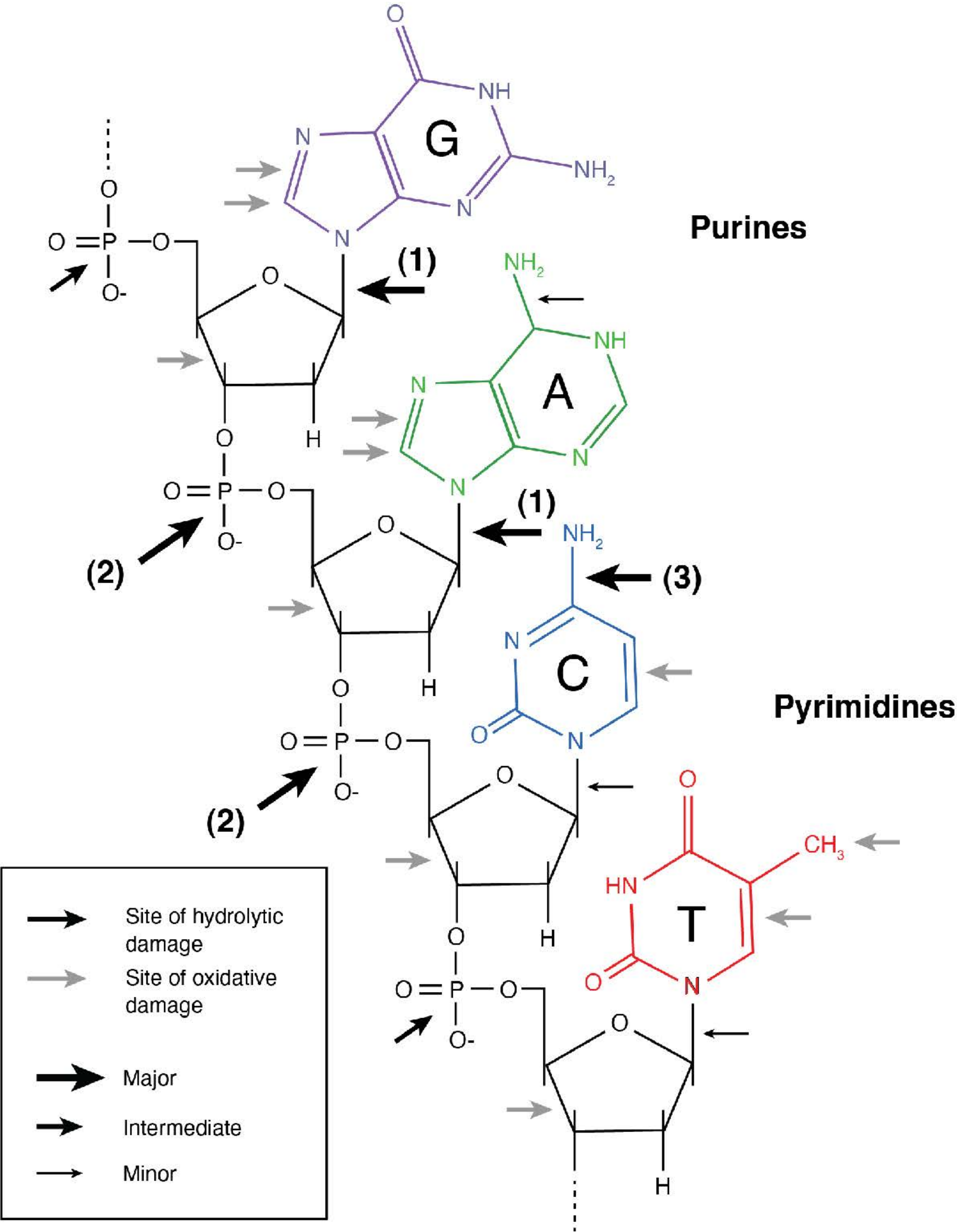
# Characteristics of ancient DNA



5,000 year-old human remains from the Provadia salt mine, Bulgaria

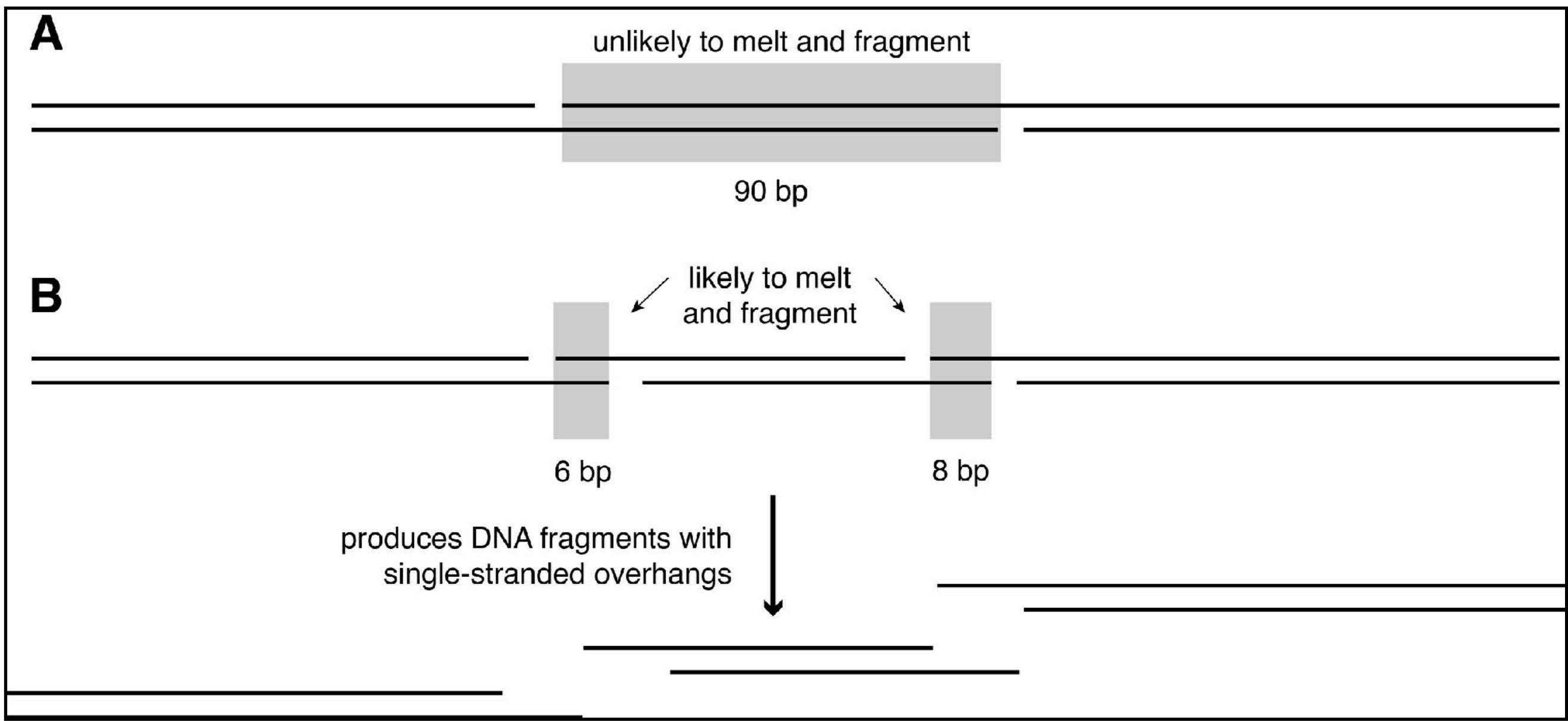
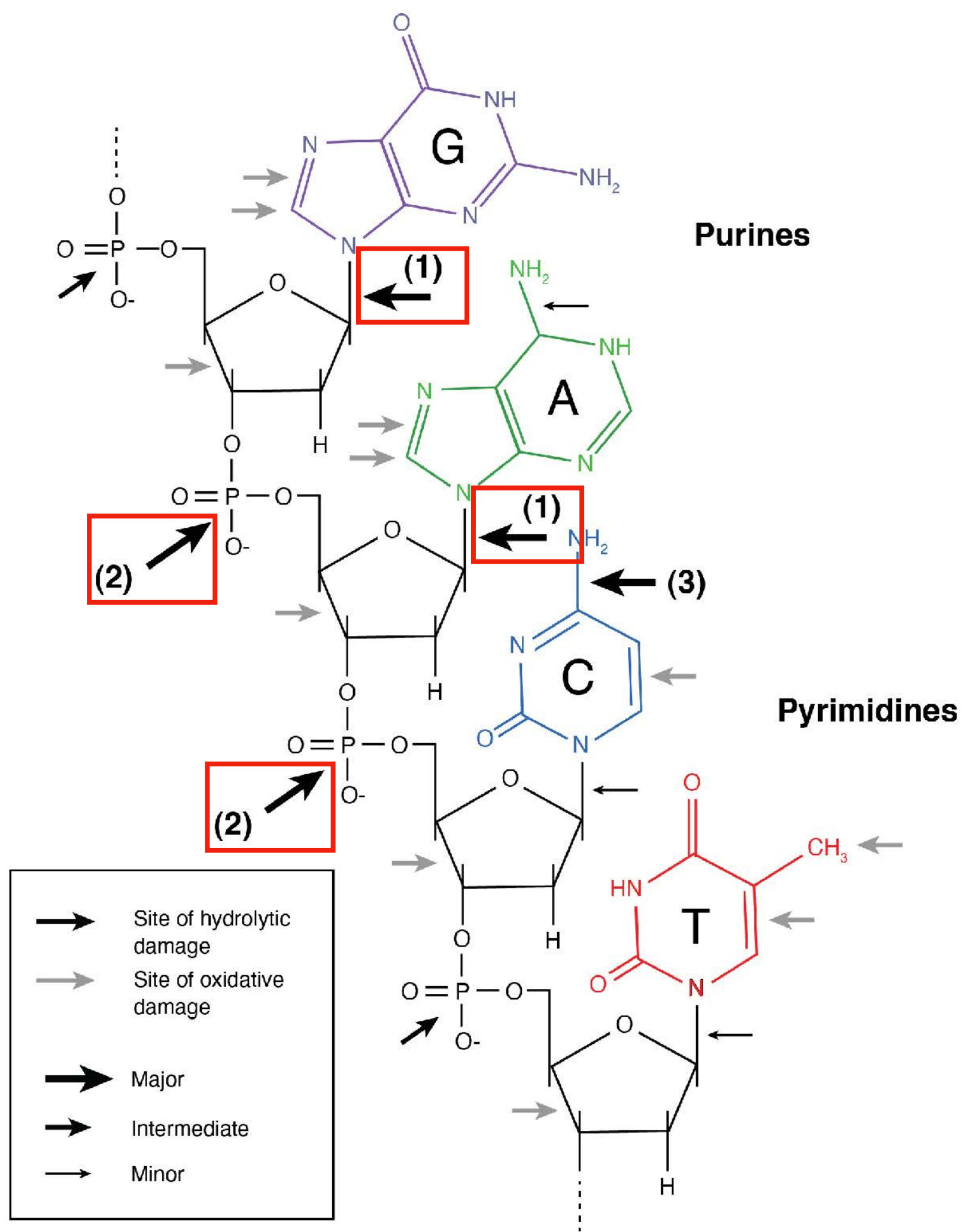


# How does DNA degrade?





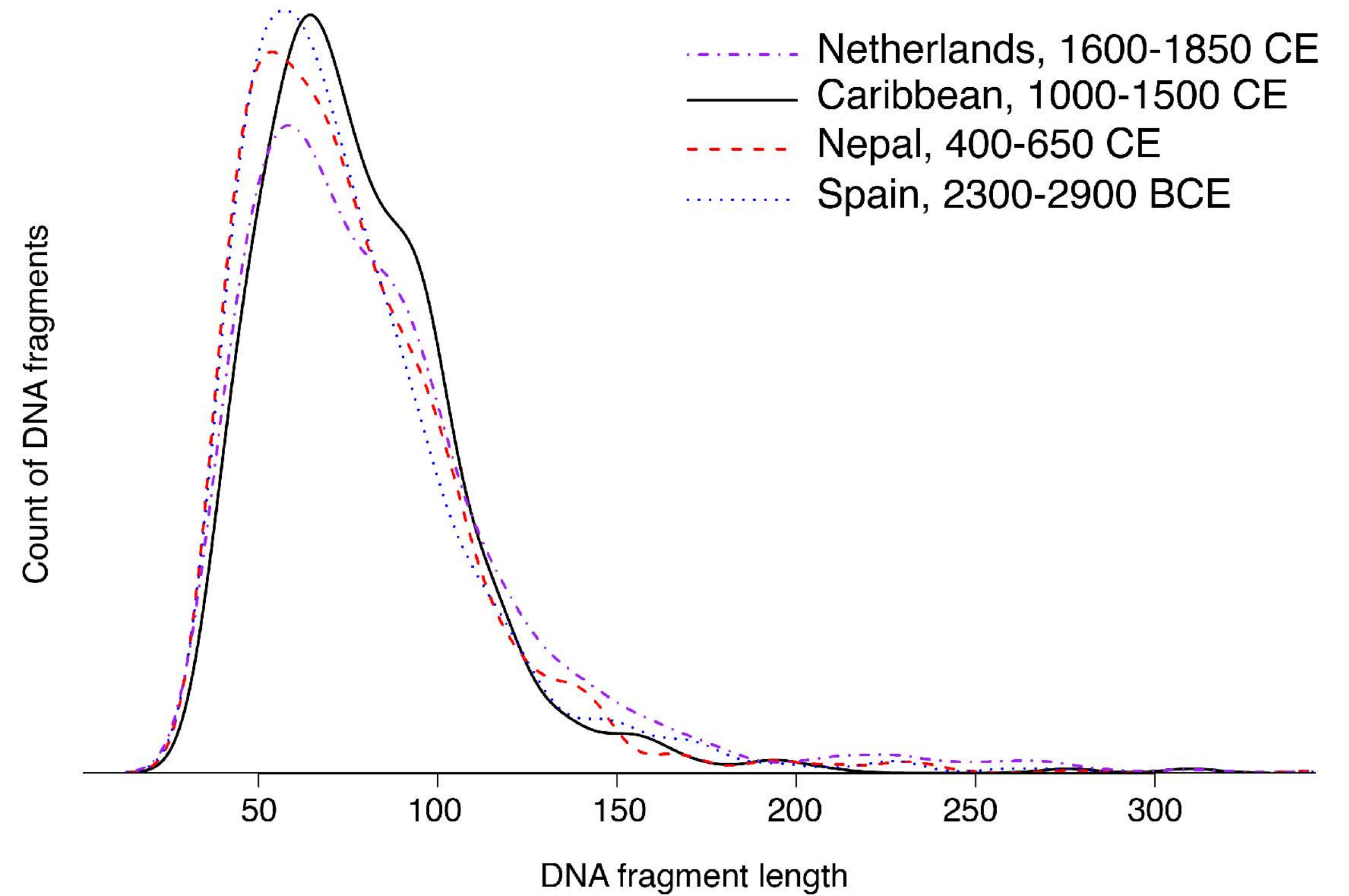
# Fragmentation of DNA



Depurination and nicking results in DNA fragmentation



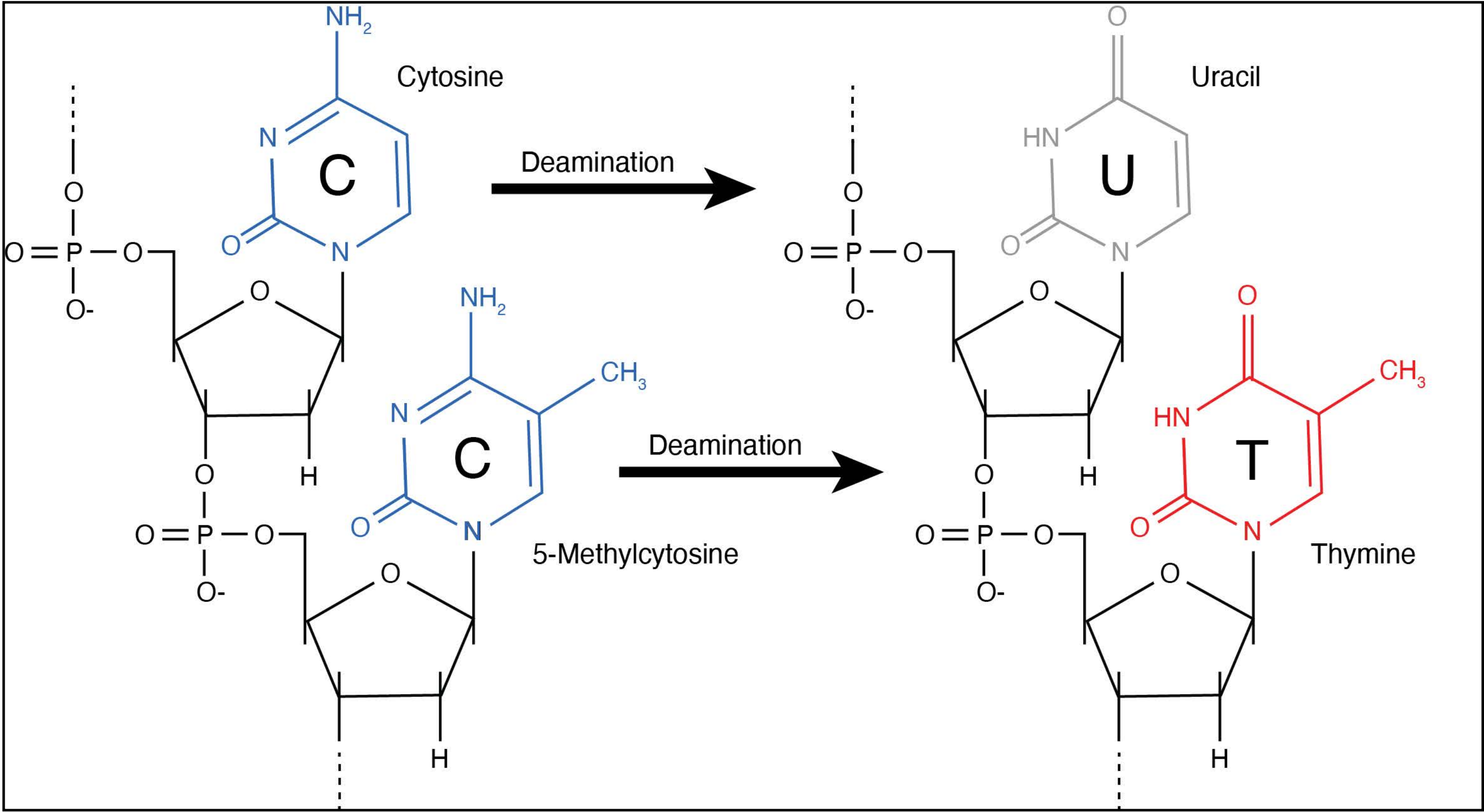
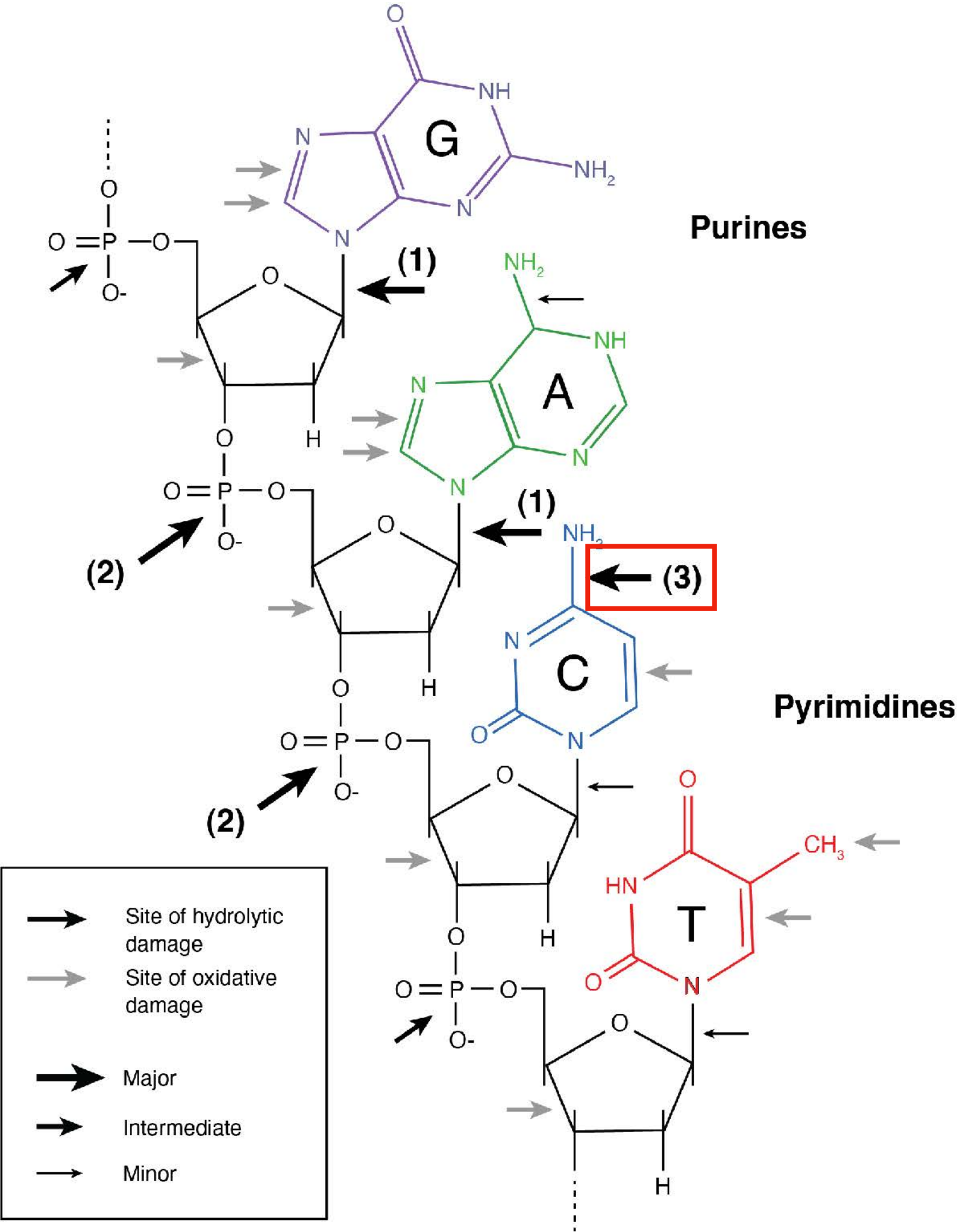
# Characteristics of ancient DNA



