

Ancient DNA

Next Generation Sequencing Analysis

DTU, 9/1/2026



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Globe Institute

University of Copenhagen

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1984 - the birth of ancient DNA

NATURE VOL. 312 15 NOVEMBER 1984

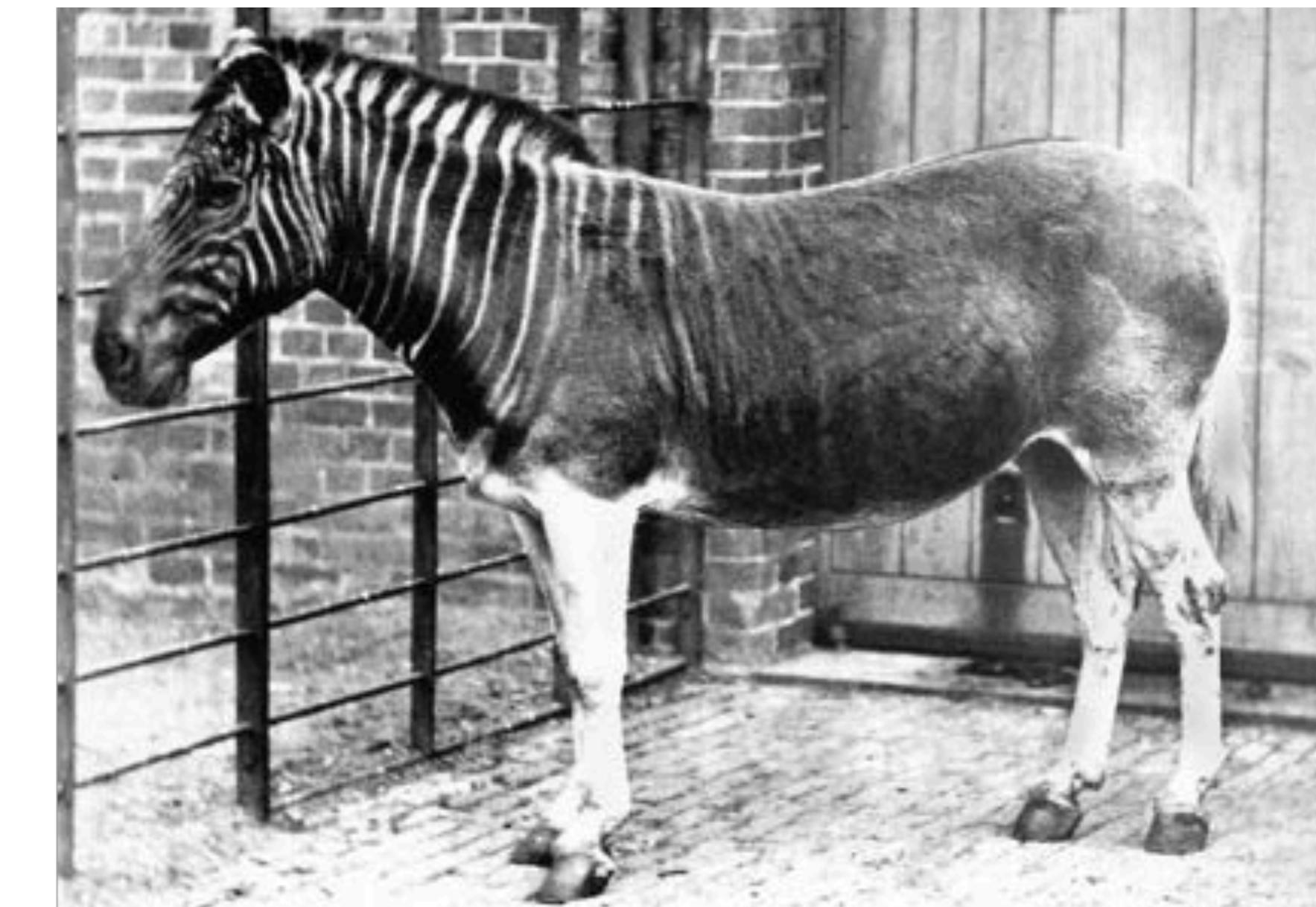
DNA sequences from the quagga, an extinct member of the horse family