Ancient DNA is short and fragmented



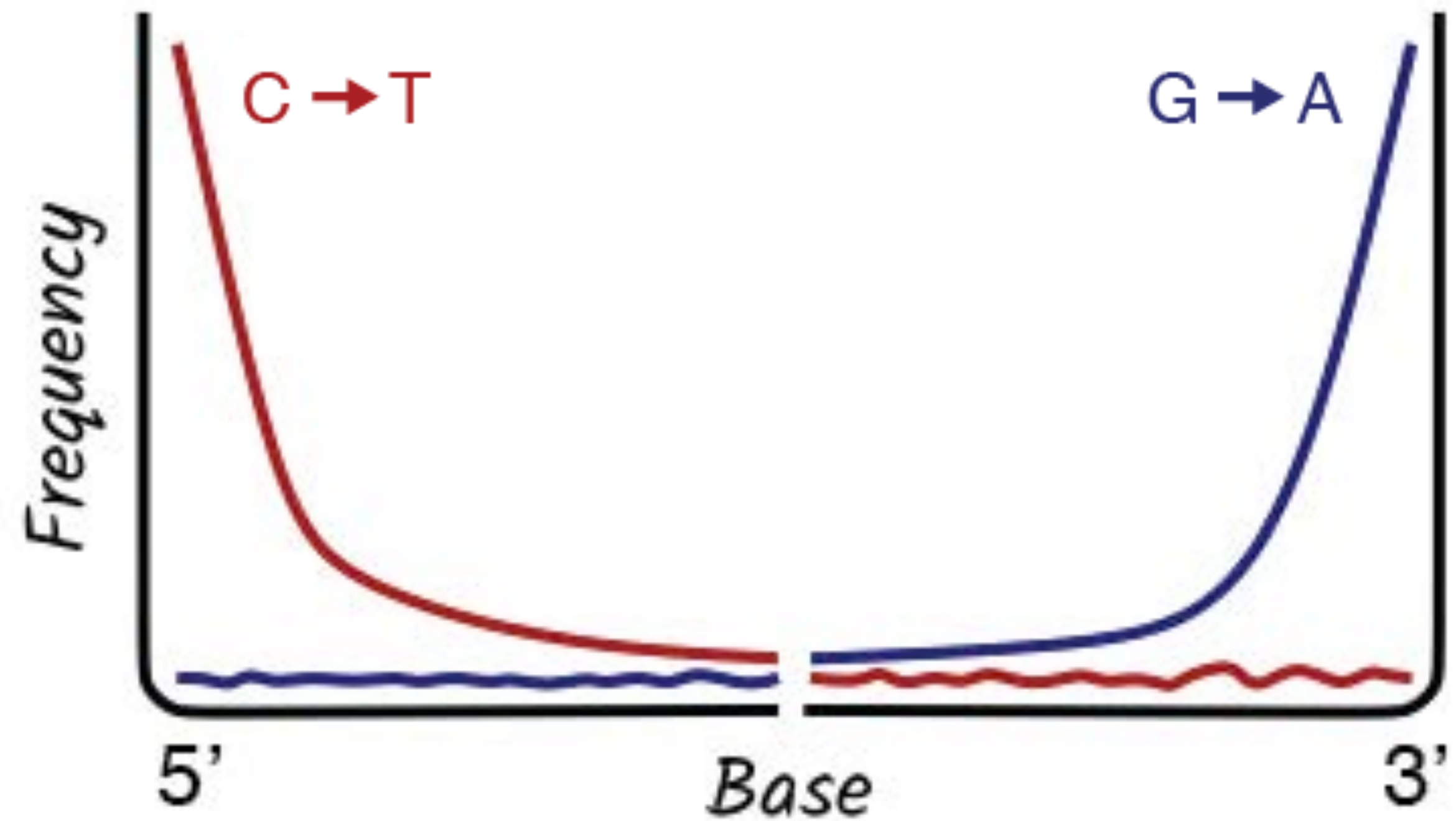
# Accumulation of DNA damage



Deamination results in base substitutions



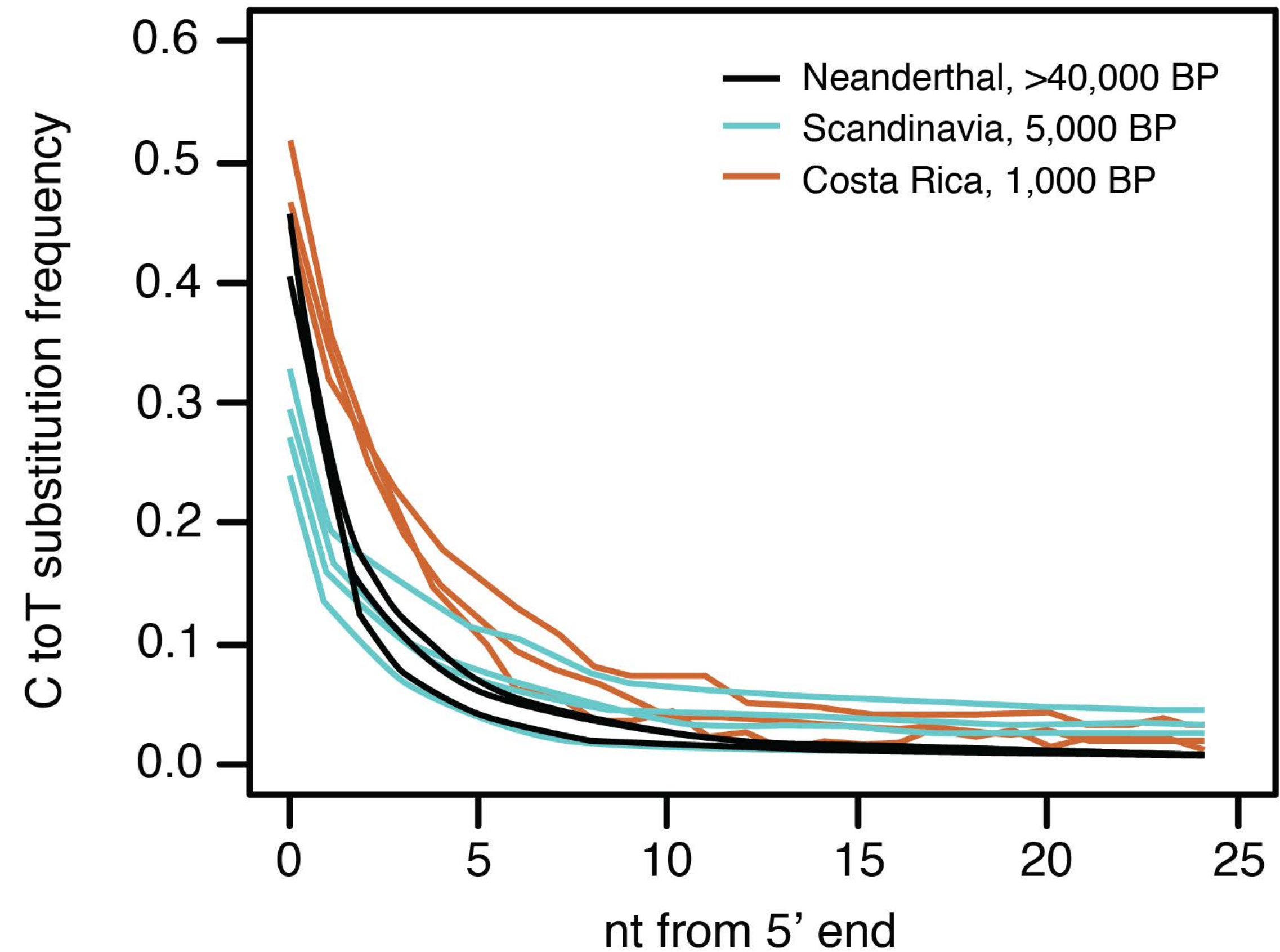
# Characteristics of ancient DNA



Ancient DNA is damaged with characteristic substitution patterns



# Characteristics of ancient DNA



Ancient DNA damage rates depend on microenvironmental conditions





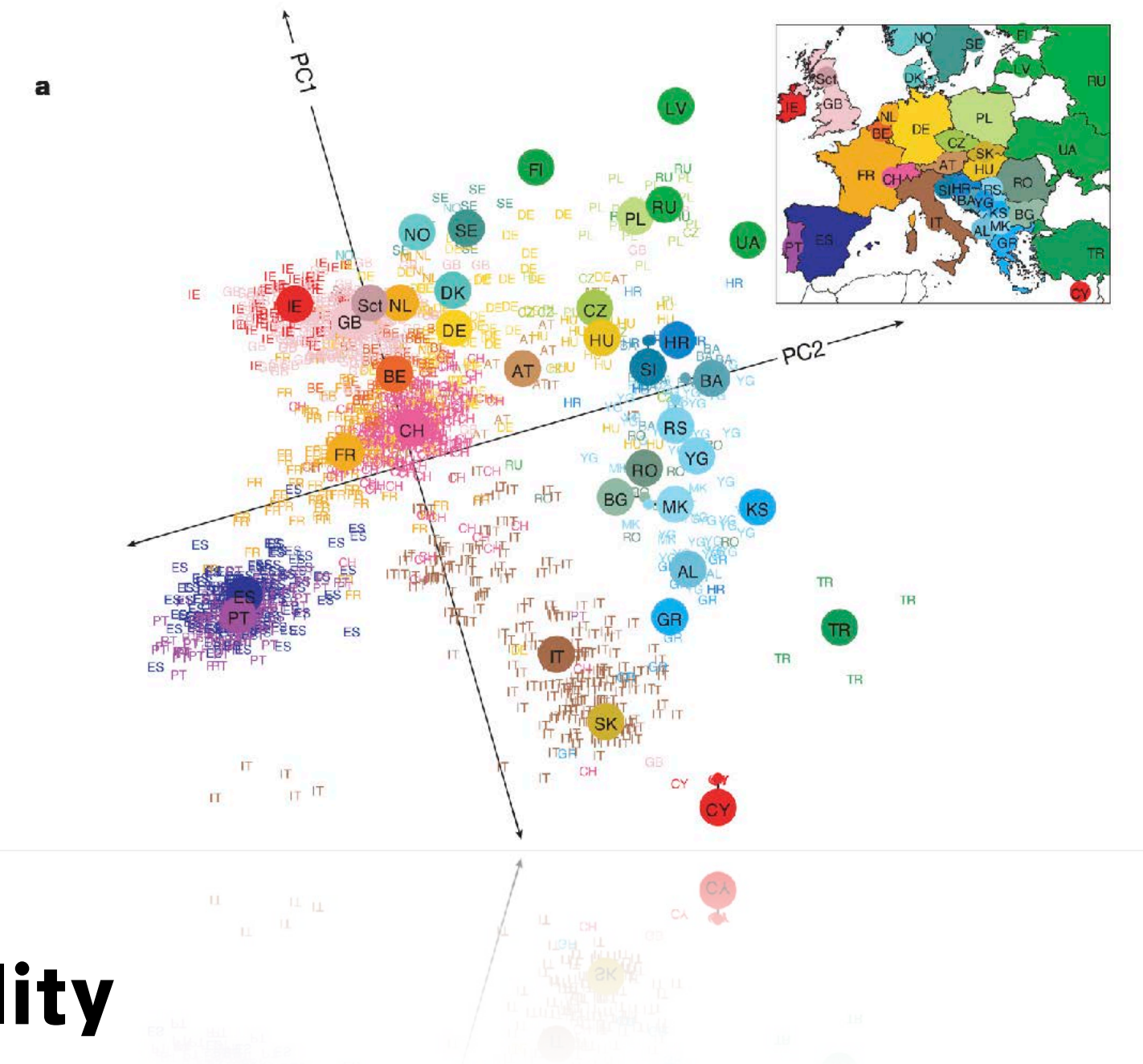
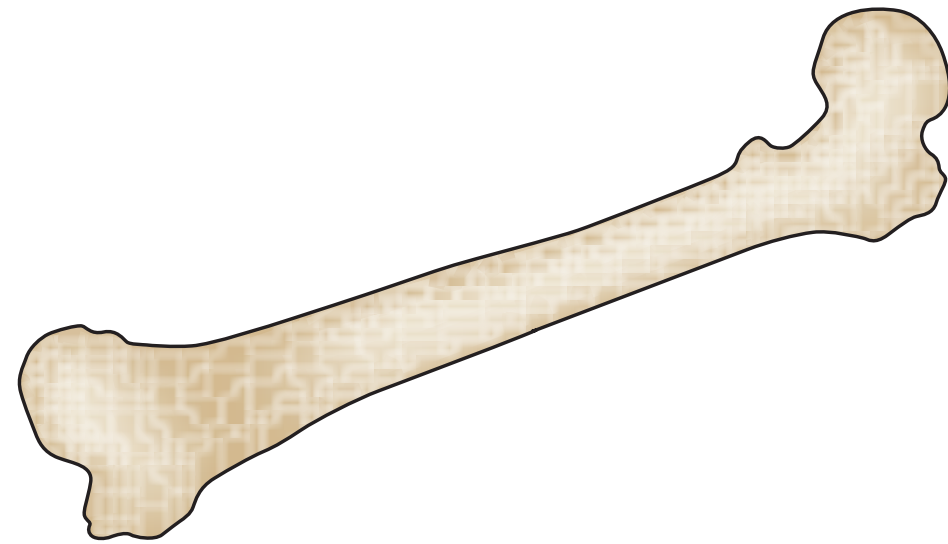


# Ancient DNA challenges

# Sample

## DNA sequences

## Analysis results



## DNA recovery

Preservation  
Sample material

## Data quality

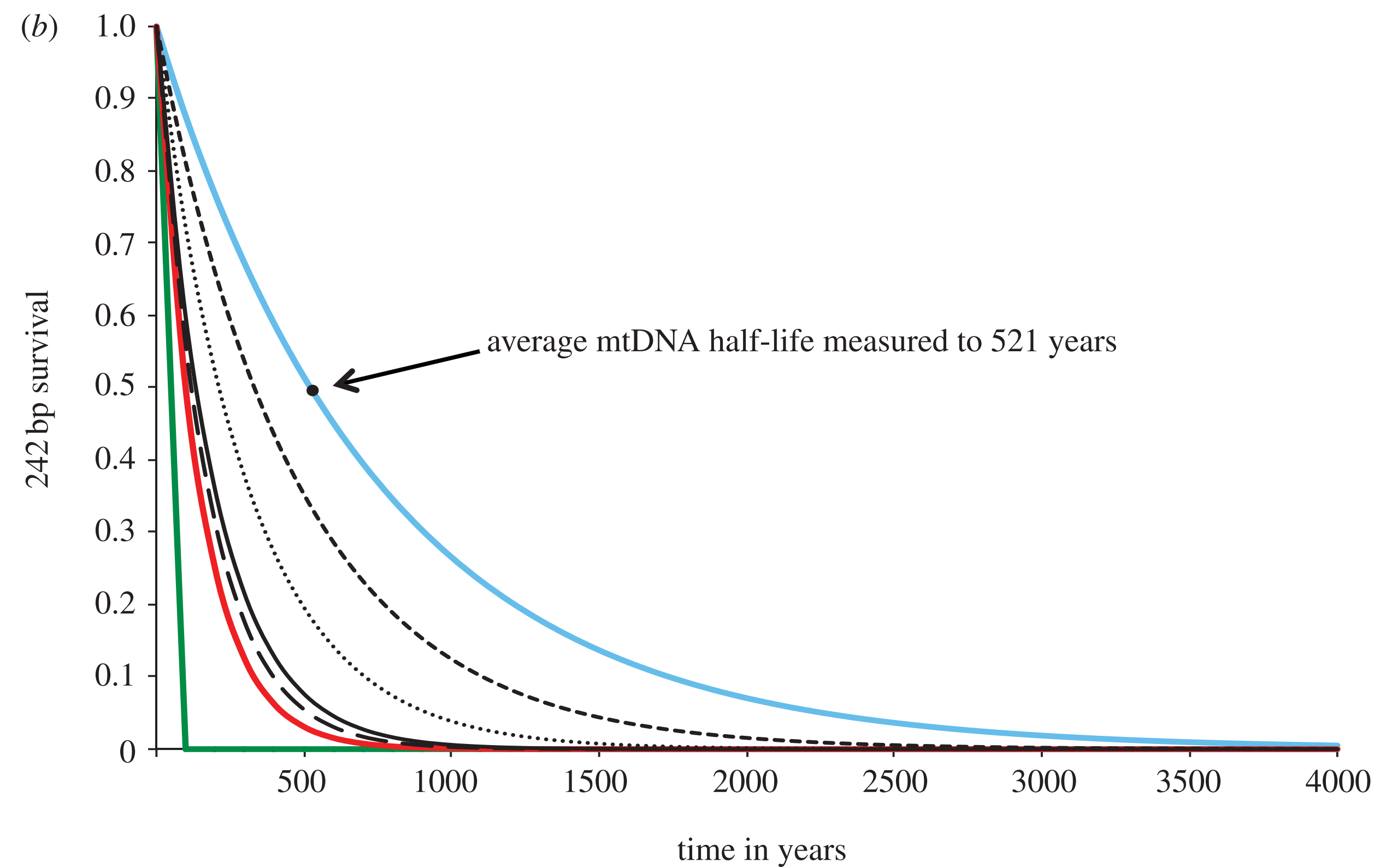
Contamination  
Genome coverage  
Error rates







# Challenges - Ancient DNA preservation

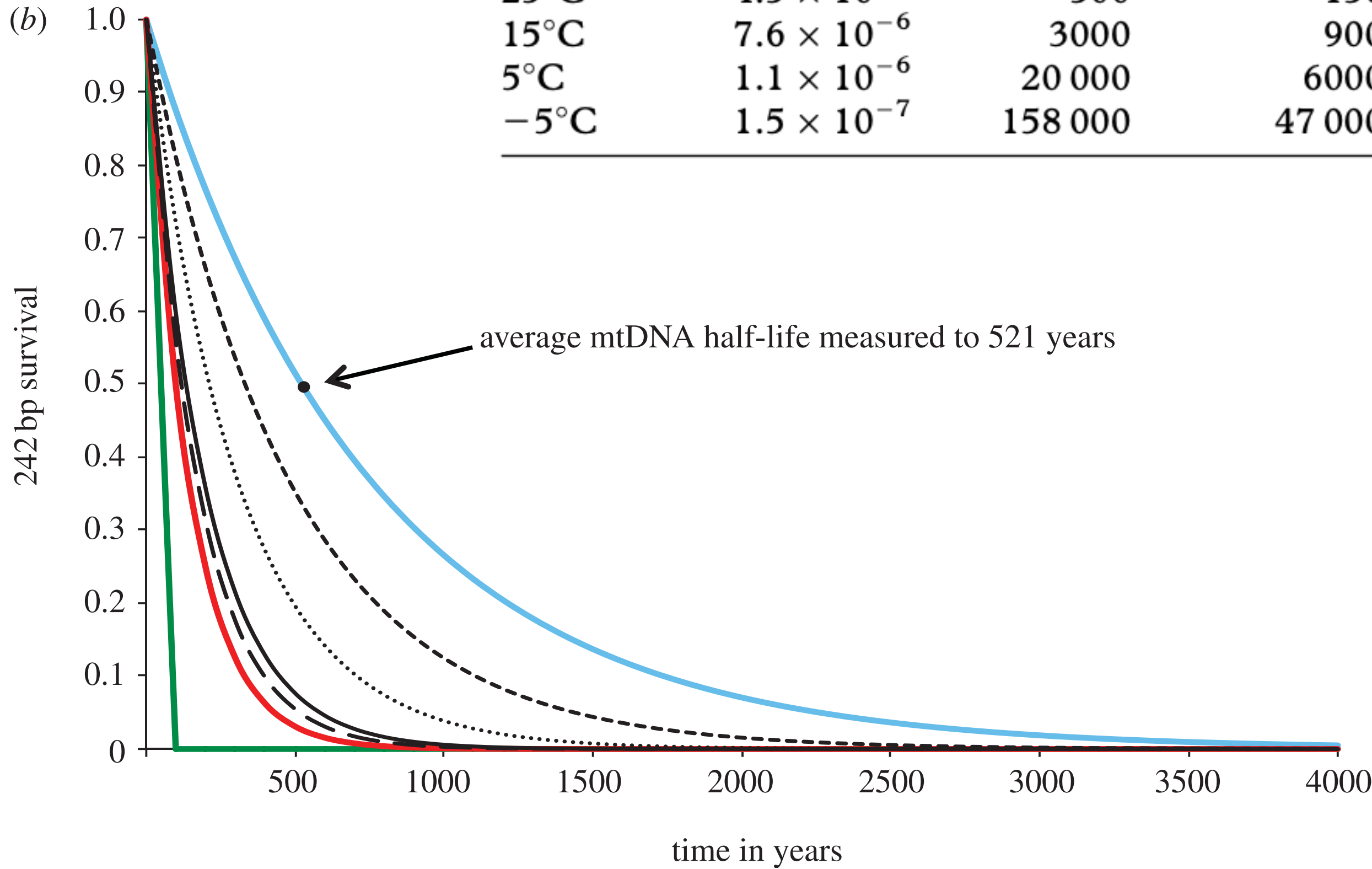


Ancient DNA fragmentation follows exponential decay dynamics



# Challenges - Ancient DNA preservation

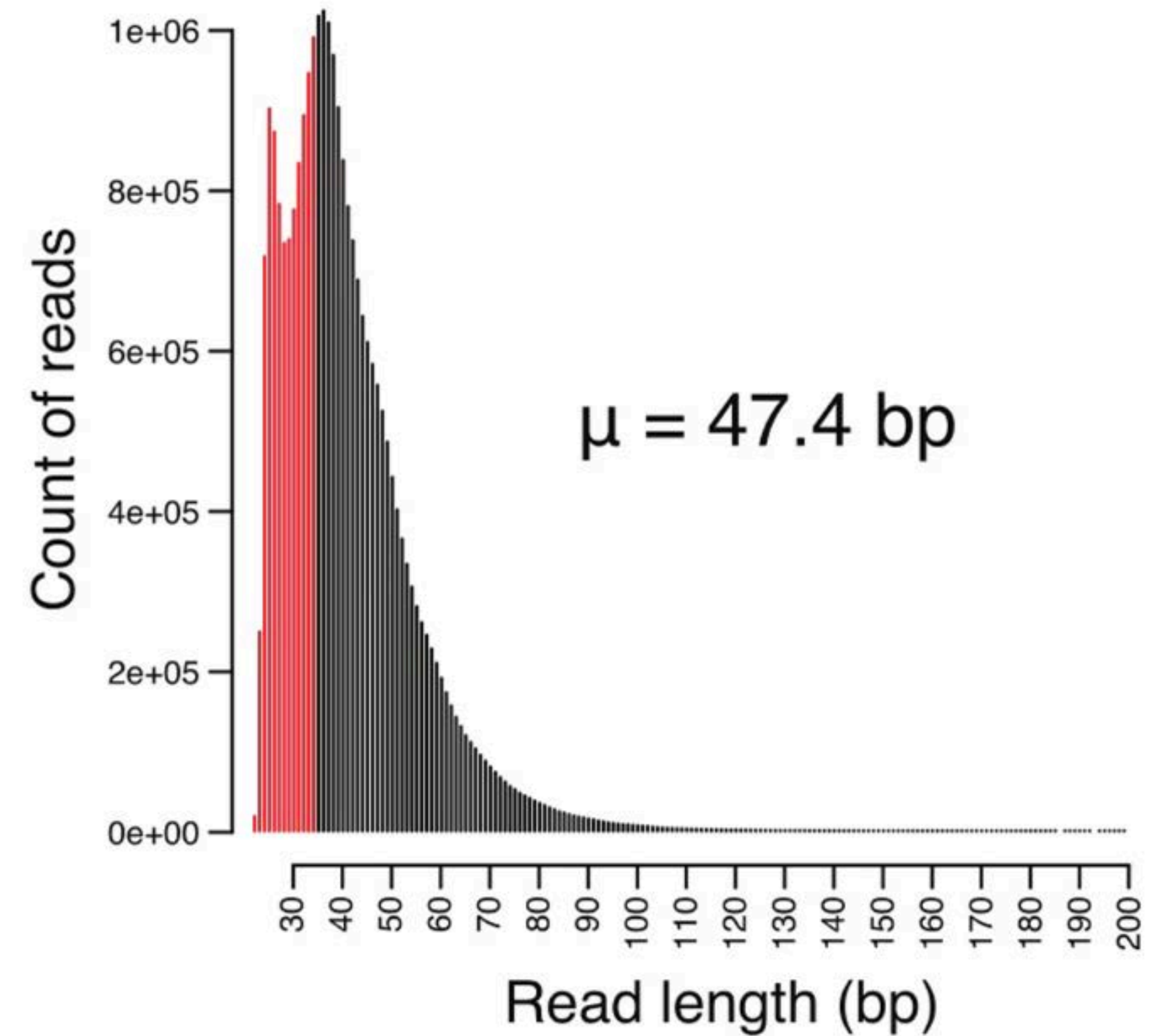
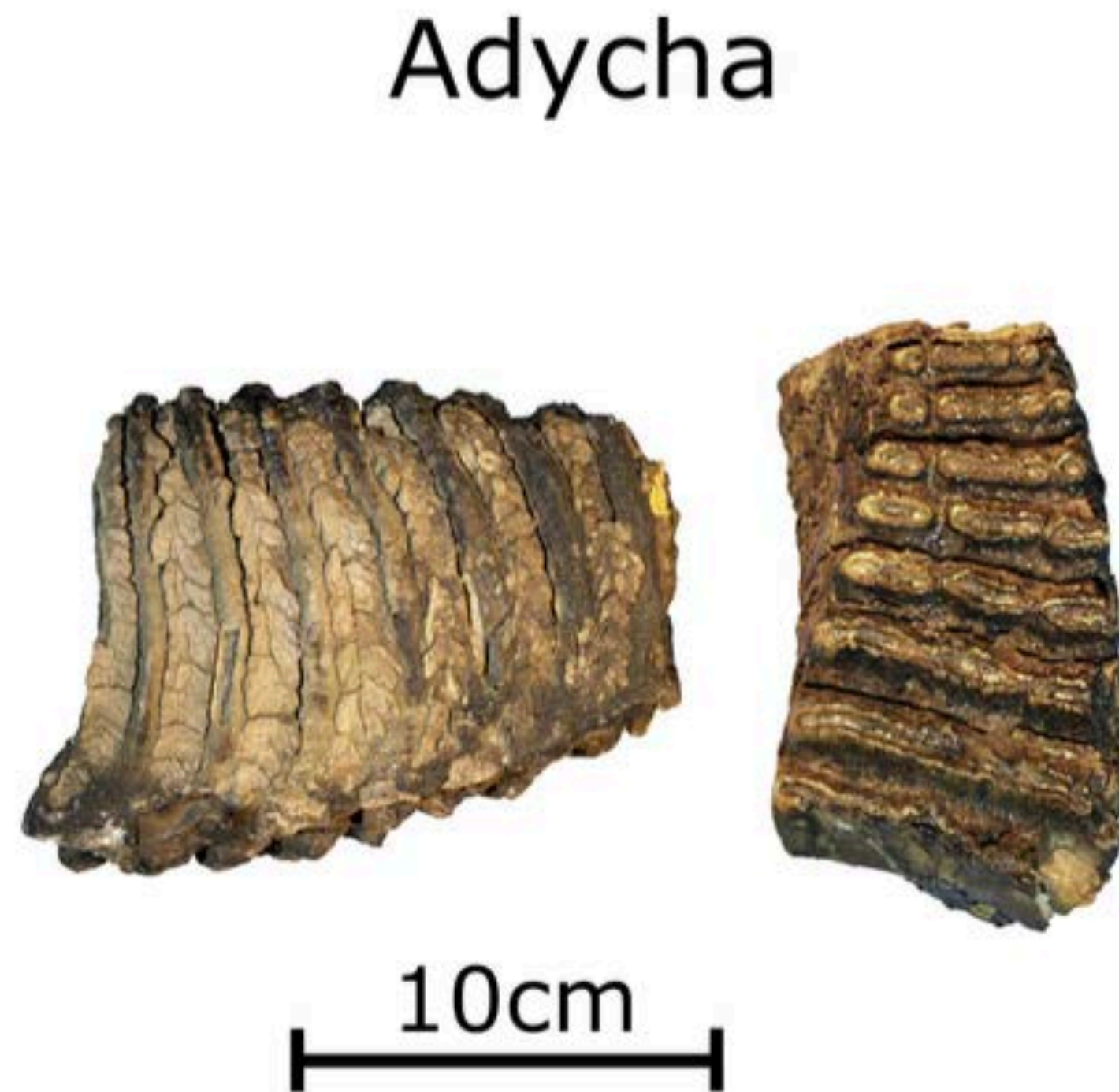
temperature	<i>k</i> per site per year	half-life (years), 30 bp	half-life (years), 100 bp	half-life (years), 500 bp	average length at 10 kyr	time (years) until average length = 1 bp
25°C	$4.5 \times 10^{-5}$	500	150	30	2 bp	22 000
15°C	$7.6 \times 10^{-6}$	3000	900	180	13 bp	131 000
5°C	$1.1 \times 10^{-6}$	20 000	6000	1200	88 bp	882 000
−5°C	$1.5 \times 10^{-7}$	158 000	47 000	9500	683 bp	6 830 000



Average fragment length of 6.8 million-year old DNA  
1 base pair



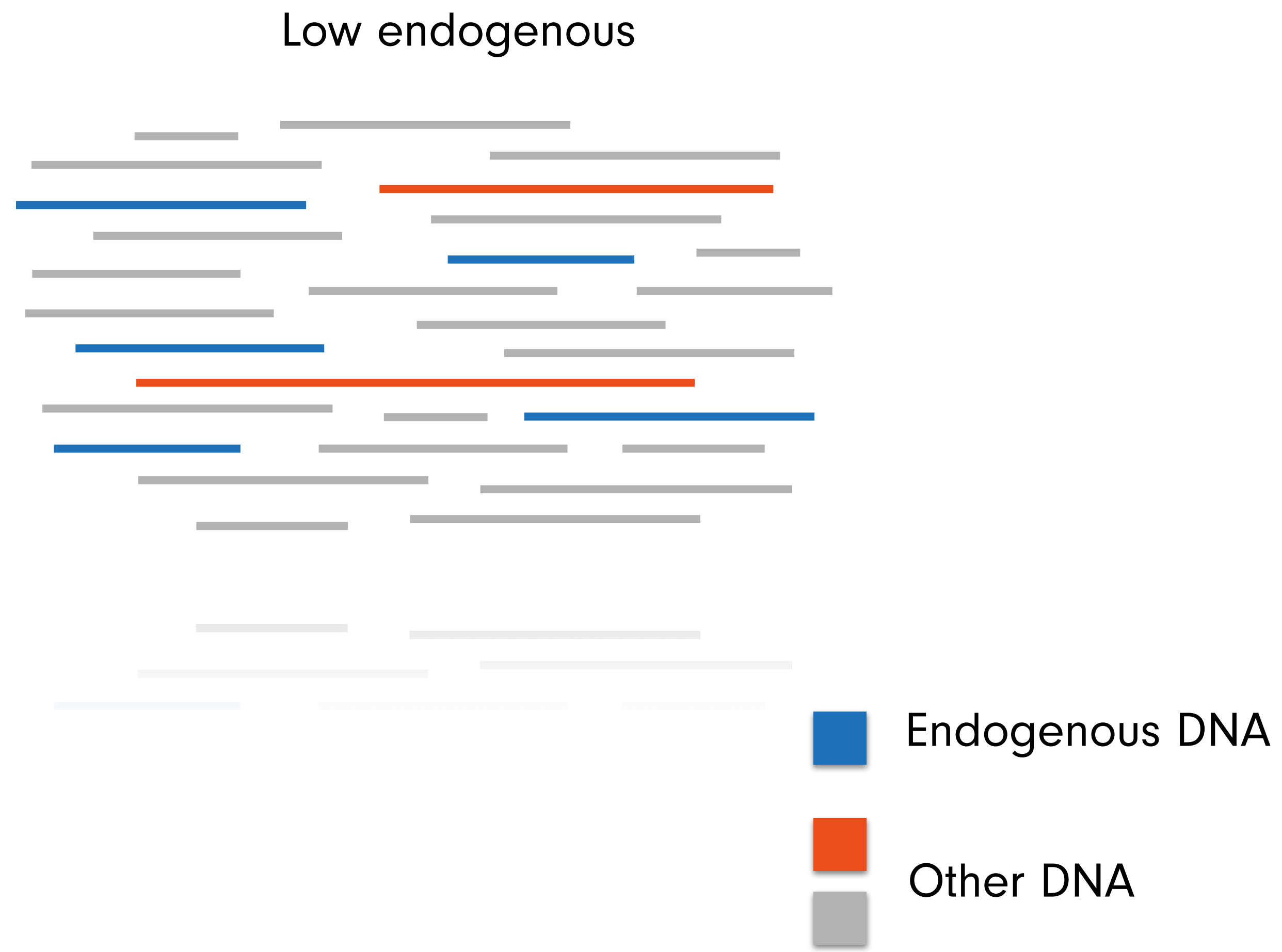
# Challenges - Ancient DNA preservation



DNA from a 1.2 million year old mammoth tooth is highly fragmented

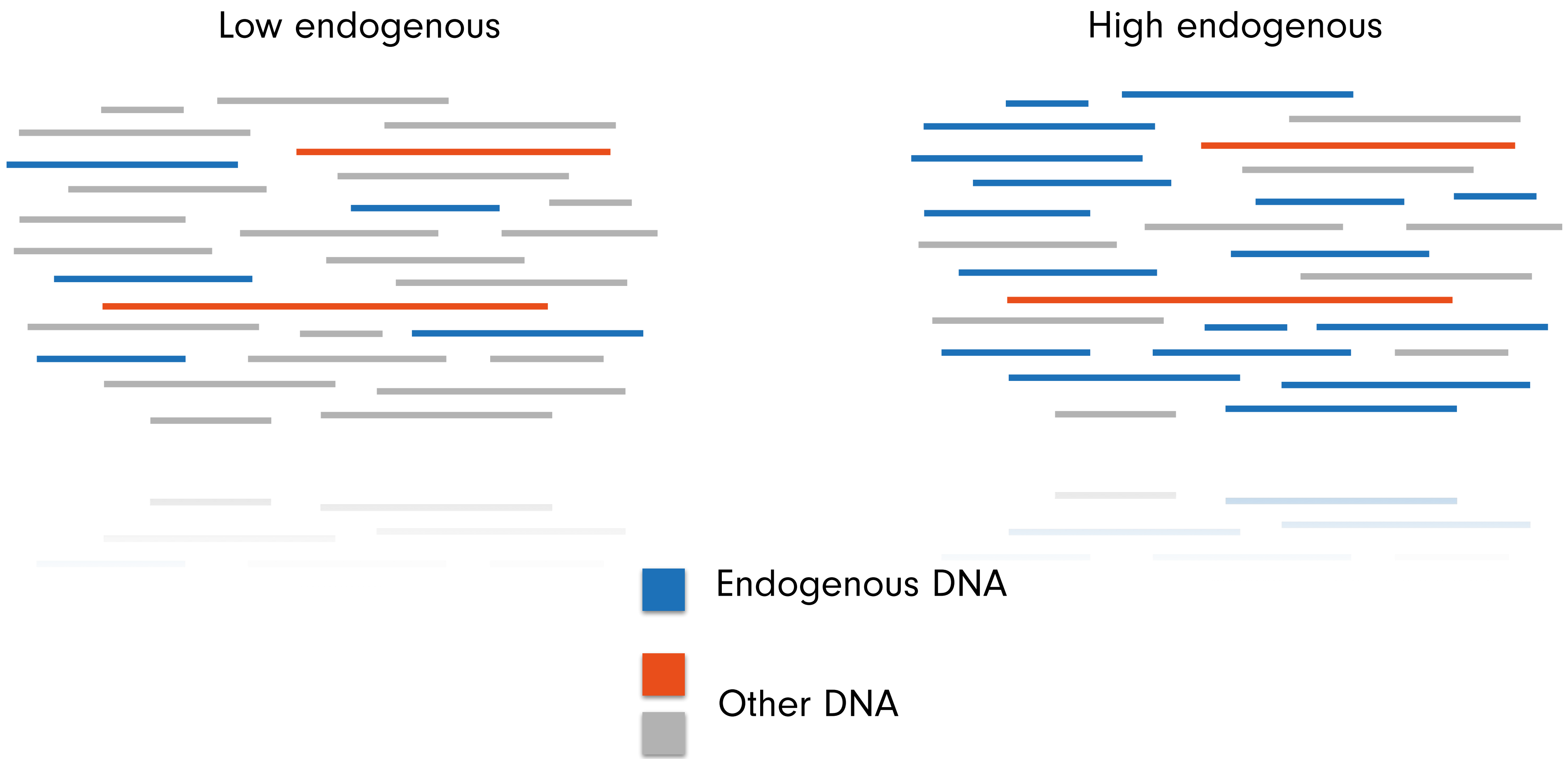


# Challenges - Endogenous DNA content





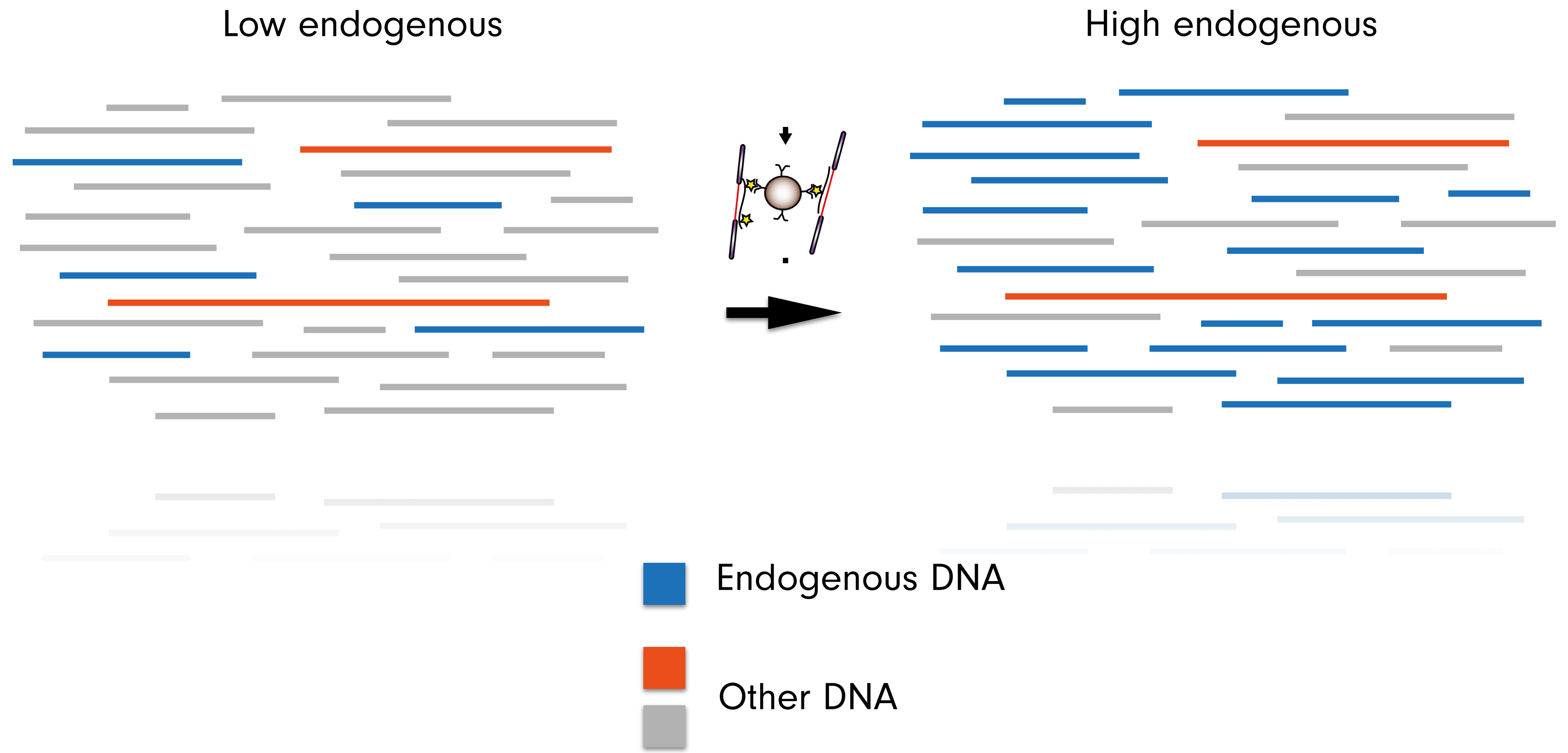
# Challenges - Endogenous DNA content



Endogenous DNA content varies substantially between samples



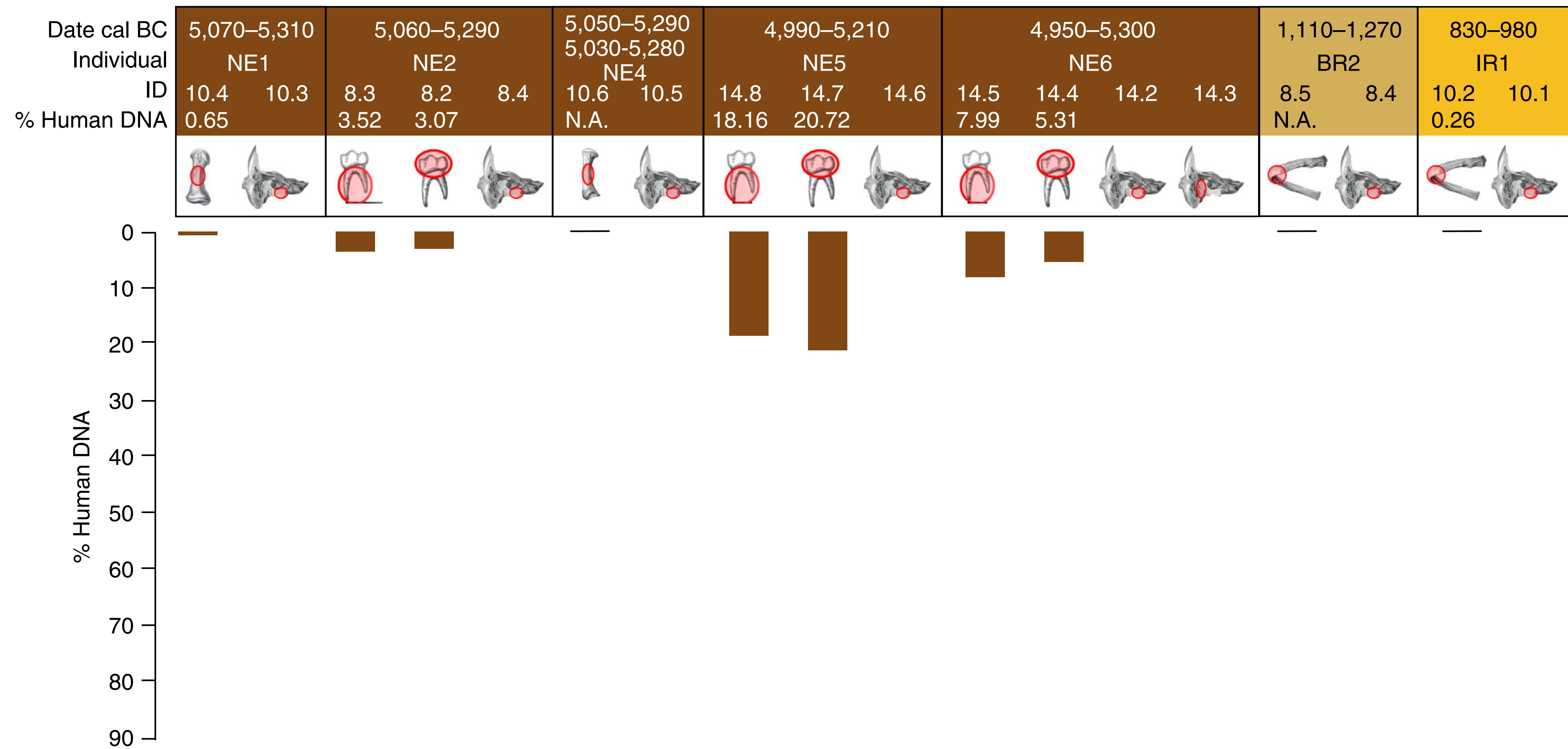
# Challenges - Endogenous DNA content



DNA capture enrichment (targeted or whole genome) to increase endogenous DNA

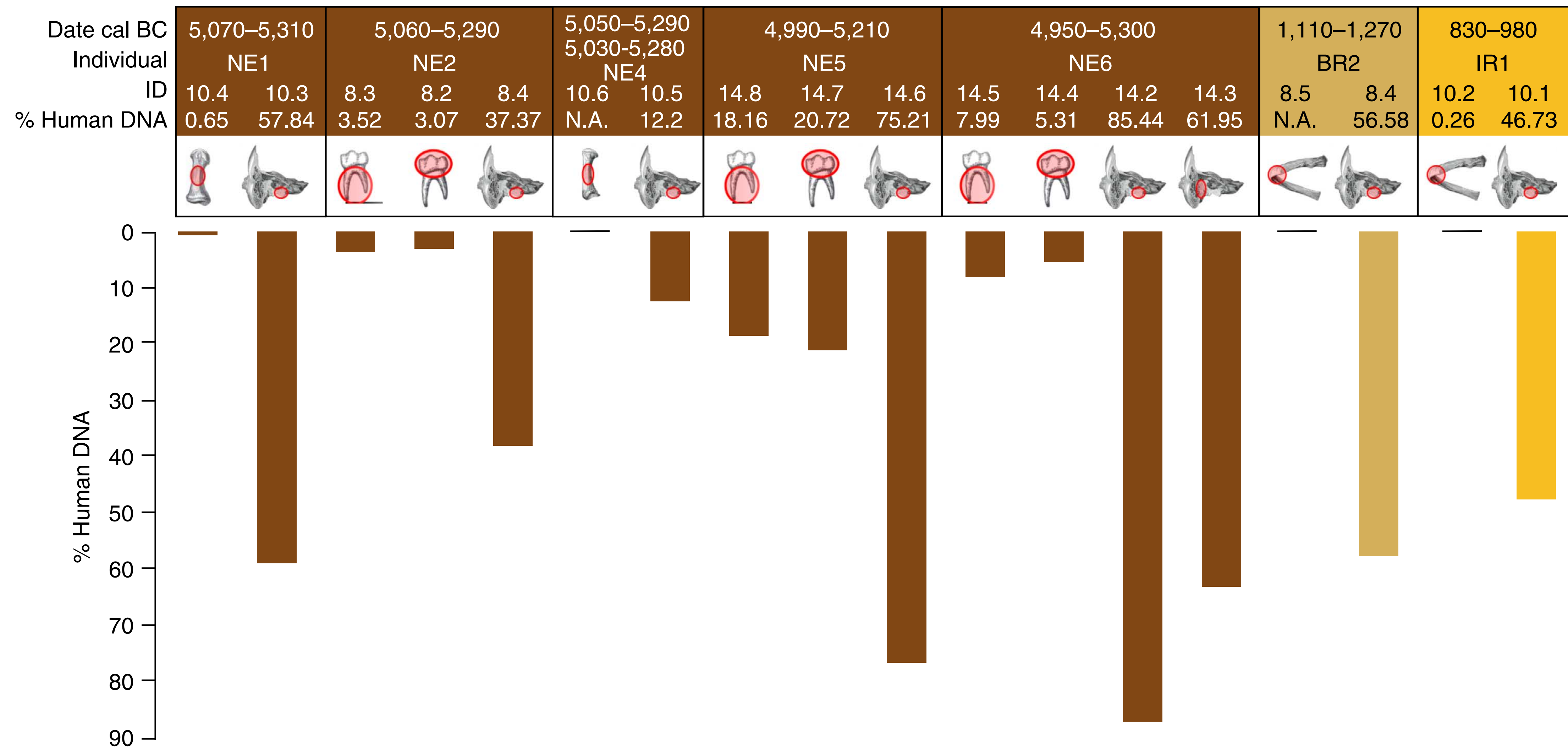


# Impact of sample material





# Impact of sample material





# Ancient DNA challenges

Sample

DNA sequences

Analysis results

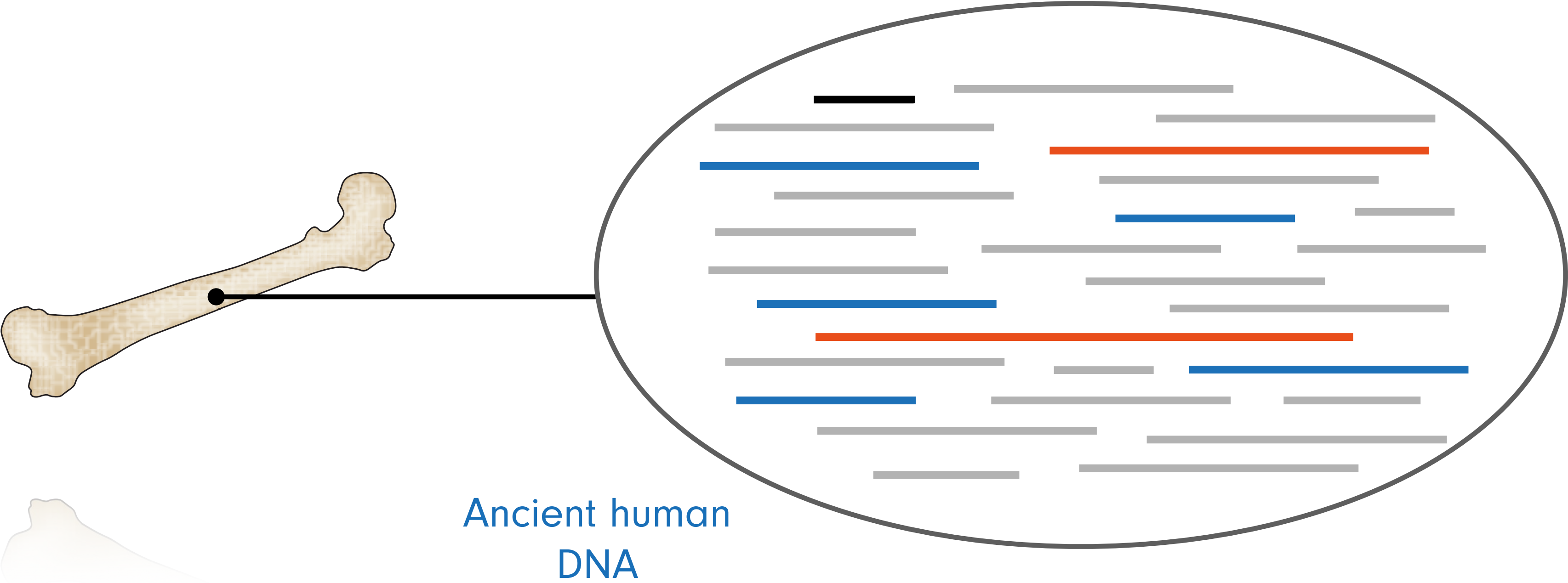


**Data quality**

- Contamination
- Genome coverage
- Error rates

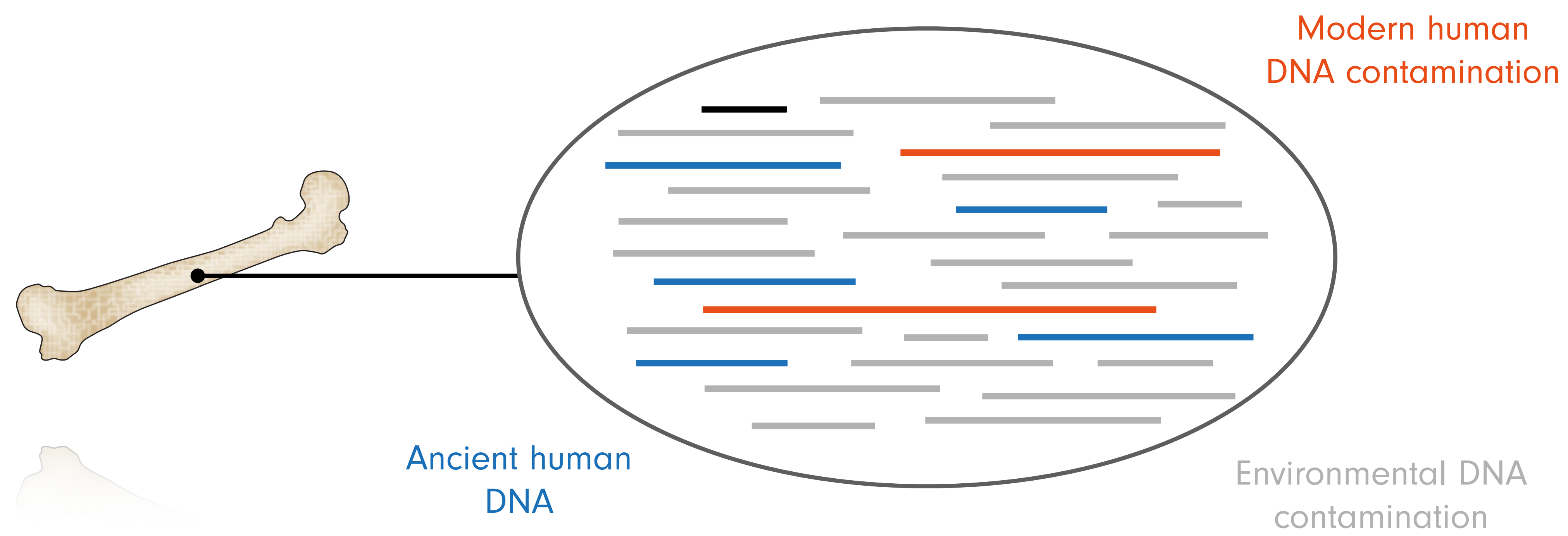


# Challenges - Contamination





# Challenges - Contamination



Ancient DNA data is a complex mixture of DNA from different sources



# Challenges - Contamination

Barcelona lab	Monti Lessini (L906-H924)	TCATCTACGCCTTCCACAGCCAGGAGCTCCGCAGGACGCTCAAGGAGGTGCTGACAT
	B.2.1	.....G.....
	B.2.2	.....G.....
	B.2.3	.....G.....
	B.2.4	..........
	B.2.5	..........
	B.2.6	..........
	B.2.7	..........
	B.2.8	..........
	B.2.9	..........
	B.2.10	..........
	B.2.11	..........
	B.2.12	..........
Florence lab	Monti Lessini (L884-H936)	TGCAATGCCATCATCGACCCCCTCATCTACGCCTTCCACAGCCAGGAGCTCCGCAGGACGCTCAAGGAGGTGCTGACATG
	CTCCTGGTGA	
	F.1.1	.....G.....
	F.1.2	.....G.....
	F.1.3	.....G.....
	F.1.4	.....G.....
	F.1.5	.....G.....
	F.1.6	.....C.....
	F.1.7	..C.....
	F.1.8	..........
	F.1.9	..........
	F.1.10	..........
	F.1.11	..........
	F.1.12	..........
	F.1.13	..........
	F.1.14	..........
	F.1.15	..........
	F.1.16	..........
	F.1.17	..........
	F.1.18	..........
	F.1.19	..........
	F.1.20	..........
	F.1.21	..........
	F.1.22	..........
	F.1.23	..........

MC1R gene fragments amplified from Monti Lessini Neandertal DNA extract

# Challenges - Contamination

Barcelona lab

Monti Lessini (L906-H924)

- B.2.1
- B.2.2
- B.2.3
- B.2.4
- B.2.5
- B.2.6
- B.2.7
- B.2.8
- B.2.9
- B.2.10
- B.2.11
- B.2.12

TCATCTACGCCTTCCACAGCCAGGAGCTCCGCAGGACGCTCAAGGAGGTGCTGACAT

.....G.....
.....G.....
.....G.....
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.....