Russell Higuchi*, Barbara Bowman*, Mary Freiberger*,
Oliver A. Ryder† & Allan C. Wilson*

* Department of Biochemistry, University of California, Berkeley,
California 94720, USA

† Research Department, San Diego Zoo, San Diego,
California 92103, USA

To determine whether DNA survives and can be recovered from the remains of extinct creatures, we have examined dried muscle from a museum specimen of the quagga, a zebra-like species (*Equus quagga*) that became extinct in 1883 (ref. 1). We report that DNA



Unidentified reading frame 1

Quagga	C CCA ATC CTG CTC GCC GTC GCA TTC CTC ACA CTA GTT GAA CGA AAA GTC TTA GGC TAC ATA CAA CTT CGT AAA GGA CCC AAC ATC GTC GGC CCC TAT GGC CTA CTA CAA CCC ATT AC
ZebraT.....G.....T.....C.....G*

Cytochrome oxidase I

Quagga	A GGA GGA TTC GTT CAC TGA TTC CCT CTA TTC TCA GGA TAC ACA CTC AAC CAA ACC TGA GCA AAA ATT CAC TTT ACA ATT ATA TTC GTC AAC ATA ATT TTC TTC CCA
Zebra	G.....T.....G.....C.....A.....T.....C*

Fig. 1 Sequences of the coding strands determined for two pieces of quagga mtDNA. The sequences are arranged in triplets corresponding to the amino acids that they encode. At 12 positions, the quagga sequences differ from those of mtDNA from a mountain zebra; only for these positions is the nature of the base specified for the zebra. The two asterisks identify triplets at which the zebra and quagga differ by an amino acid replacement.

Nobel prize in Medicine 2022

The Nobel Prize in Physiology or
Medicine 2022

Svante Pääbo

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Svante Pääbo Facts



III. Niklas Elmehed © Nobel
Prize Outreach

Svante Pääbo

The Nobel Prize in Physiology or Medicine 2022

Born: 20 April 1955, Stockholm, Sweden

Affiliation at the time of the award: Max Planck Institute for
Evolutionary Anthropology, Leipzig, Germany, Okinawa
Institute of Science and Technology, Okinawa, Japan

Prize motivation: “for his discoveries concerning the
genomes of extinct hominins and human evolution”