Neandertal sequences

Florence lab

Monti Lessini (L884-H936)

- CTCCTGGTGA
- F.1.1
  - F.1.2
  - F.1.3
  - F.1.4
  - F.1.5
  - F.1.6
  - F.1.7
  - F.1.8
  - F.1.9
  - F.1.10
  - F.1.11
  - F.1.12
  - F.1.13
  - F.1.14
  - F.1.15
  - F.1.16
  - F.1.17
  - F.1.18
  - F.1.19
  - F.1.20
  - F.1.21
  - F.1.22
  - F.1.23

TGCAATGCCATCATCGACCCCTCATCTACGCCTTCCACAGCCAGGAGCTCCGCAGGACGCTCAAGGAGGTGCTGACATG

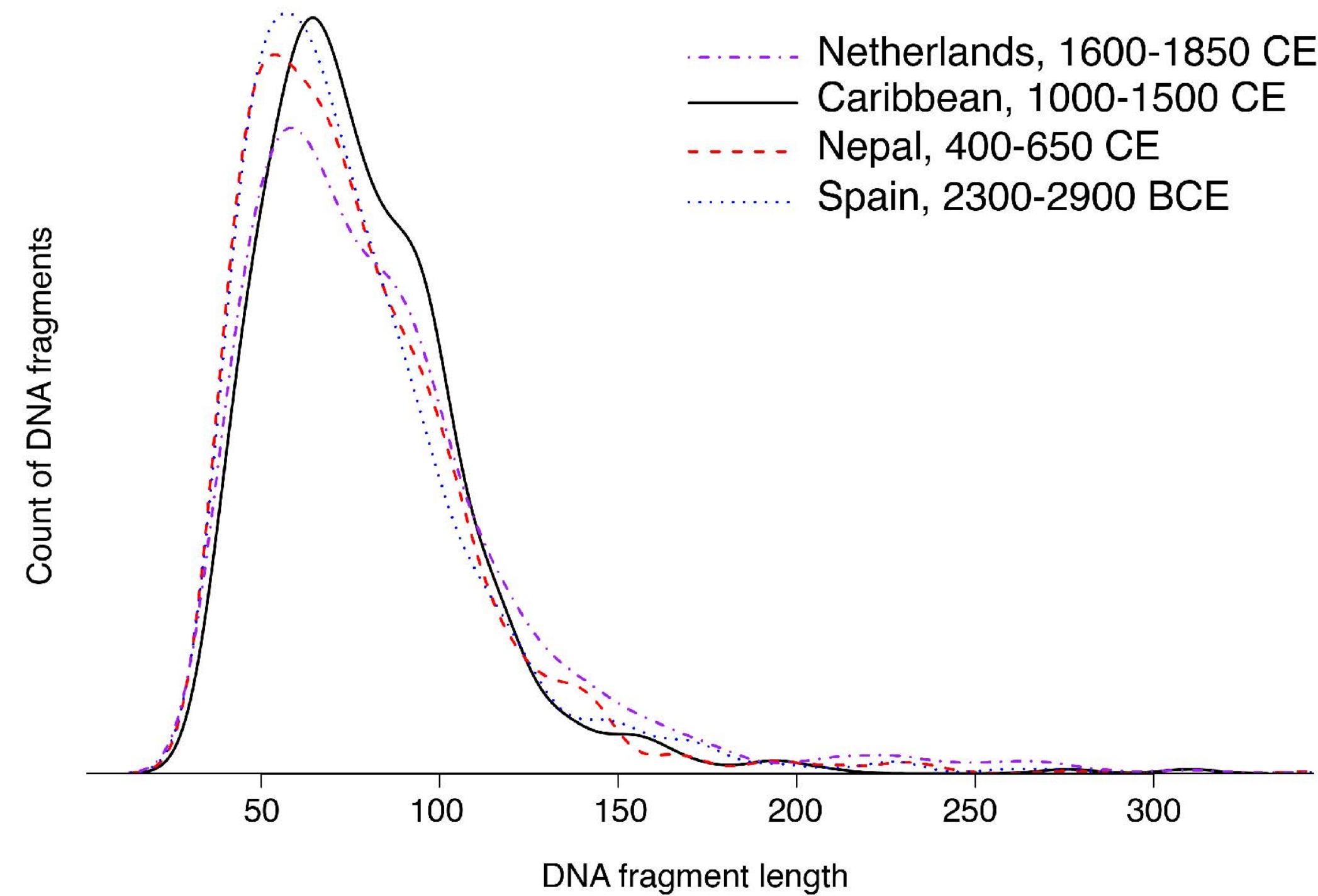
.....G.....
.....G.....
.....G.....
.....G.....
.....G.....
.....C.....
..C.....
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Neandertal sequences

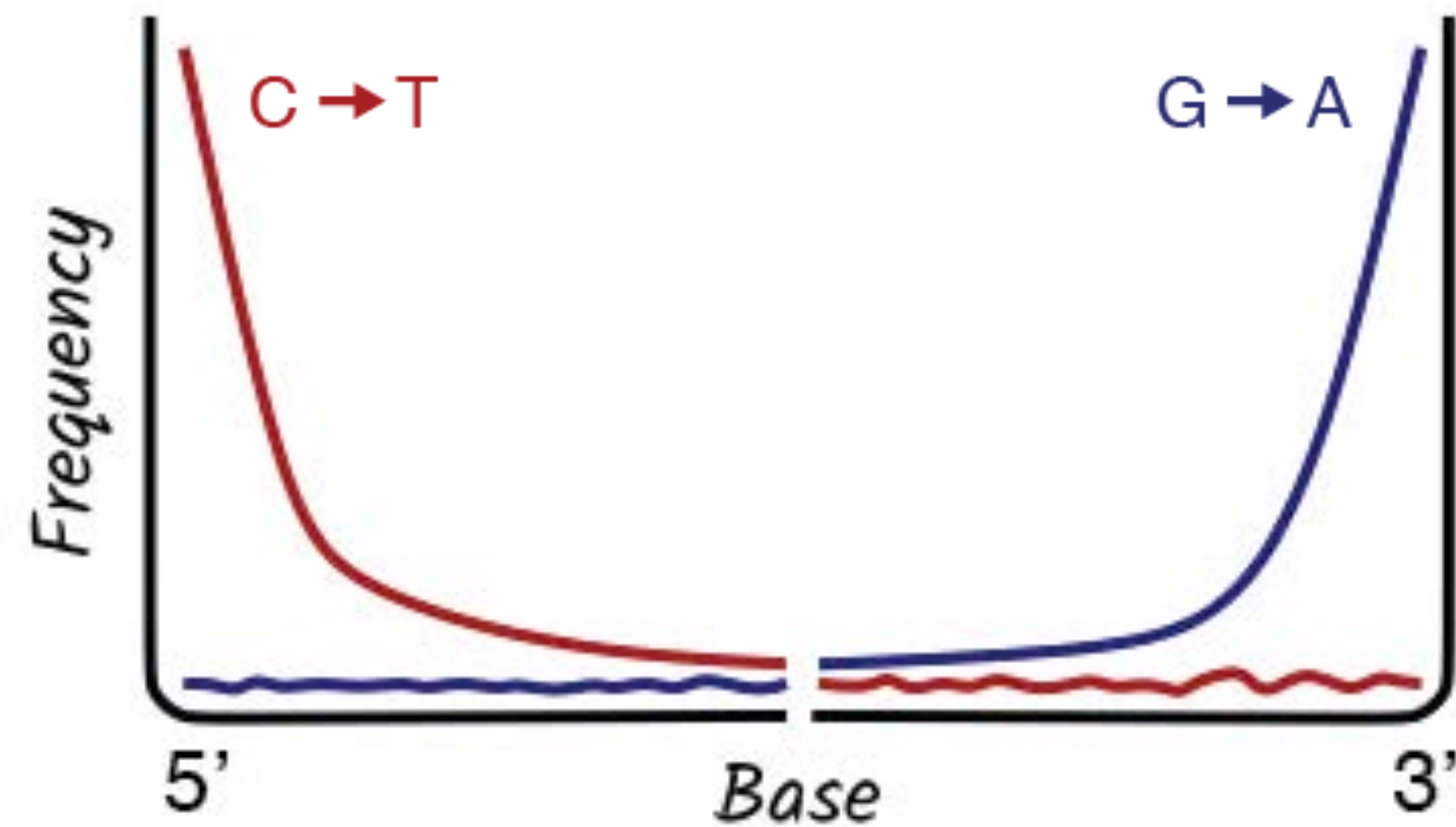
MC1R gene fragments amplified from Monti Lessini Neandertal DNA extract



# Authentication of ancient DNA



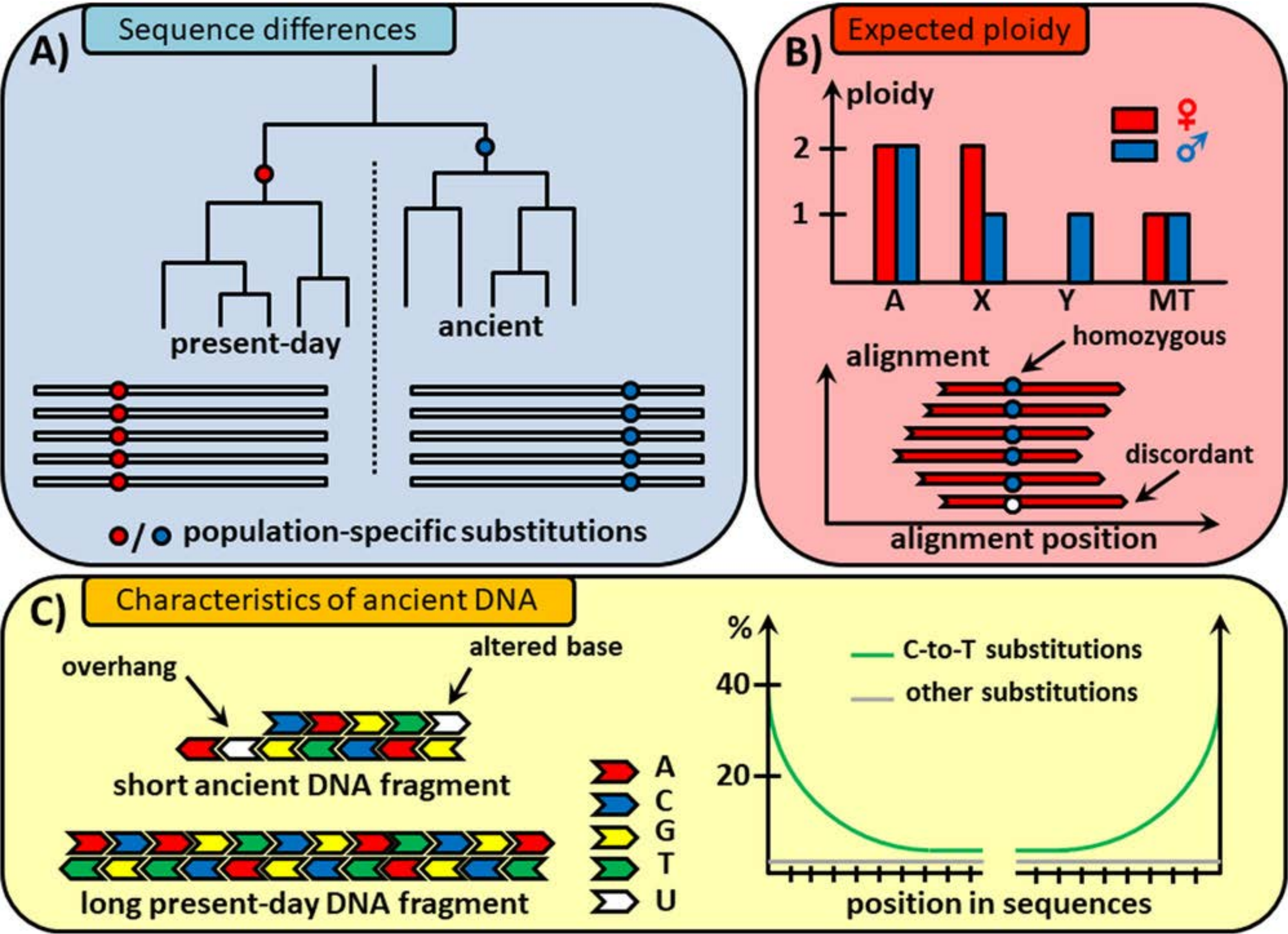
Ancient DNA is short and fragmented



Ancient DNA is damaged with characteristic substitution patterns



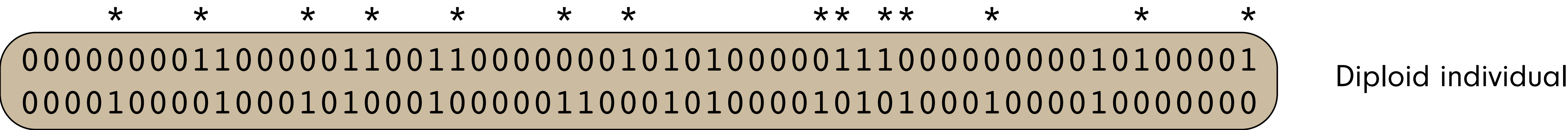
# Authentication of ancient DNA



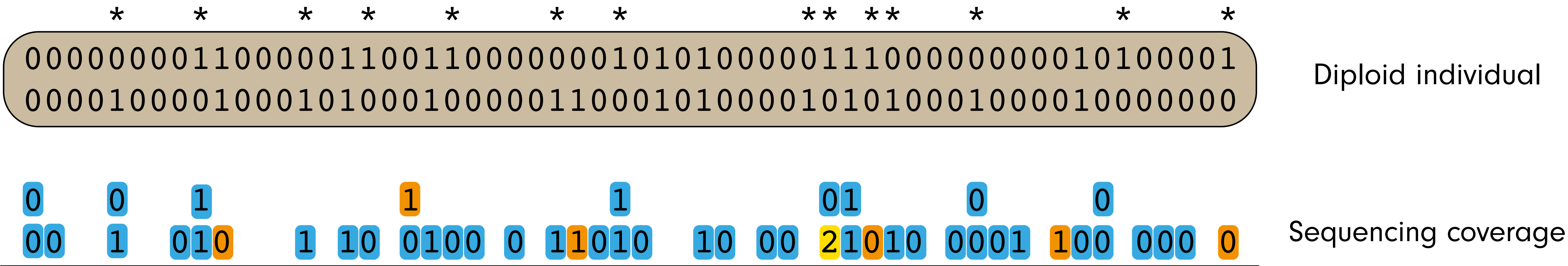
Signatures used to estimate contamination levels



# Challenges - Genome coverage and errors

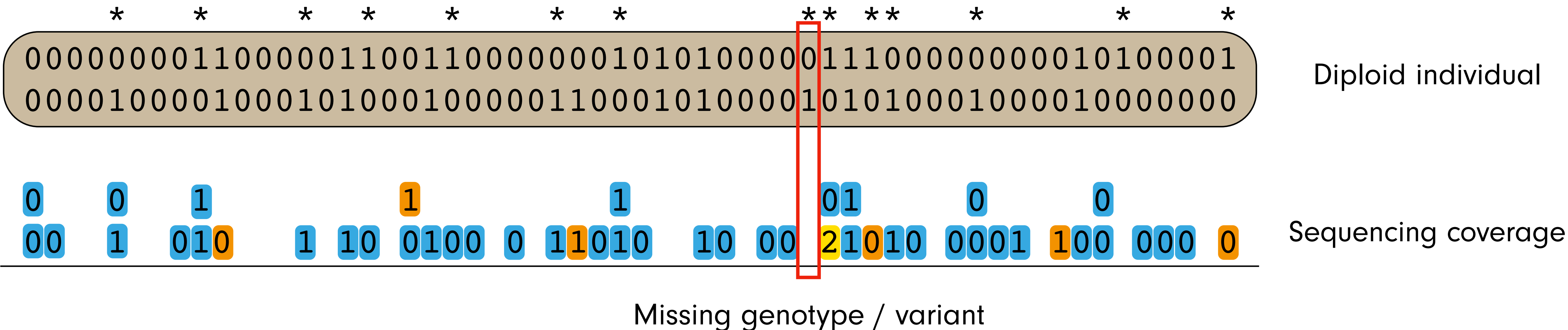


# Challenges - Genome coverage and errors

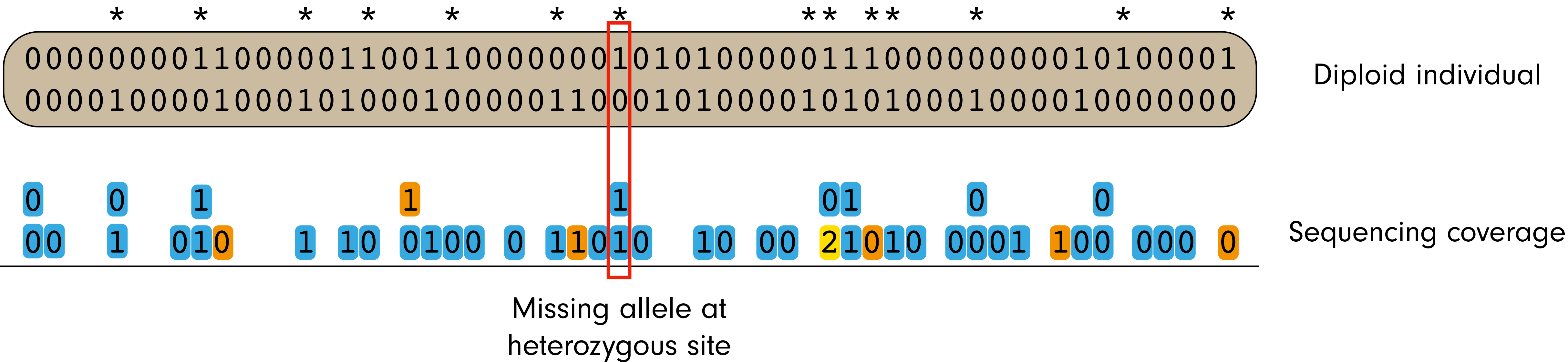




# Challenges - Genome coverage and errors

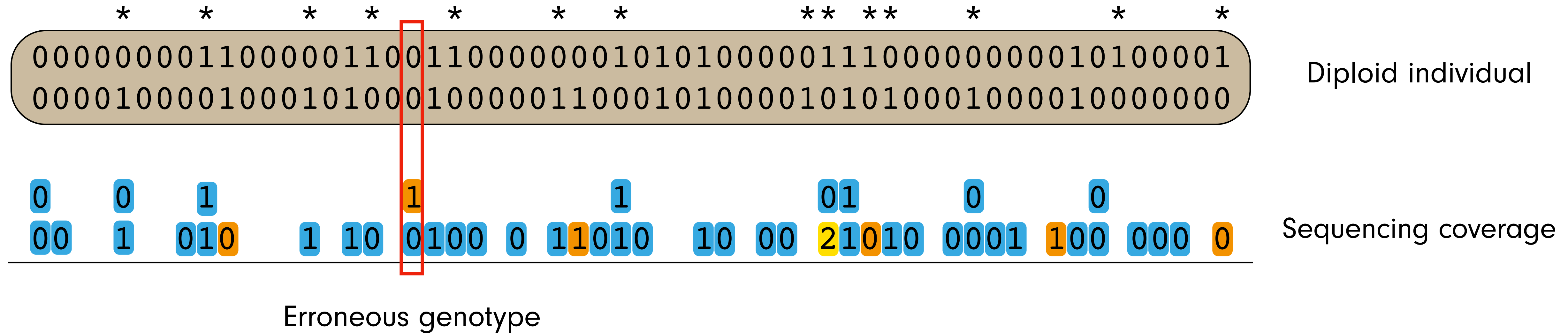


# Challenges - Genome coverage and errors





# Challenges - Genome coverage and errors



# Challenges - Genome coverage and errors

\* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \*  
0000000011000001100110000000101010000011100000000010100001  
000010000100010100010000011000101000001010100010000100000000

# Diploid individual

0 0 1 1 1 0 1 0 0 1 0 1 0 0 0 0 0

00 1 010 1 10 0100 0 11010 10 00 21010 00001 100 000 0

## Sequencing coverage

00..1..010...1.10.1100.1.11010..10.00.21010.0001.100.000.0

# Pseudo-haploid genotypes



# Challenges - Genome coverage and errors

The diagram shows a horizontal bar representing a 32-bit register. Above the bar, three asterisks (\*) are positioned at the 10th, 18th, and 26th bit positions, indicating three multi-bit multipliers. The register contains the following 32 bits (from left to right): 0, 0, 0, 0, 0, 0, 1, 1, 0, 0, 0, 0, 1, 1, 0, 0, 1, 1, 0, 0, 0, 0, 0, 0, 1, 0, 1, 0, 1, 0, 0, 0, 0, 0, 1, 1, 1, 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 1, 0, 0, 0, 0, 1. The bits are grouped into four 8-bit segments, each containing a bolded '1' at the 7th bit position (0-indexed).

## Diploid individual

0 0 1 1 1 0 1 0 0 0 1 0 0 0 0 0

00 1 01 10 0100 0 11010 10 00 21010 00001 100 000 0

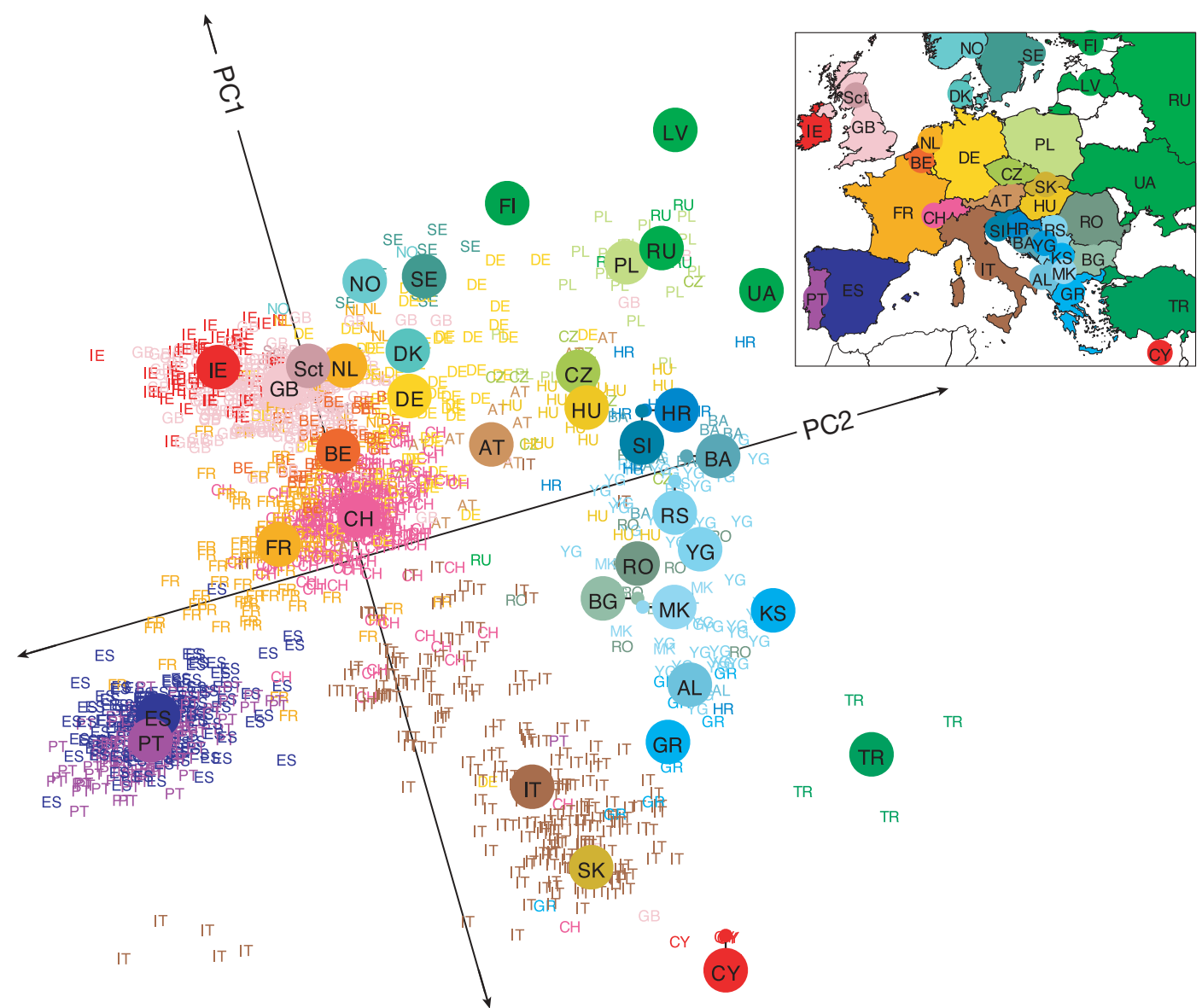
## Sequencing coverage

[illegible]

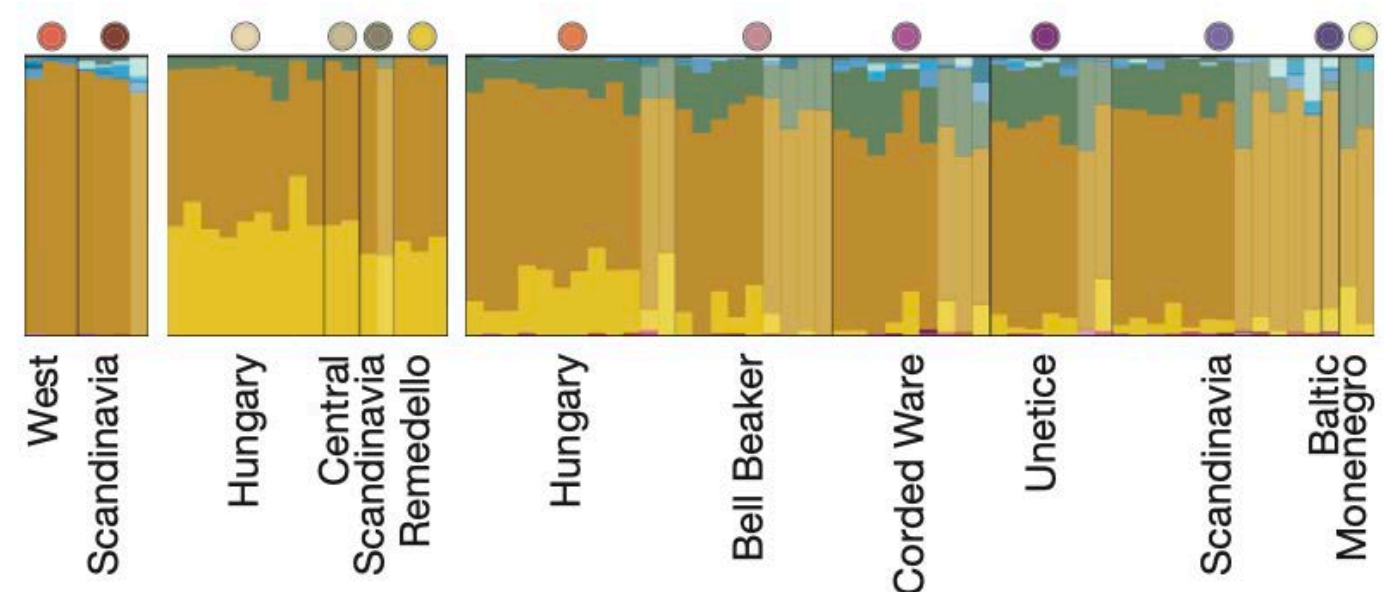
Pseudo-haploid  
genotypes at pre-  
ascertained sites

# Population genetic analysis of ancient DNA data

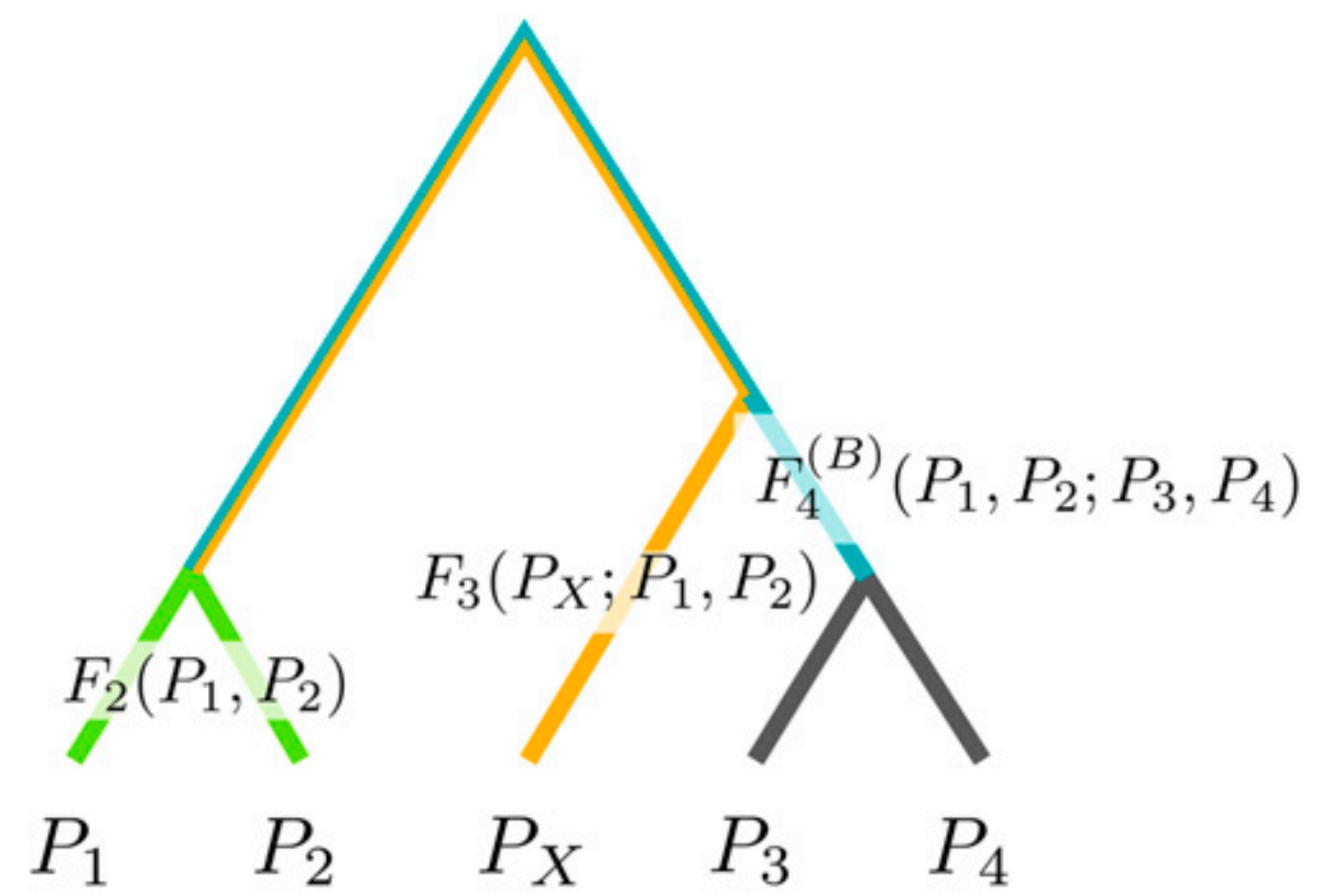
Principal component analysis



Model-based clustering



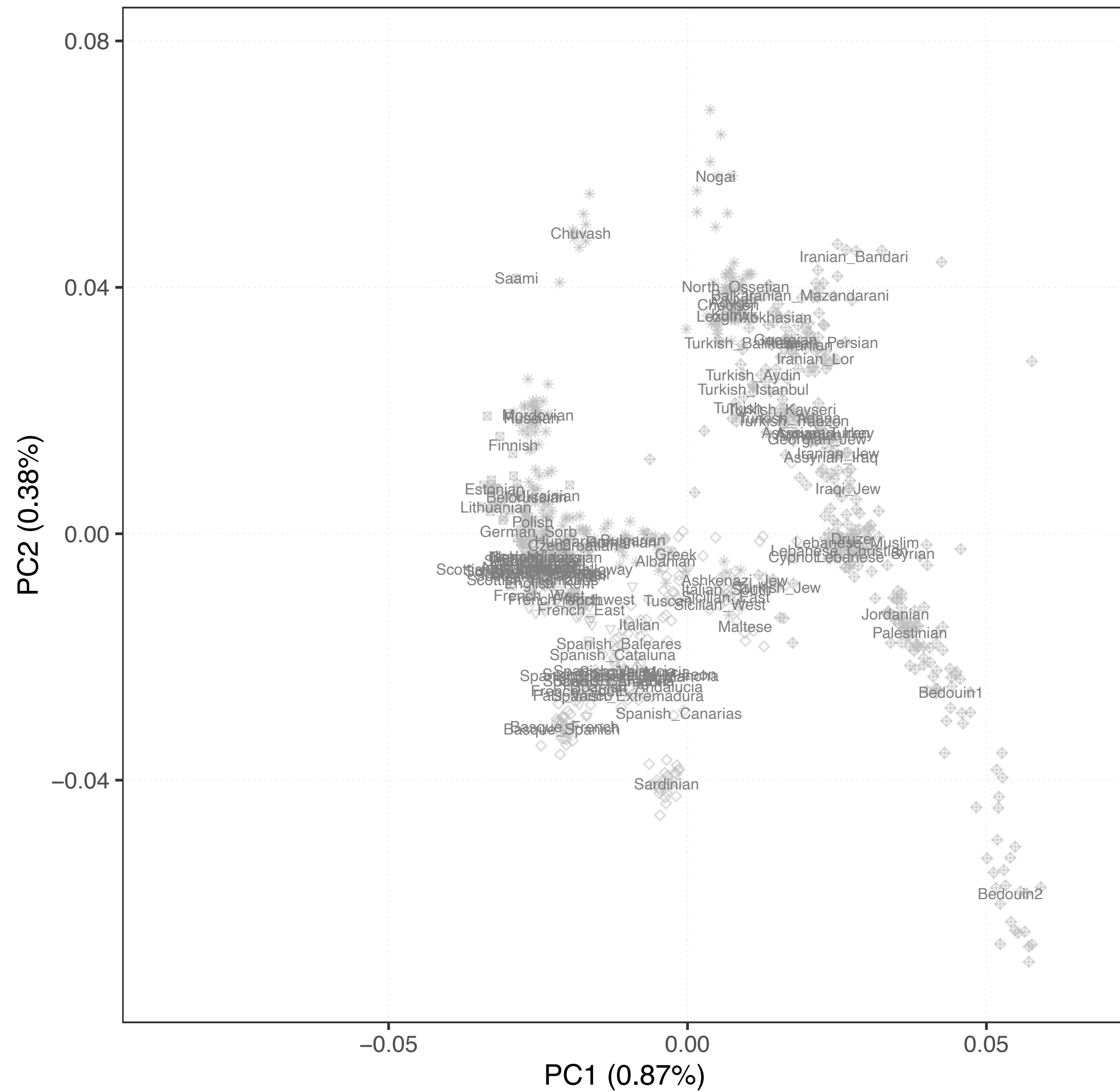
*f*-statistics



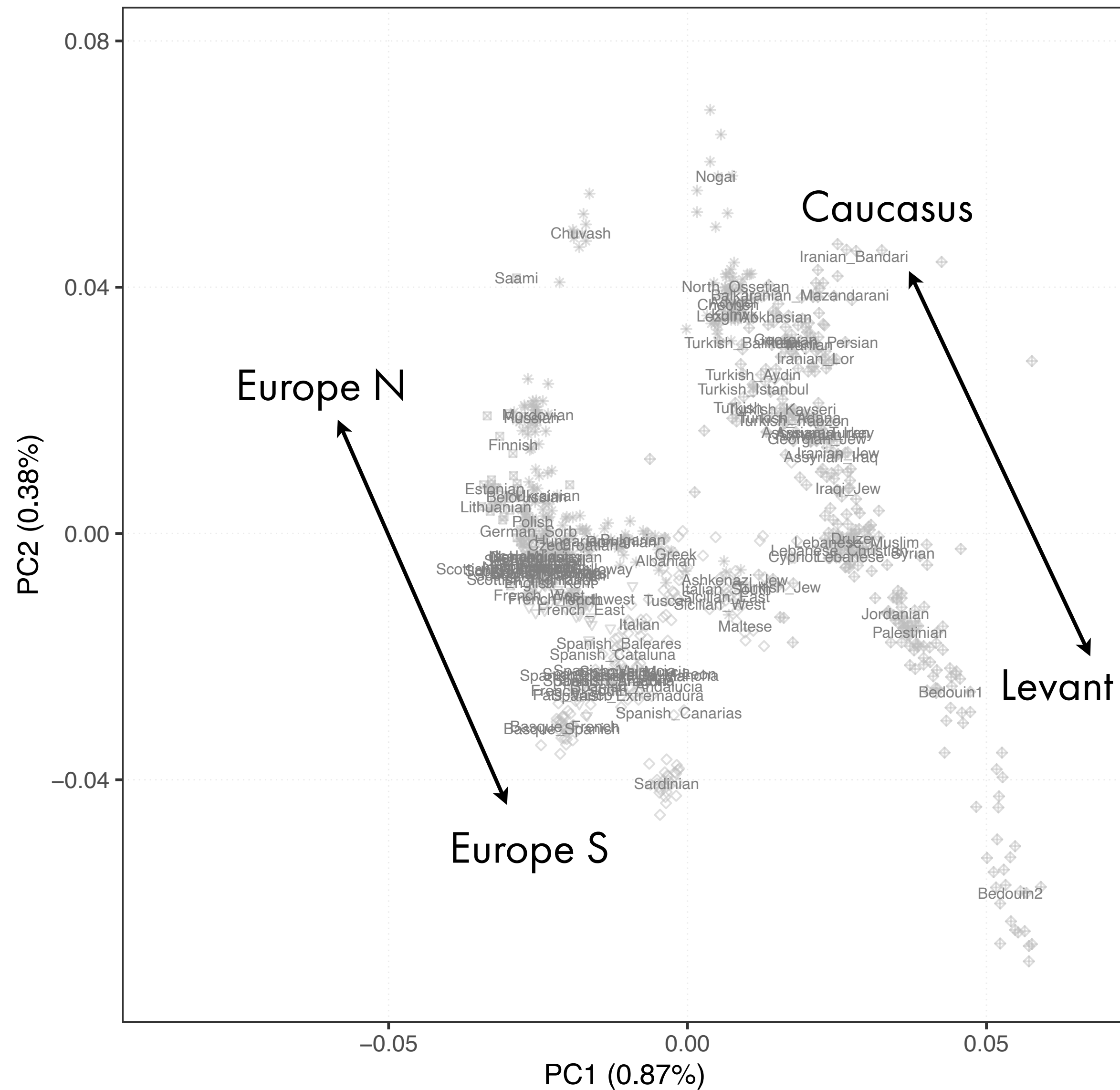
Allele-frequency based methods suited for low coverage / pseudo-haploid data