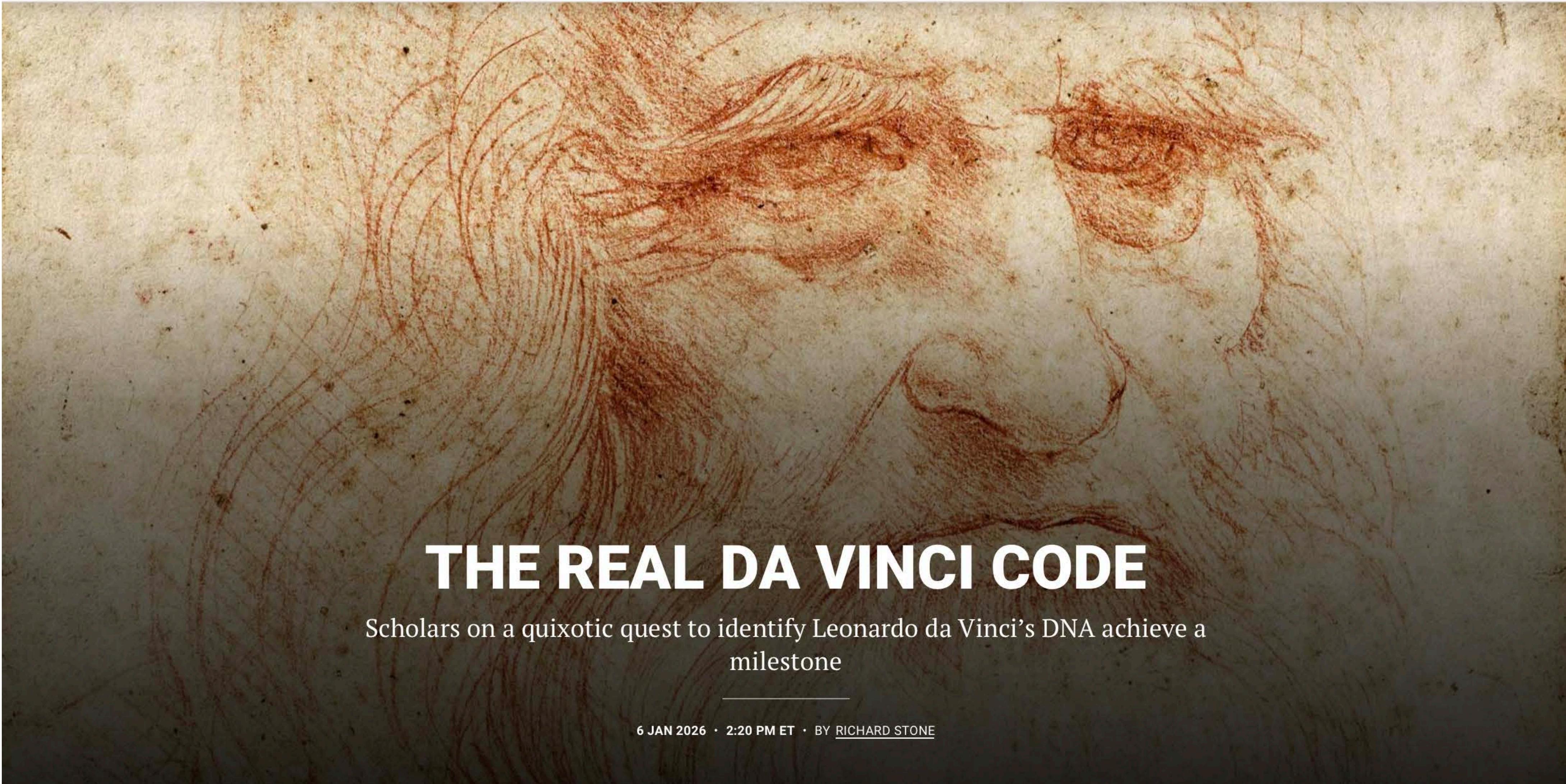
Prize share: 1/1

The week in ancient DNA

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THE REAL DA VINCI CODE

Scholars on a quixotic quest to identify Leonardo da Vinci's DNA achieve a milestone

6 JAN 2026 • 2:20 PM ET • BY [RICHARD STONE](#)

The week in ancient DNA

New Results

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Biological signatures of history: Examination of composite biomes and Y chromosome analysis from da Vinci-associated cultural artifacts