# A PCA tour of western Eurasia

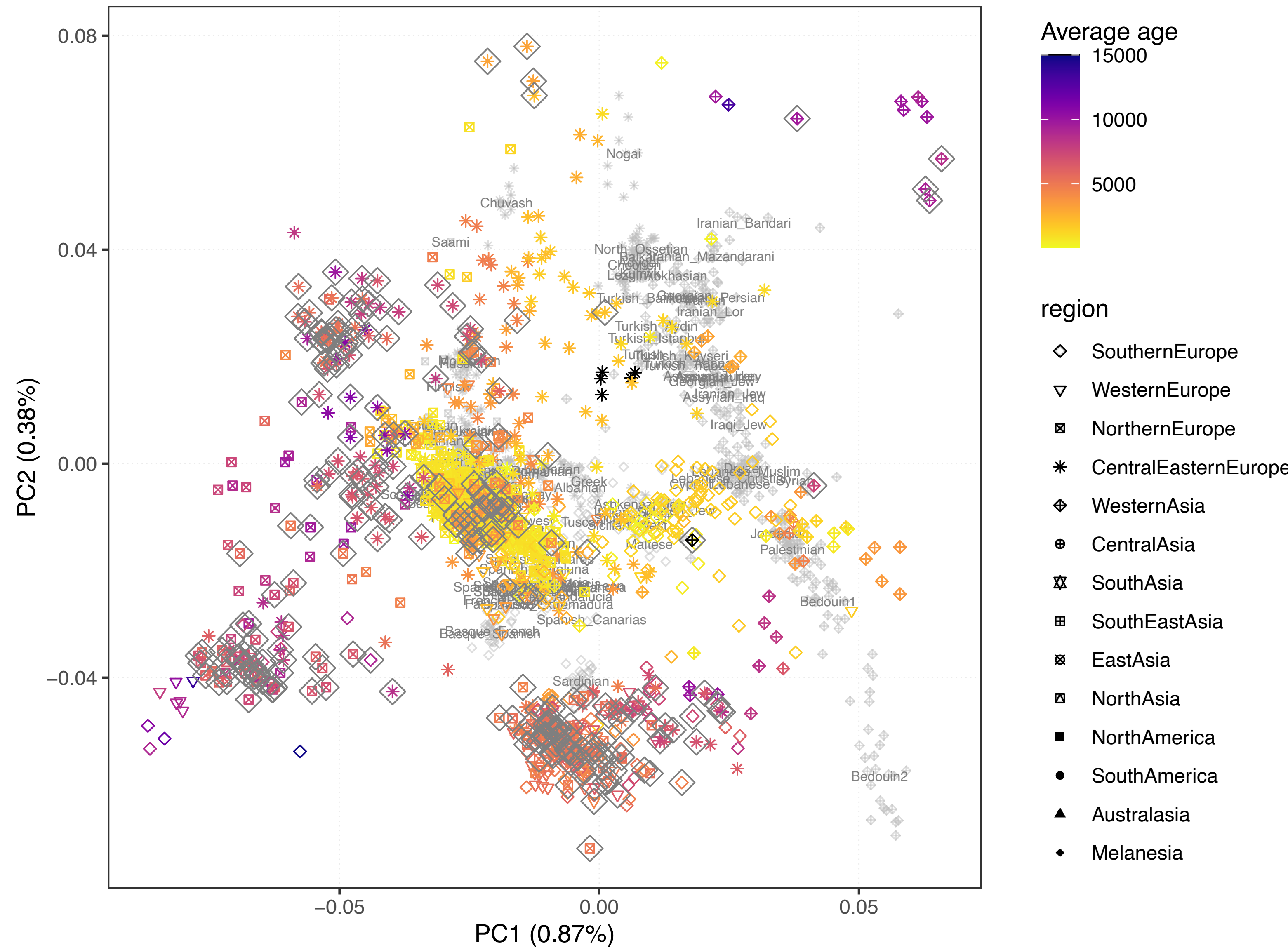


# A PCA tour of western Eurasia





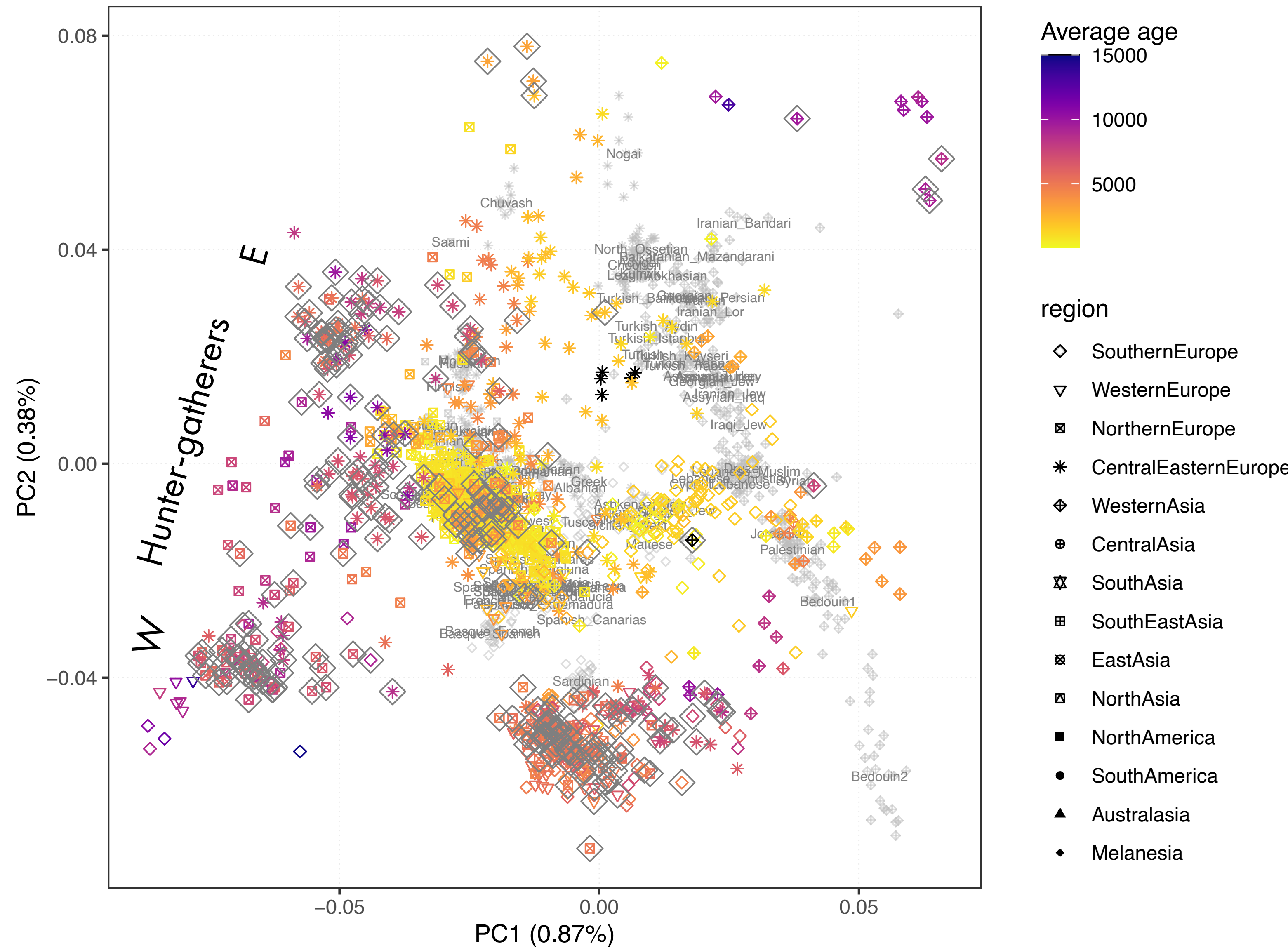
# A PCA tour of western Eurasia



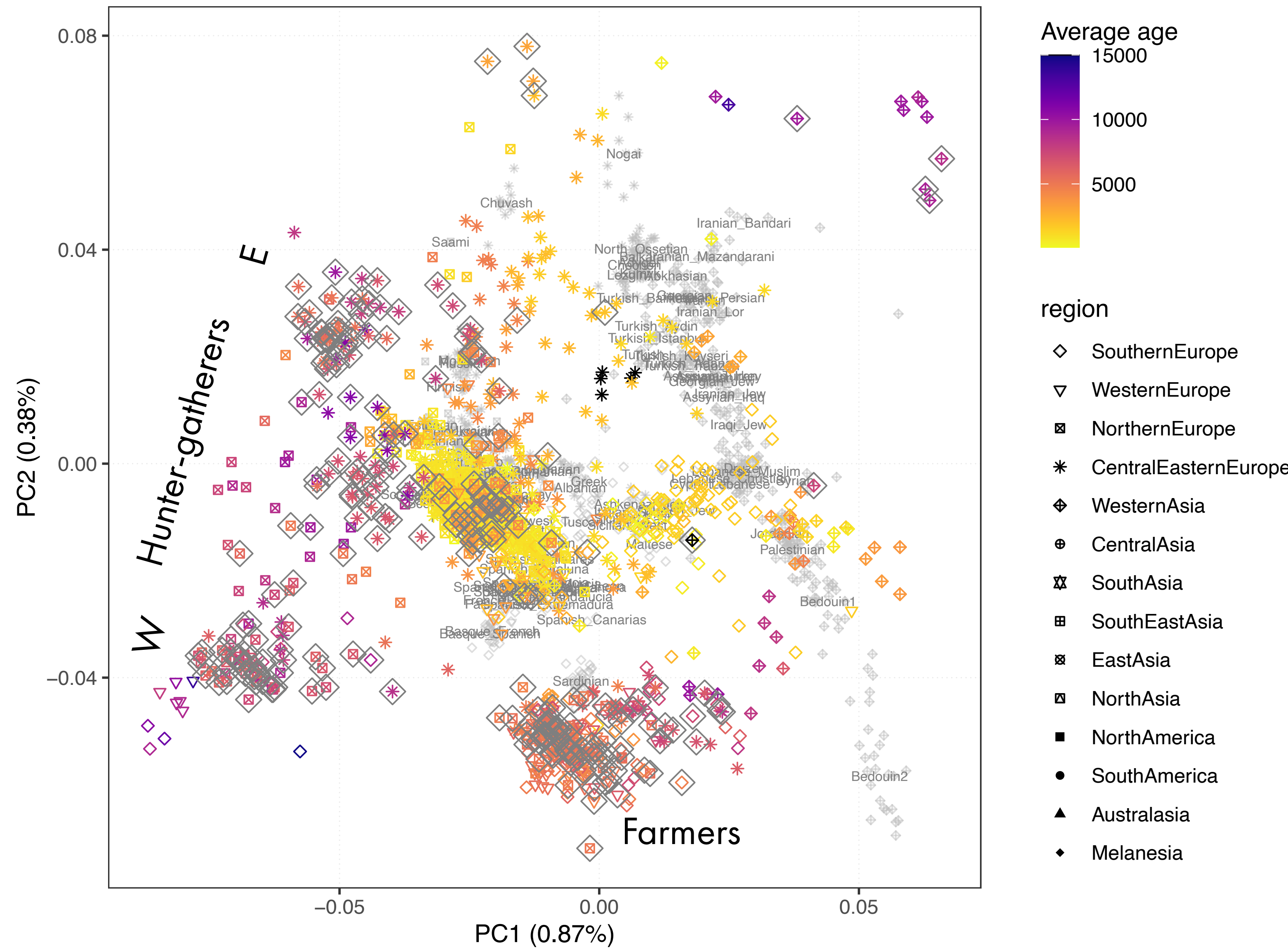




# A PCA tour of western Eurasia

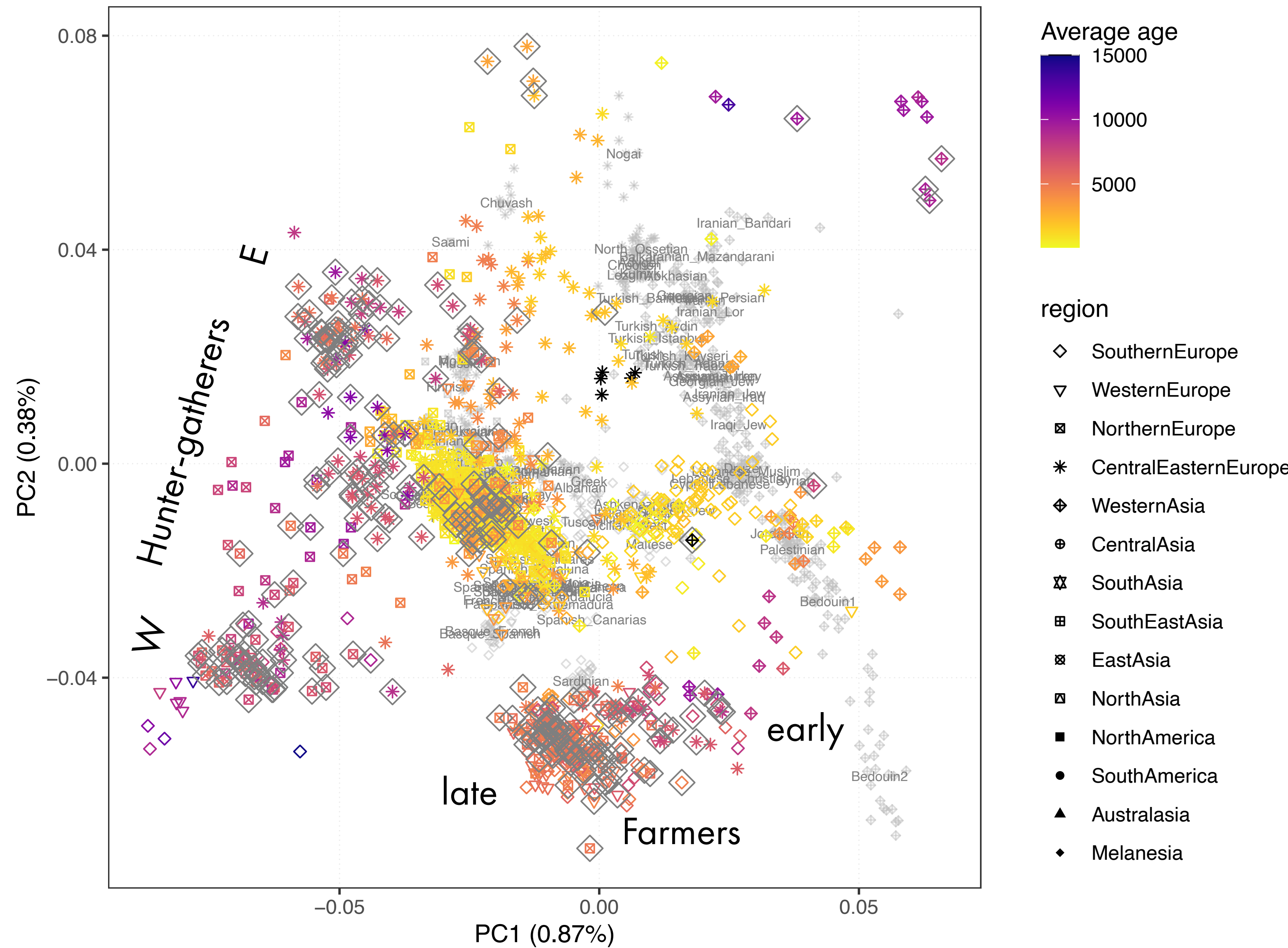


# A PCA tour of western Eurasia

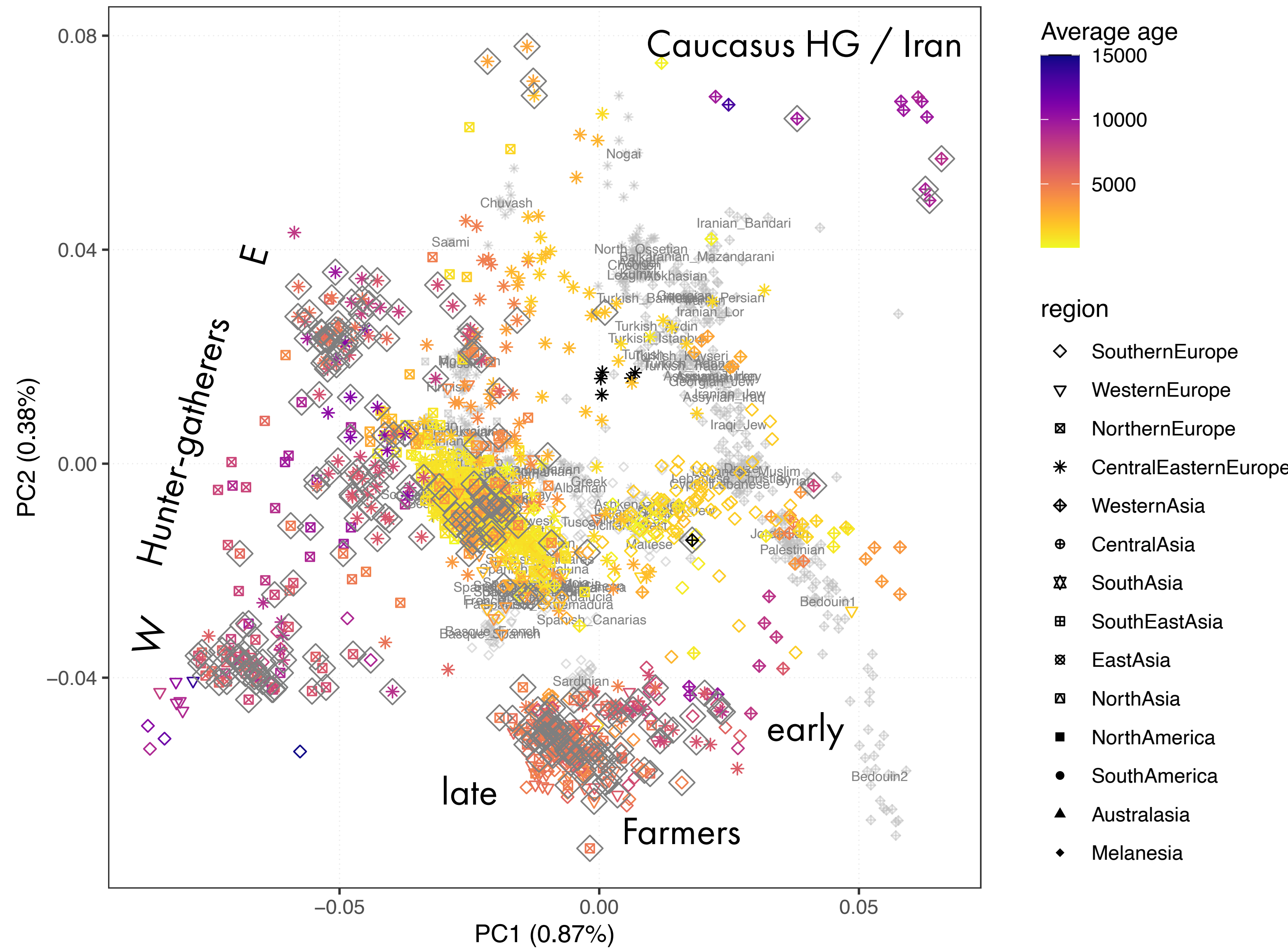




# A PCA tour of western Eurasia

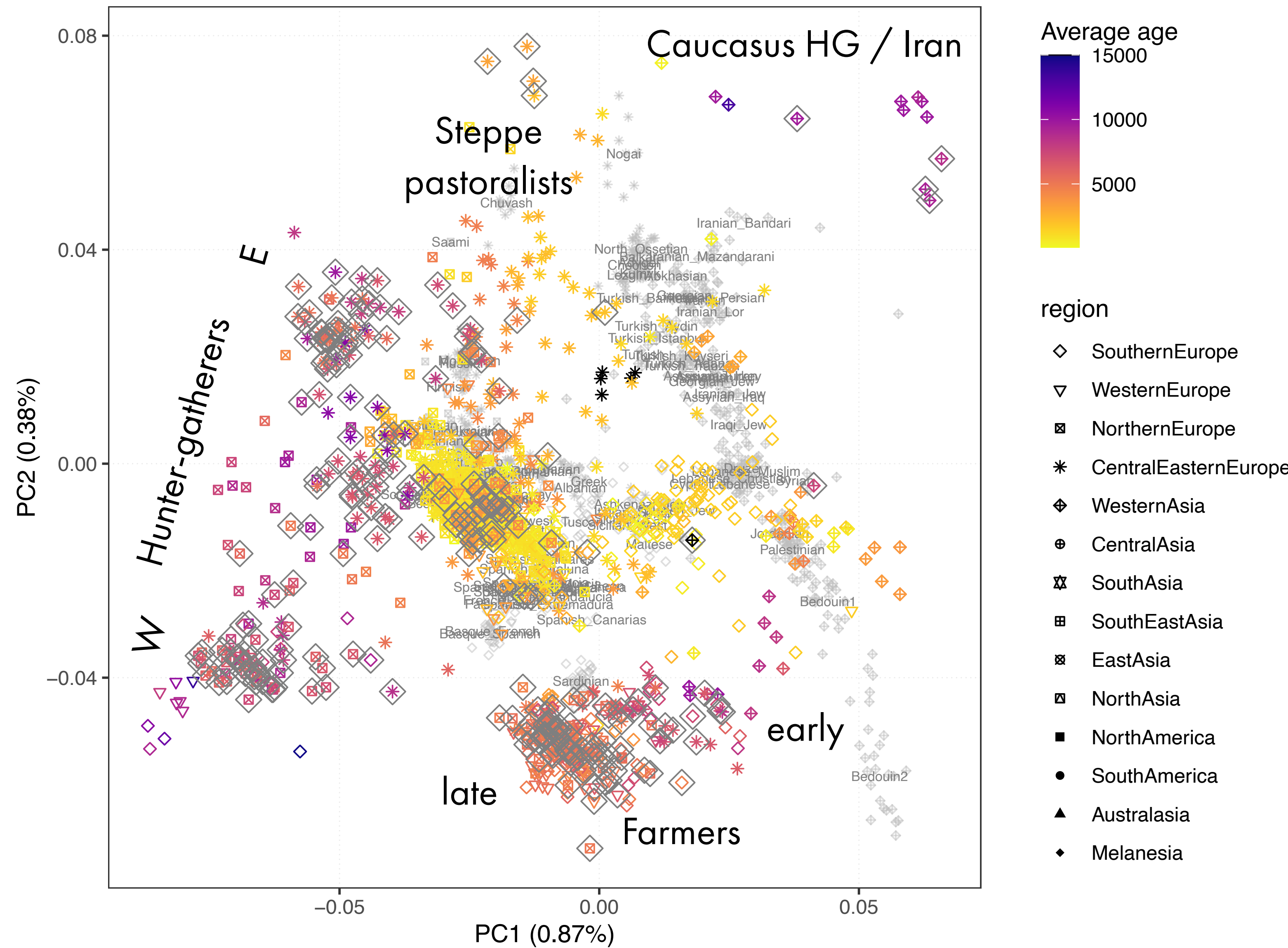


# A PCA tour of western Eurasia

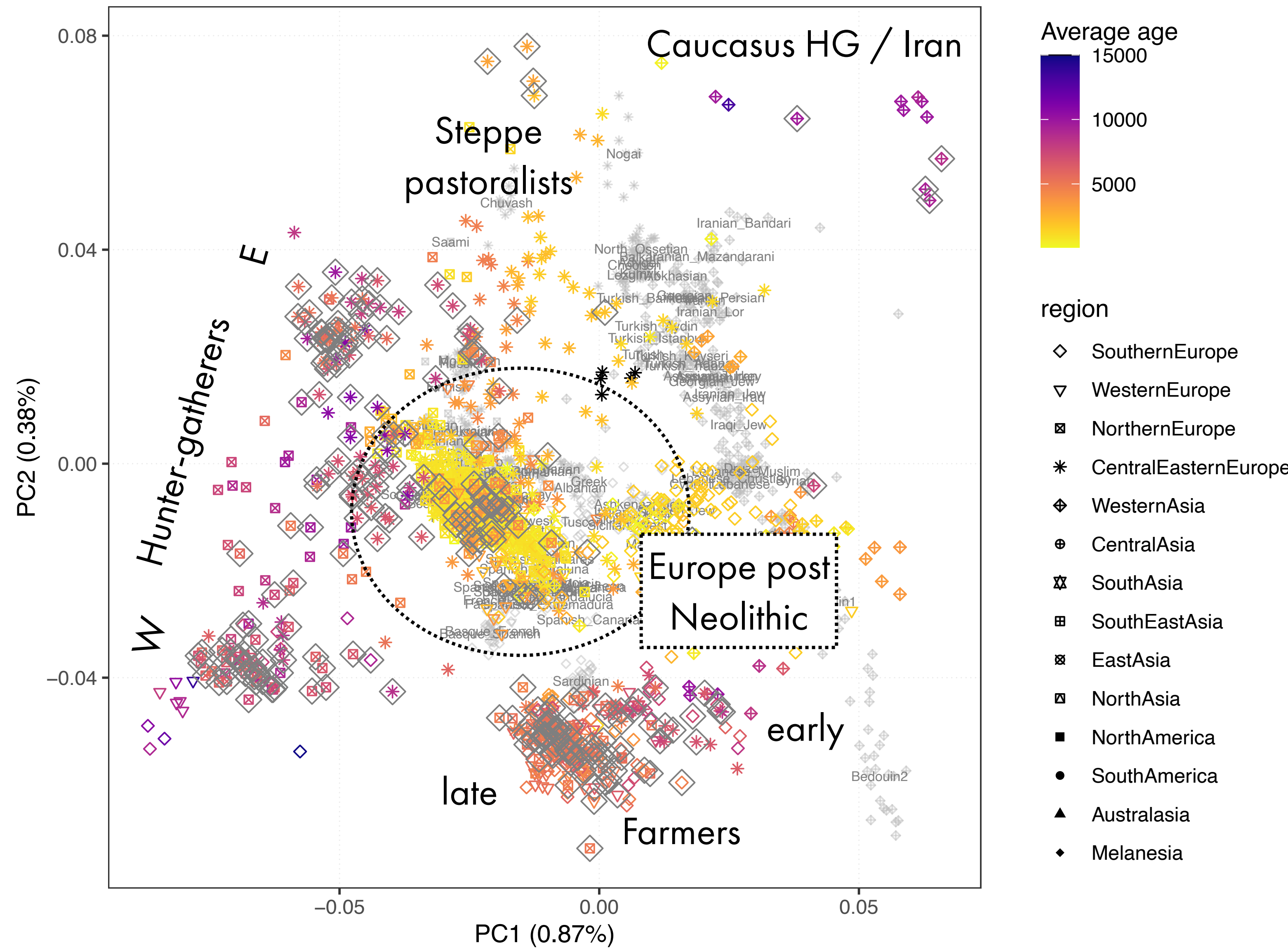




# A PCA tour of western Eurasia

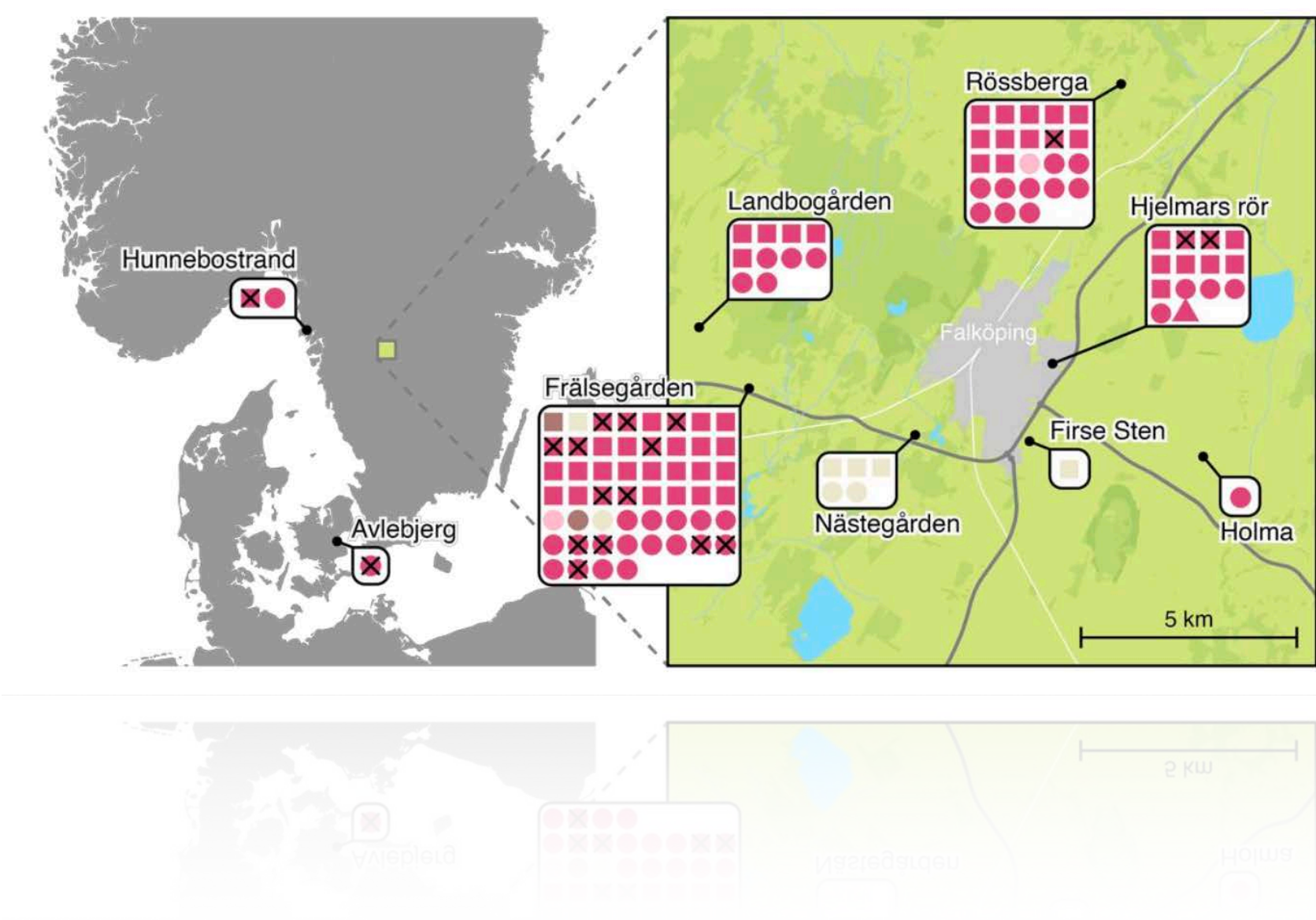
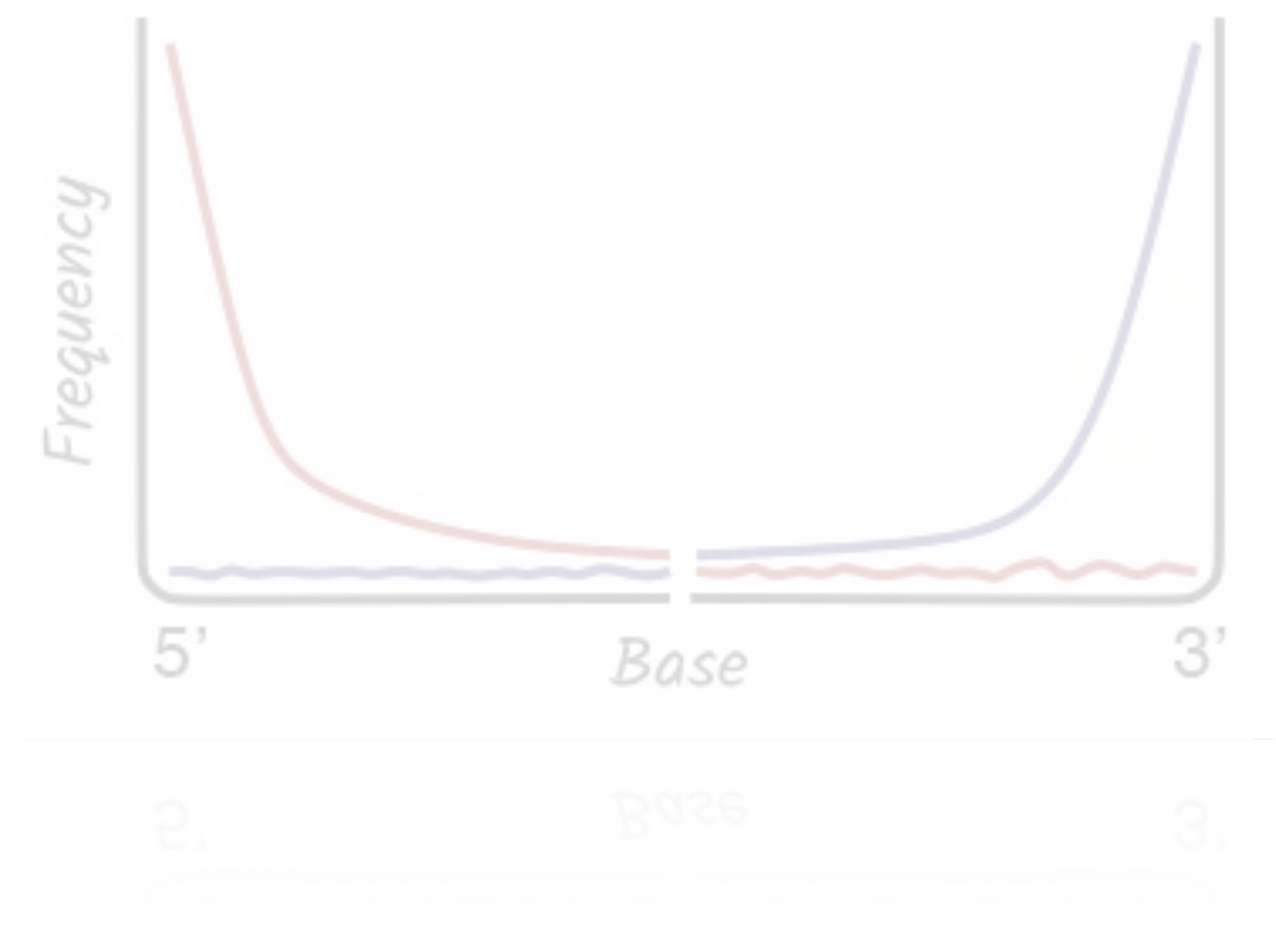


# A PCA tour of western Eurasia





# Topics for today's lecture



**Ancient DNA primer**  
Characteristics and challenges

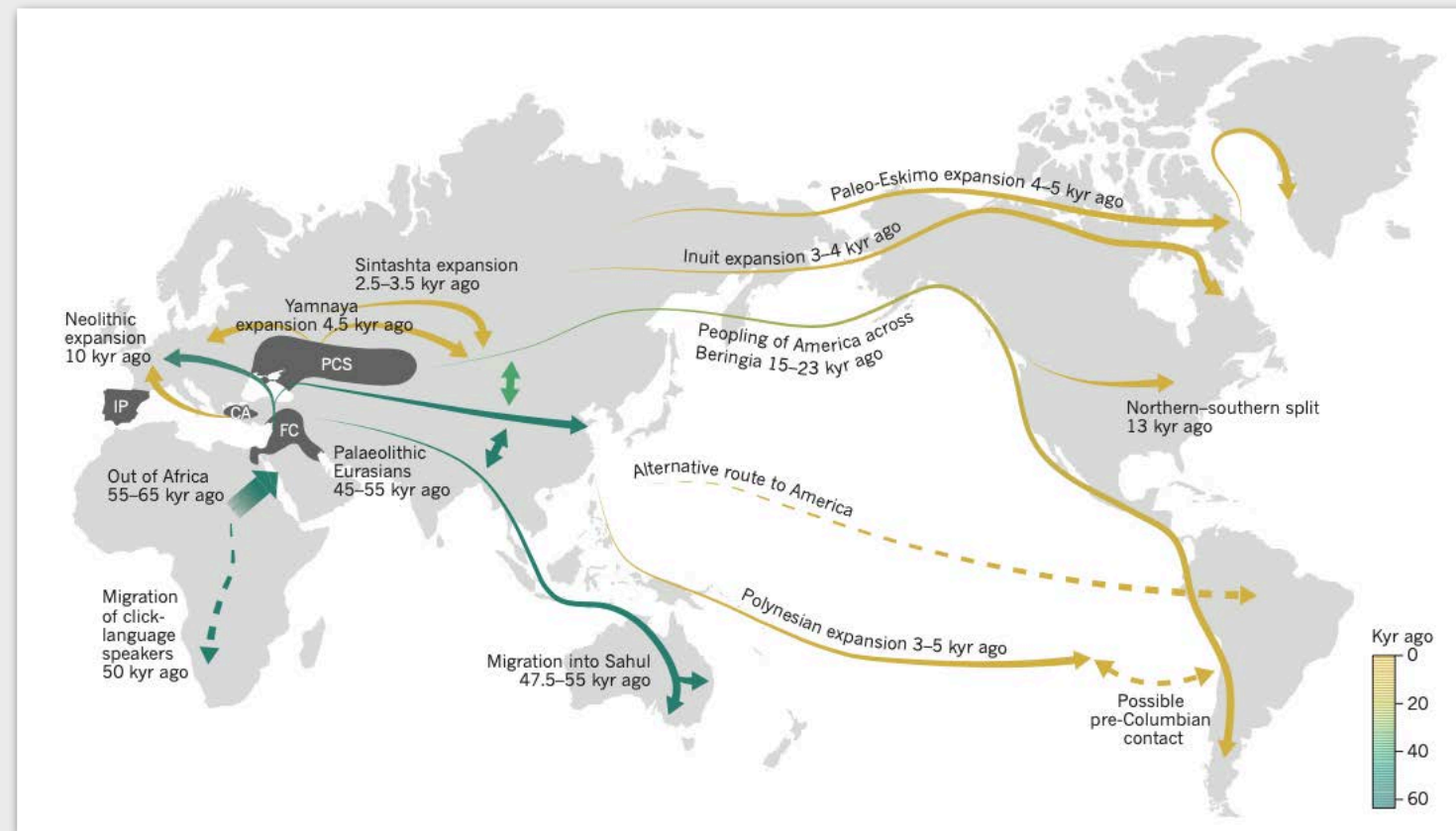
**Ancient DNA showcase**  
Kinship and plague in Stone Age Scandinavia



An abstract graphic consisting of numerous horizontal lines of varying lengths and colors. The colors include blue, orange, and grey. The lines are scattered across the page, with some appearing in groups and others in isolation. The overall effect is a dynamic, layered composition.



# Demography



# Social organisation

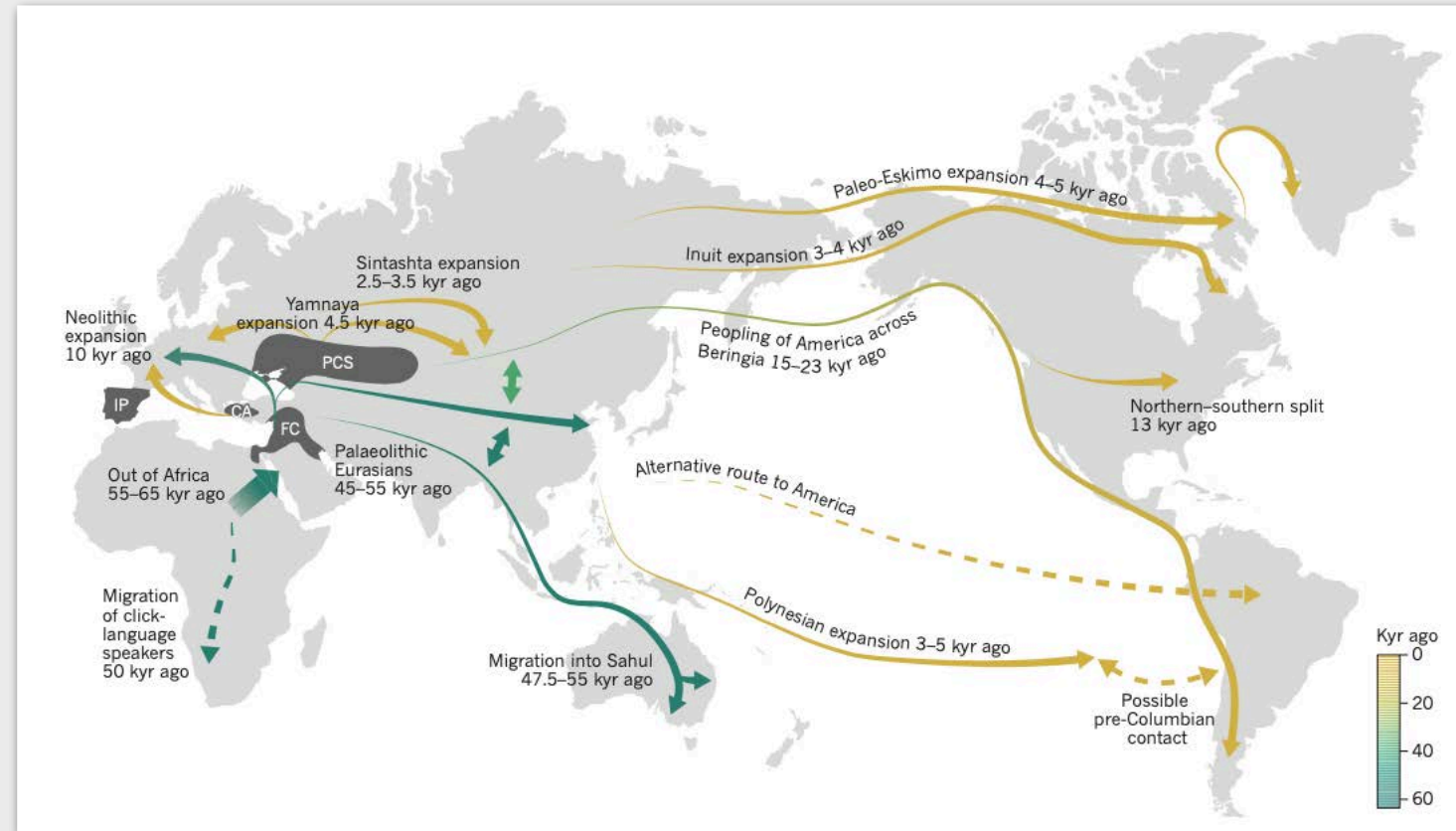


Endogenous host DNA





# Demography



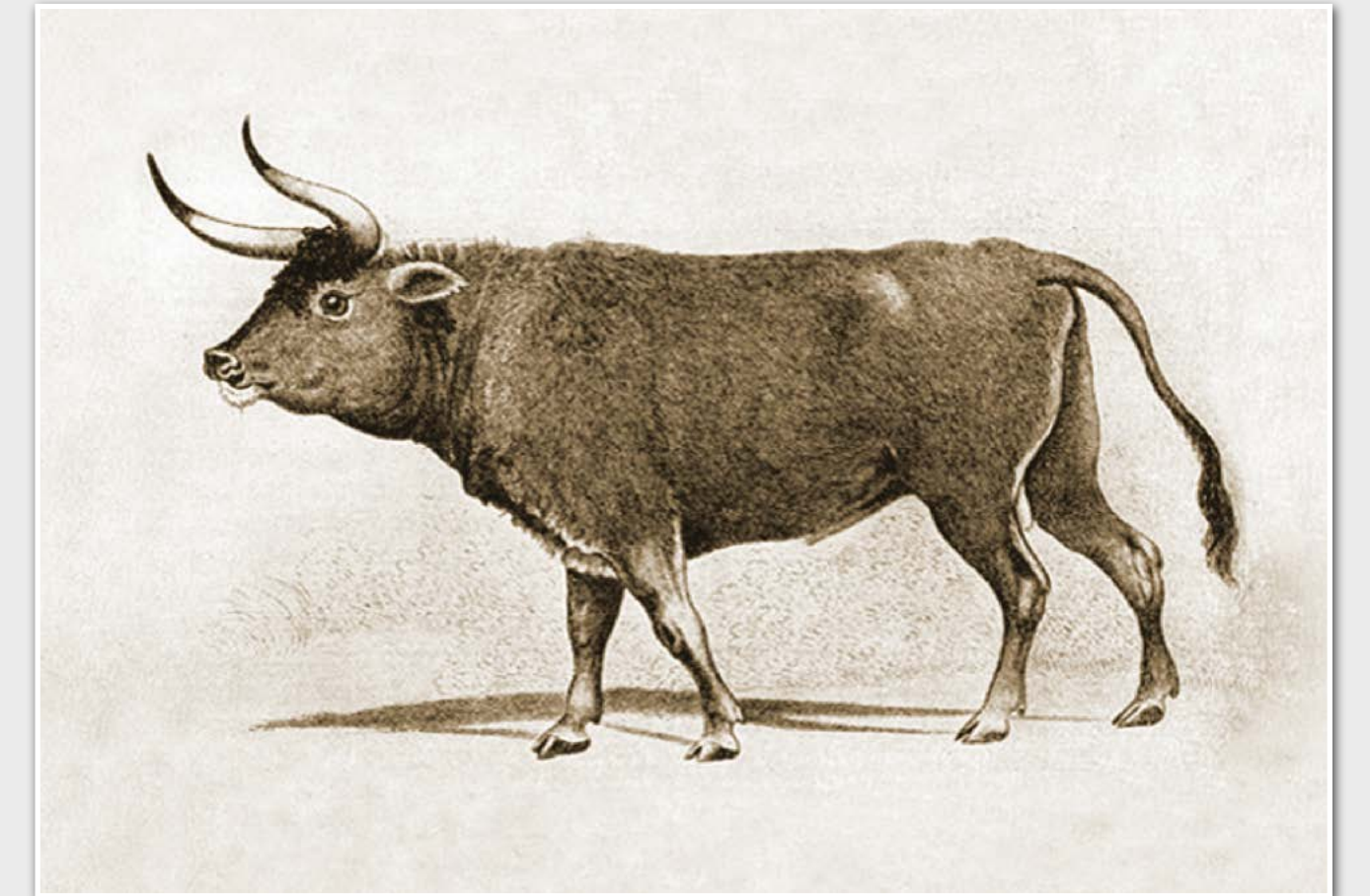
# Social organisation



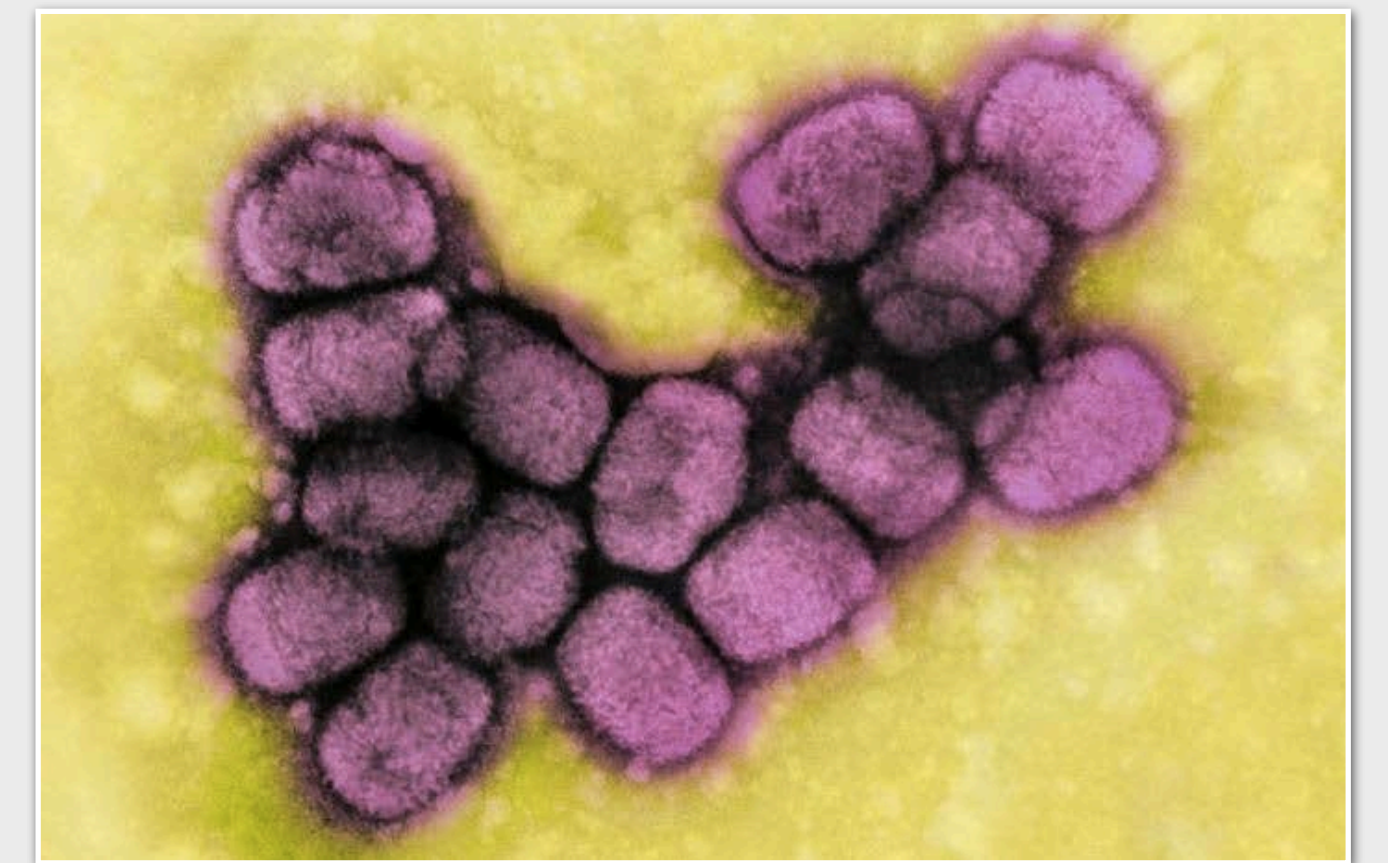
Endogenous host DNA



# Diet and microbiome



# Infectious diseases



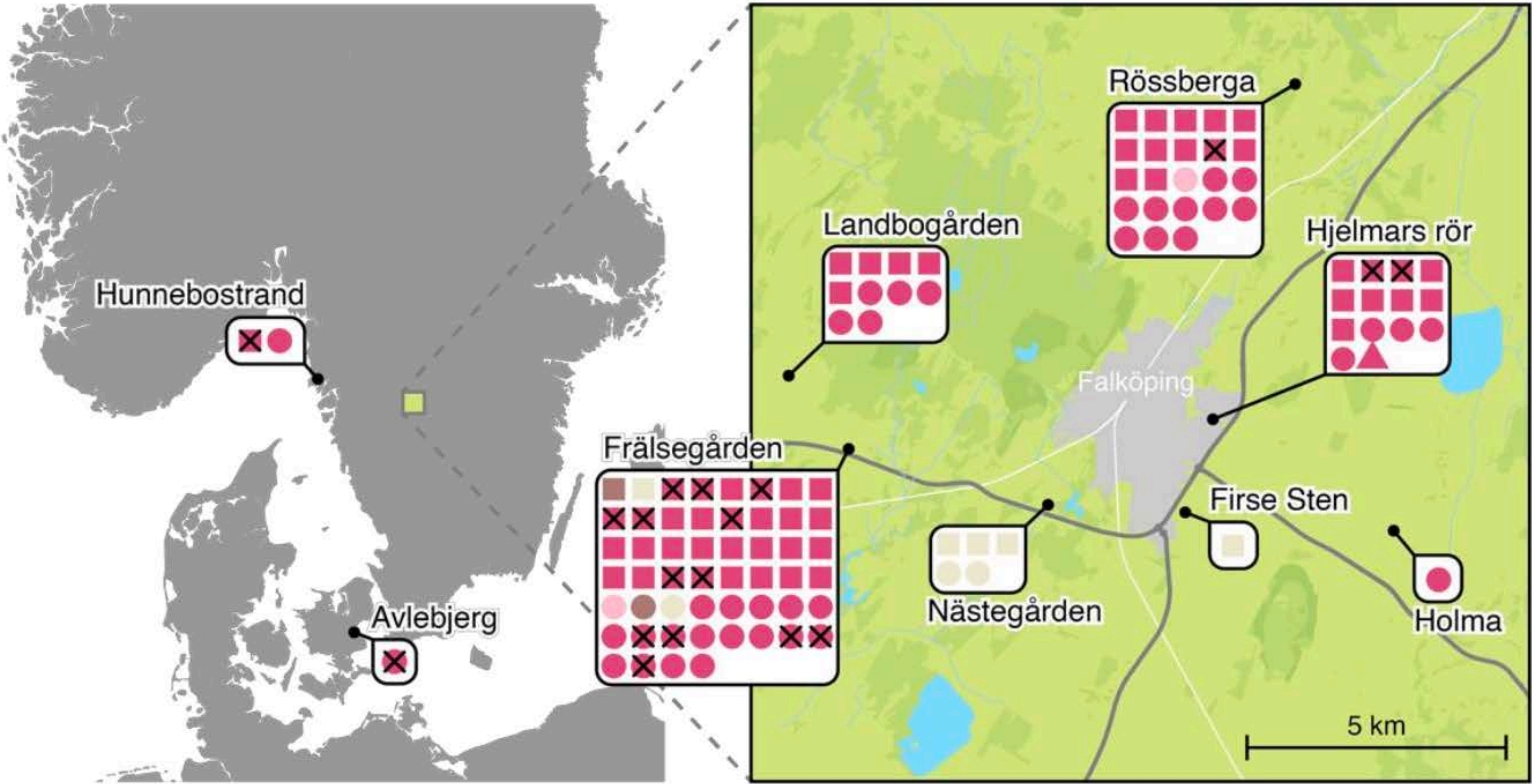
Non-host DNA



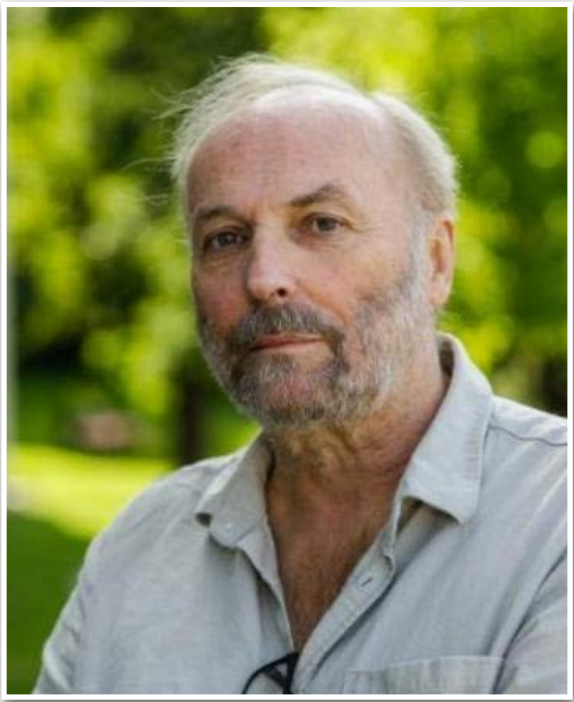




# Population genomics of late Neolithic Scandinavia



Frederik Seersholm

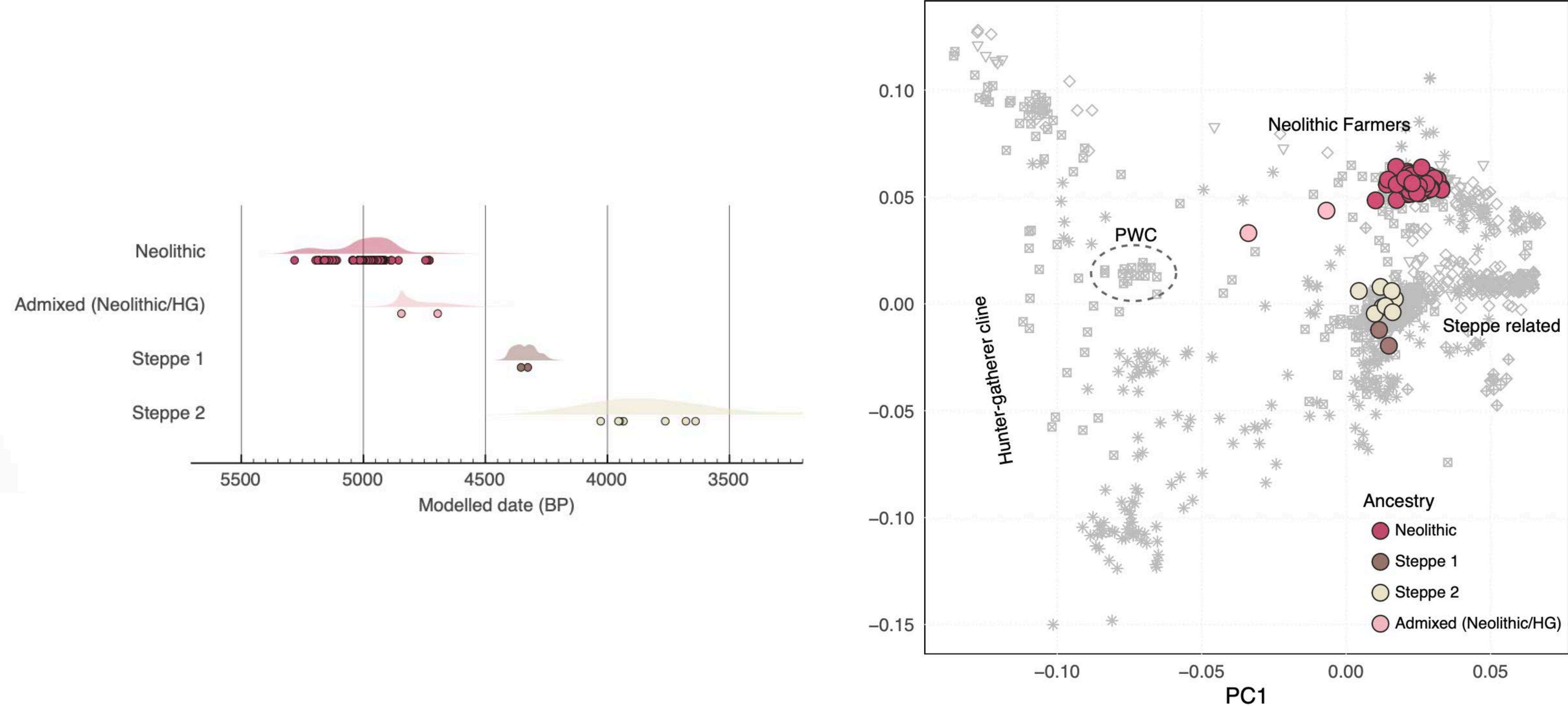


Karl-Göran Sjögren

109 ancient human genomes from megalithic graves in southern Scandinavia

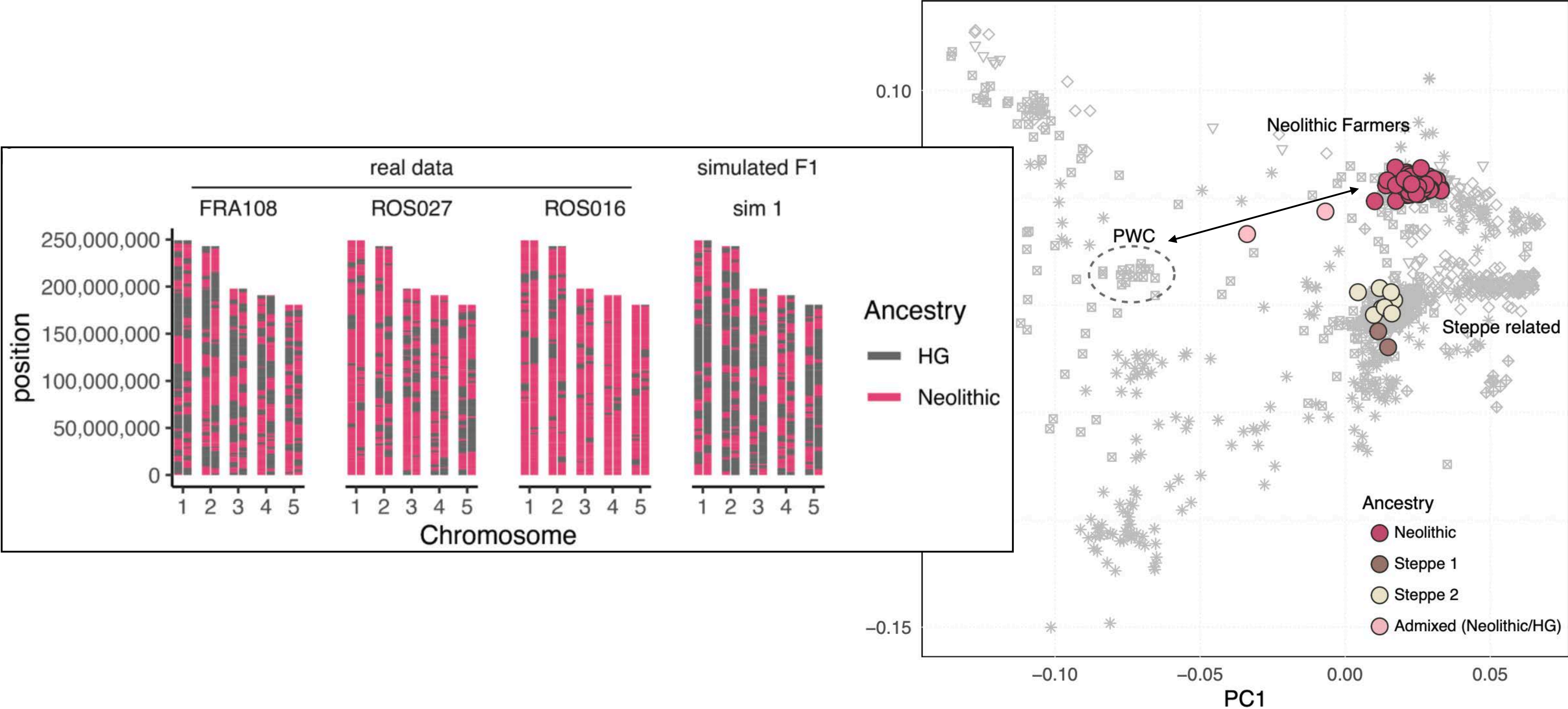


# Genetic diversity of human populations



Repeated use of megalithic graves by peoples with different ancestries

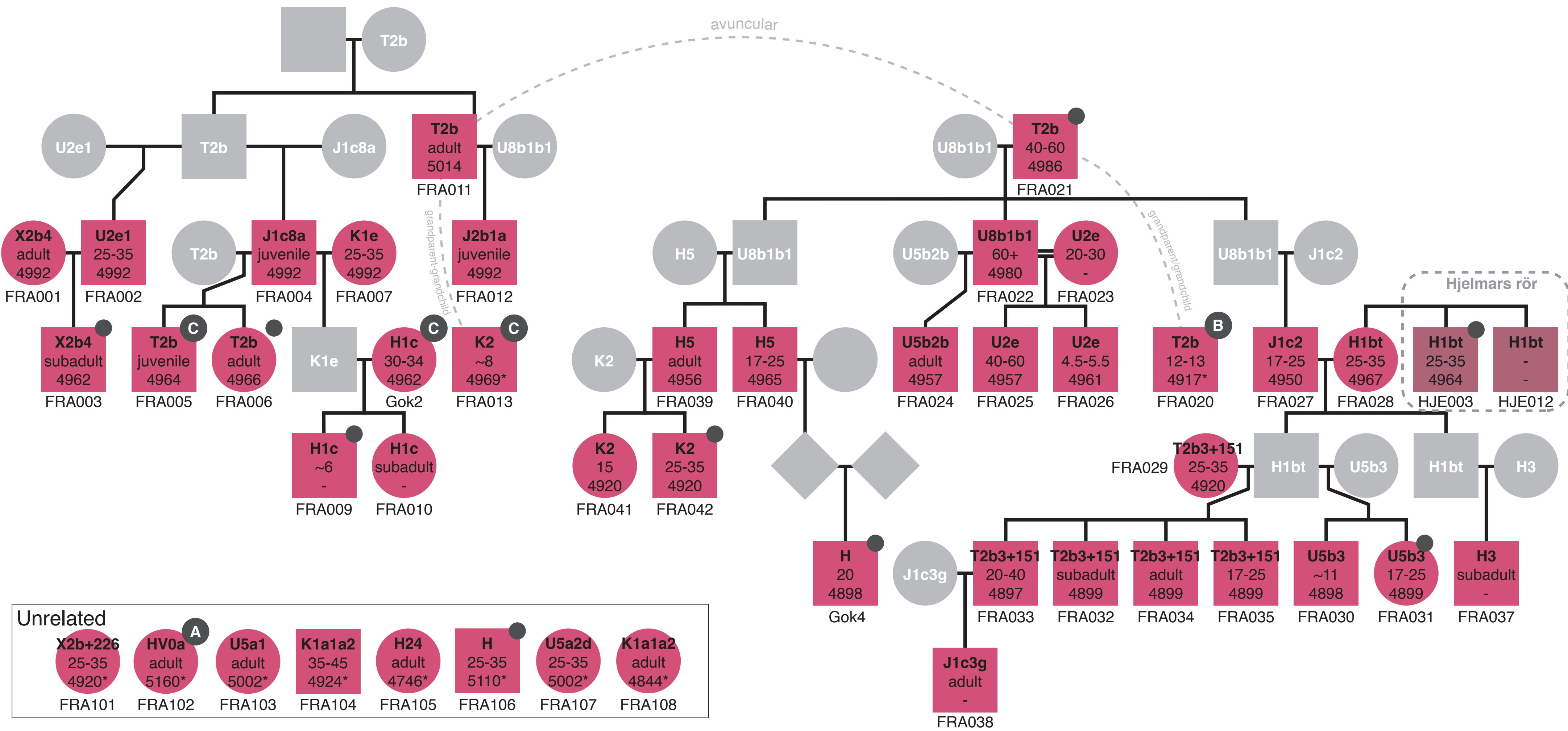
# Recent admixture with Pitted-ware culture hunter-gatherers



Likely first- and second generation offspring at Fralsegarden and Rossberga



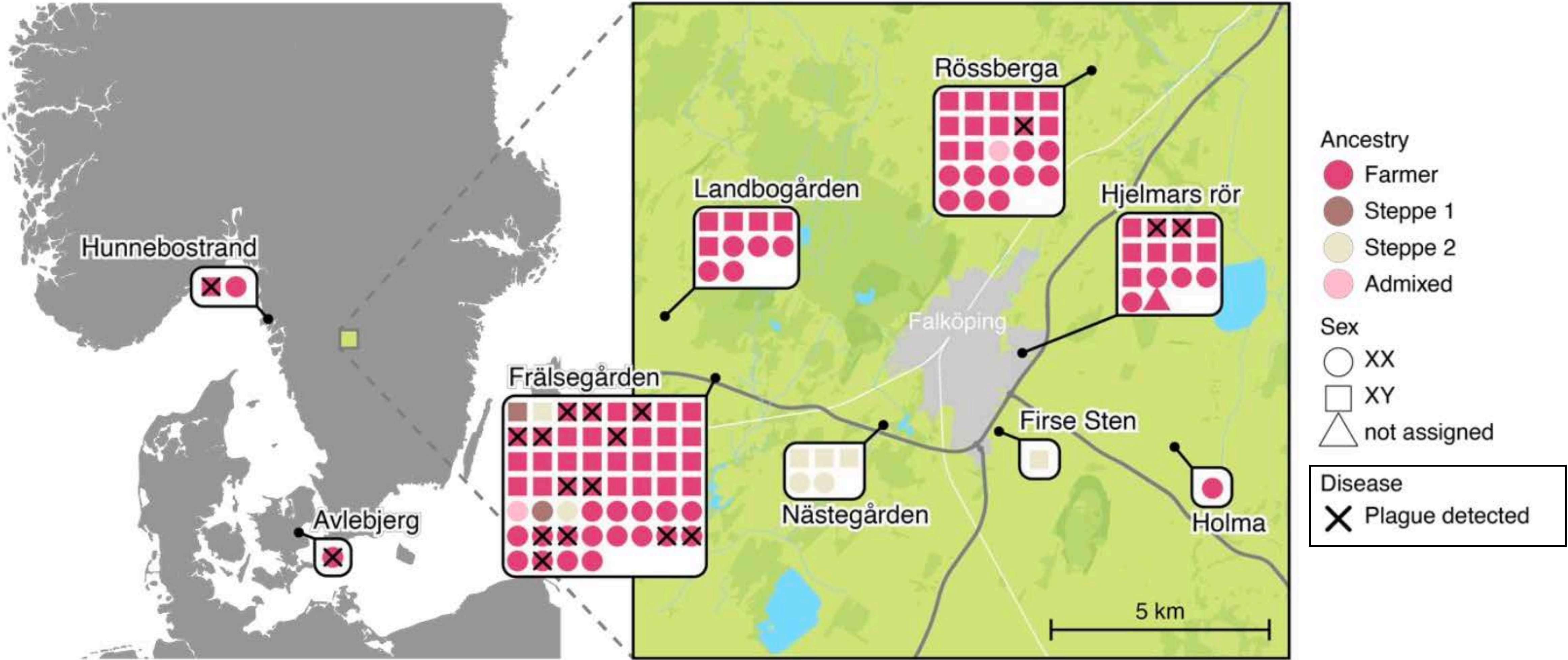
# A five generation pedigree of Neolithic farmers



Pedigree of 38 sequenced individuals suggests patrilineal and patrilocal social organisation



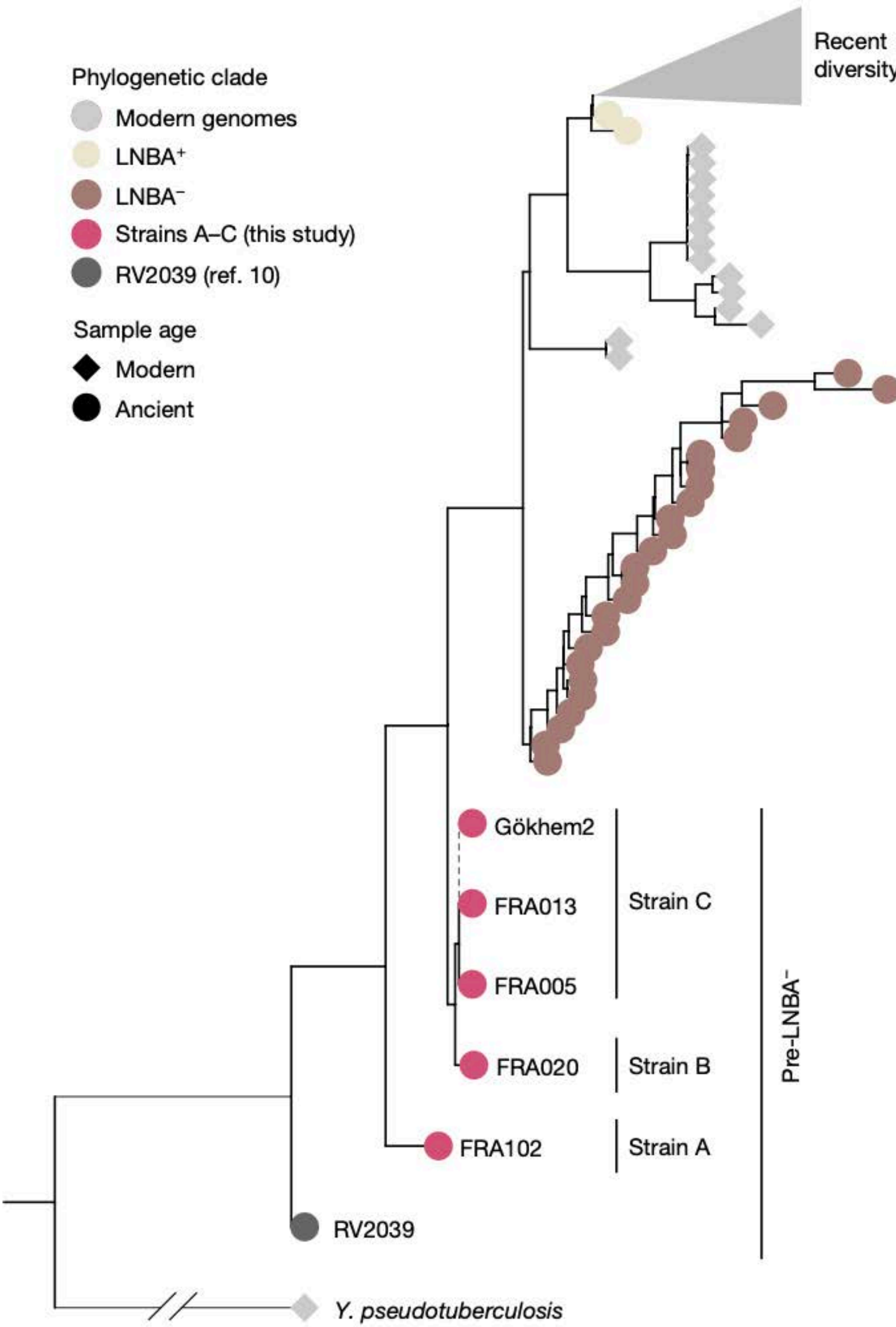
# High prevalence of plague in late Neolithic Scandinavia



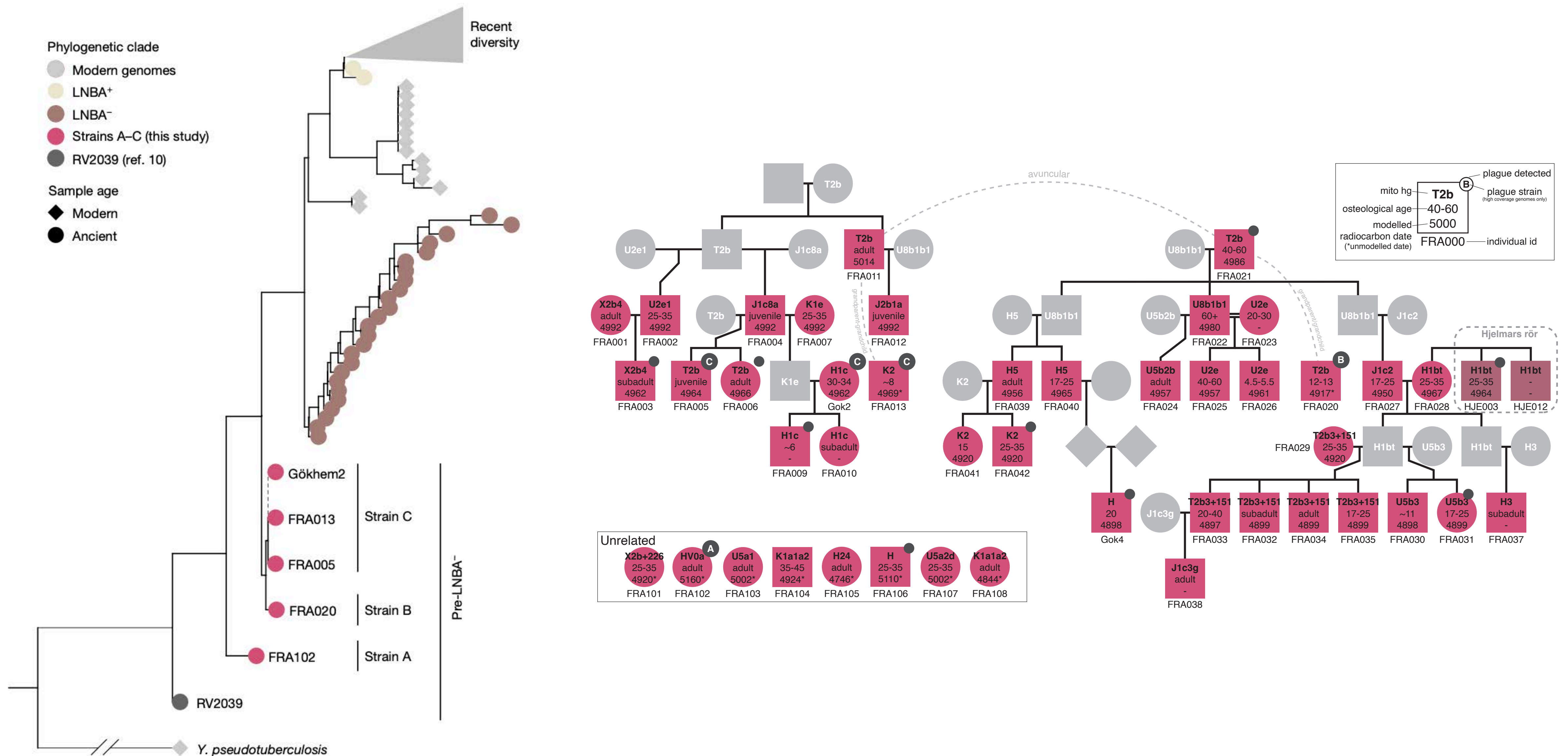
Plague detected in ~17% of individuals



# Multiple plague strains in Neolithic farmer families



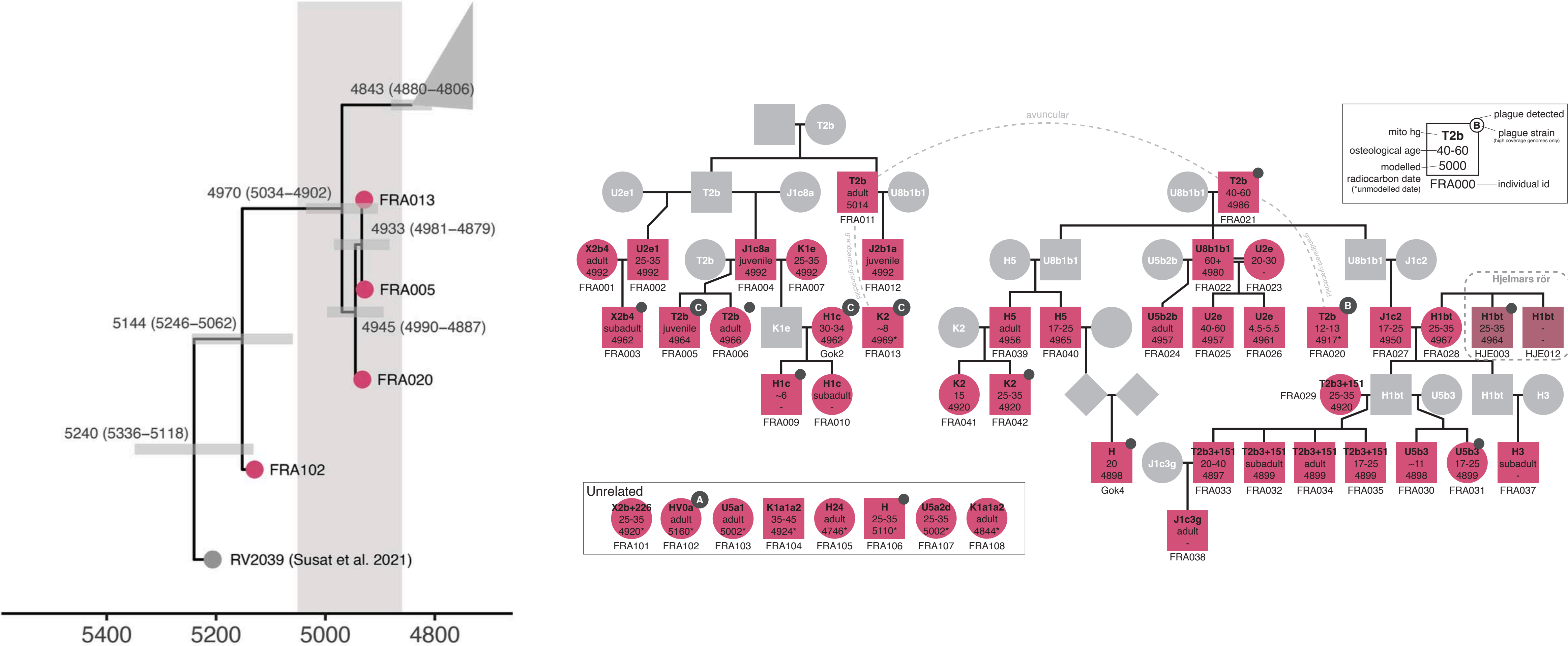
# Multiple plague strains in Neolithic farmer families



## High prevalence of plague in Fralsegarden (28%) and multiple distinct strains

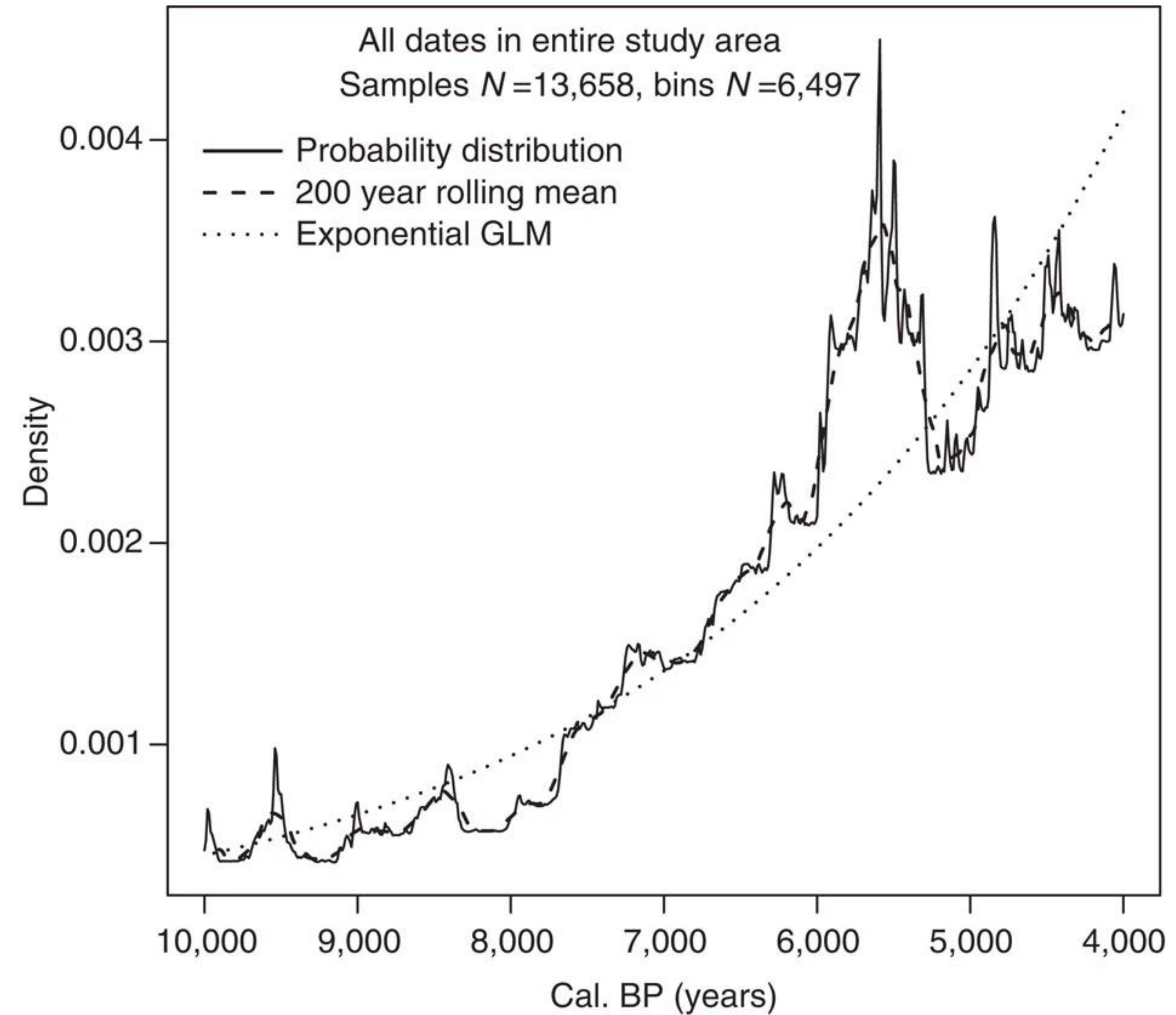


# Multiple plague strains in Neolithic farmer families



Divergence of strains A and B during Fralsegarden pedigree chronology

# Did plague play a role in the Neolithic decline?



Population change in Western European history



# Novel sources for ancient DNA

ARTICLE

<https://doi.org/10.1038/s41467-019-13549-9>




OPEN

## A 5700 year-old human genome and oral microbiome from chewed birch pitch

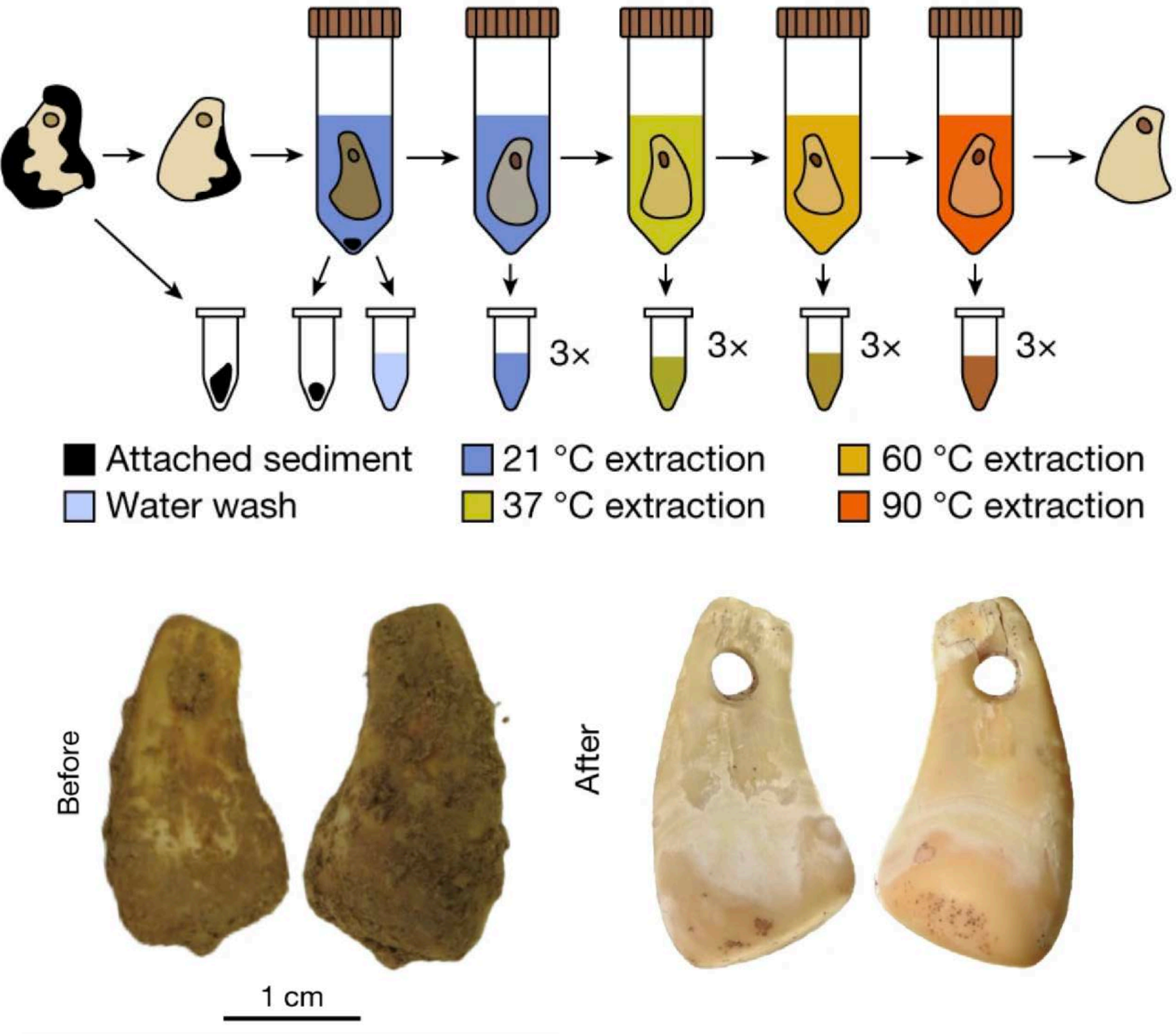
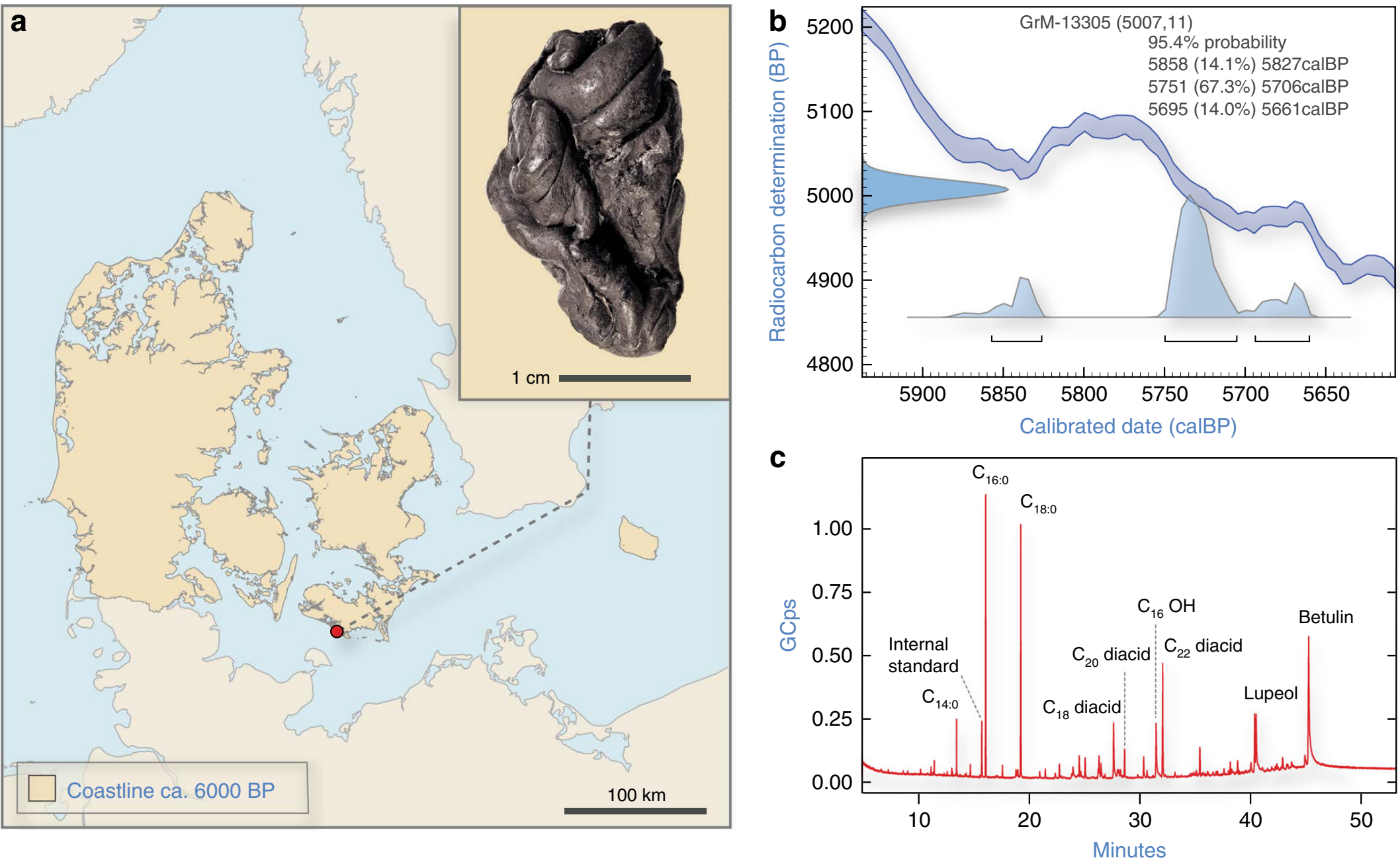
Theis Z.T. Jensen<sup>1,2,10</sup>, Jonas Niemann<sup>1,2,10</sup>, Katrine Højholt Iversen<sup>3,4,10</sup>, Anna K. Fotakis<sup>1</sup>, Shyam Gopalakrishnan<sup>1</sup>, Åshild J. Vågene<sup>1</sup>, Mikkel Winther Pedersen<sup>1</sup>, Mikkel-Holger S. Sinding<sup>1</sup>, Martin R. Ellegaard<sup>1</sup>, Morten E. Allentoft<sup>1</sup>, Liam T. Lanigan<sup>1</sup>, Alberto J. Taurozzi<sup>1</sup>, Sofie Holtsmark Nielsen<sup>1</sup>, Michael W. Dee<sup>5</sup>, Martin N. Mortensen<sup>6</sup>, Mads C. Christensen<sup>6</sup>, Søren A. Sørensen<sup>7</sup>, Matthew J. Collins<sup>1,8</sup>, M. Thomas P. Gilbert<sup>1,9</sup>, Martin Sikora<sup>1</sup>, Simon Rasmussen<sup>4</sup> & Hannes Schroeder<sup>1\*</sup>

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## Ancient human DNA recovered from a Palaeolithic pendant

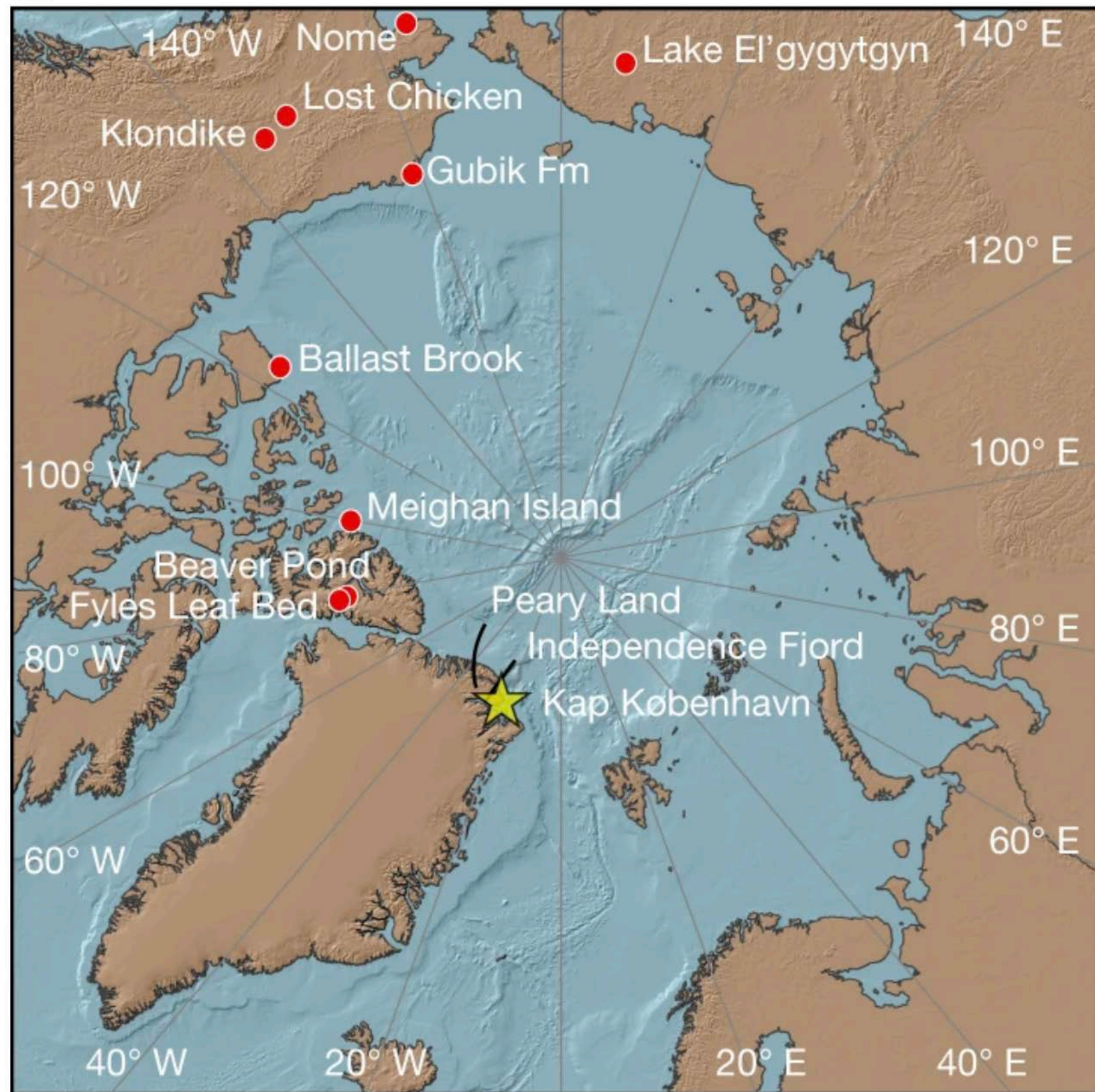
[Elena Essel](#) , [Elena I. Zavala](#), [Ellen Schulz-Kornas](#), [Maxim B. Kozlikin](#), [Helen Fewlass](#), [Benjamin Vernot](#), [Michael V. Shunkov](#), [Anatoly P. Derevianko](#), [Katerina Douka](#), [Ian Barnes](#), [Marie-Cécile Soulier](#), [Anna Schmidt](#), [Merlin Szymanski](#), [Tsenka Tsanova](#), [Nikolay Sirakov](#), [Elena Endarova](#), [Shannon P. McPherron](#), [Jean-Jacques Hublin](#), [Janet Kelso](#), [Svante Pääbo](#), [Mateja Hajdinjak](#), [Marie Soressi](#)  & [Matthias Meyer](#) 

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# Ancient environmental DNA - the next revolution



Reconstruction of a 2 million year-old ecosystem from ancient environmental DNA



# Ancient DNA

Next Generation Sequencing Analysis

DTU, 9/1/2026

Interested in a project?  
[martin.sikora@sund.ku.dk](mailto:martin.sikora@sund.ku.dk)