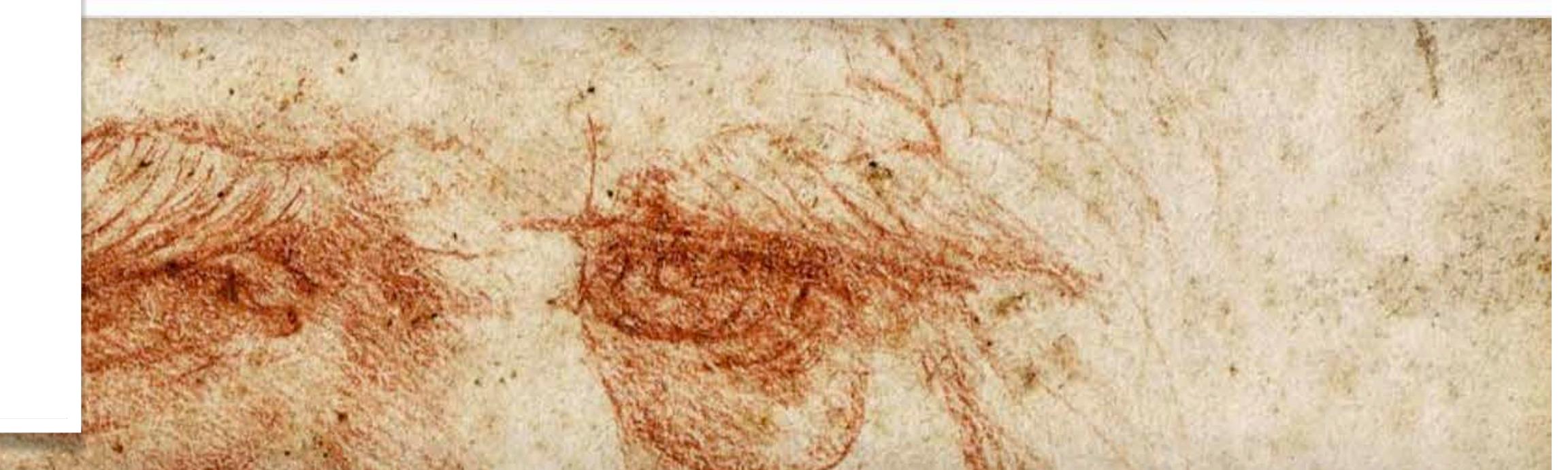
Harinder Singh, Seesandra V. Rajagopala, Rebecca Hart, Pille Hallast, Mark Loftus, Rosana Wiscovitch-Russo, Cody R. K. Conrad, David S. Thaler, Guadalupe Piñar, Karina C. Åberg, Rossella Lorenzi, José A Lorente, Jesse H. Ausubel, Thomas P. Sakmar, Rhonda K. Roby, Charles Lee, Norberto Gonzalez-Juarbe

doi: <https://doi.org/10.64898/2026.01.06.697880>

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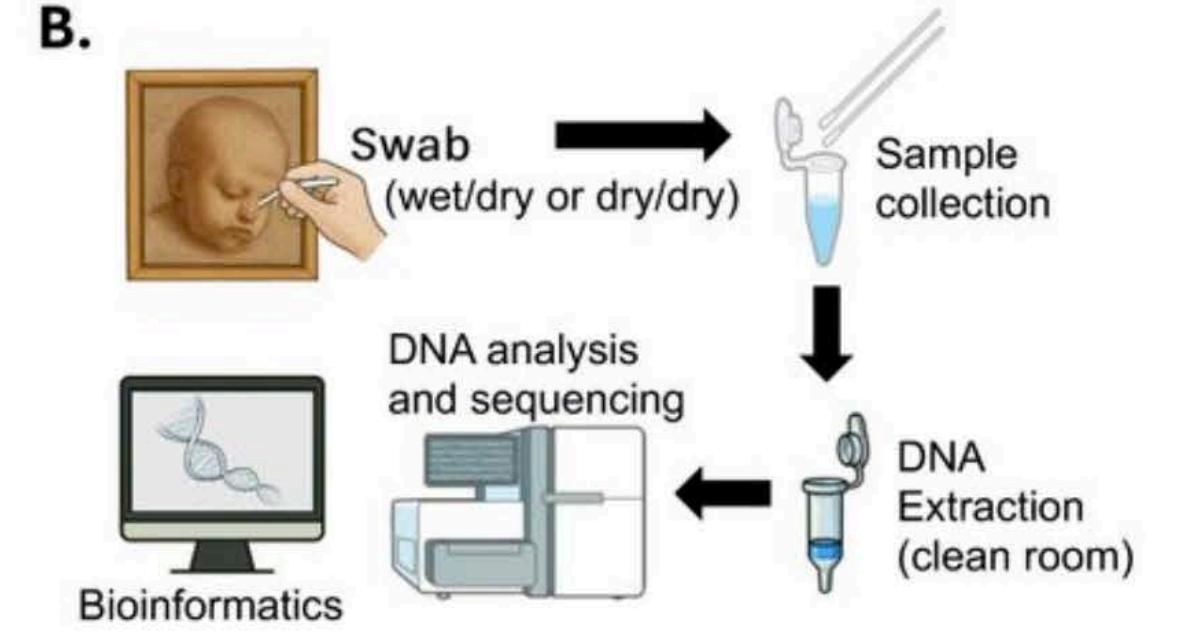
Scho

A.

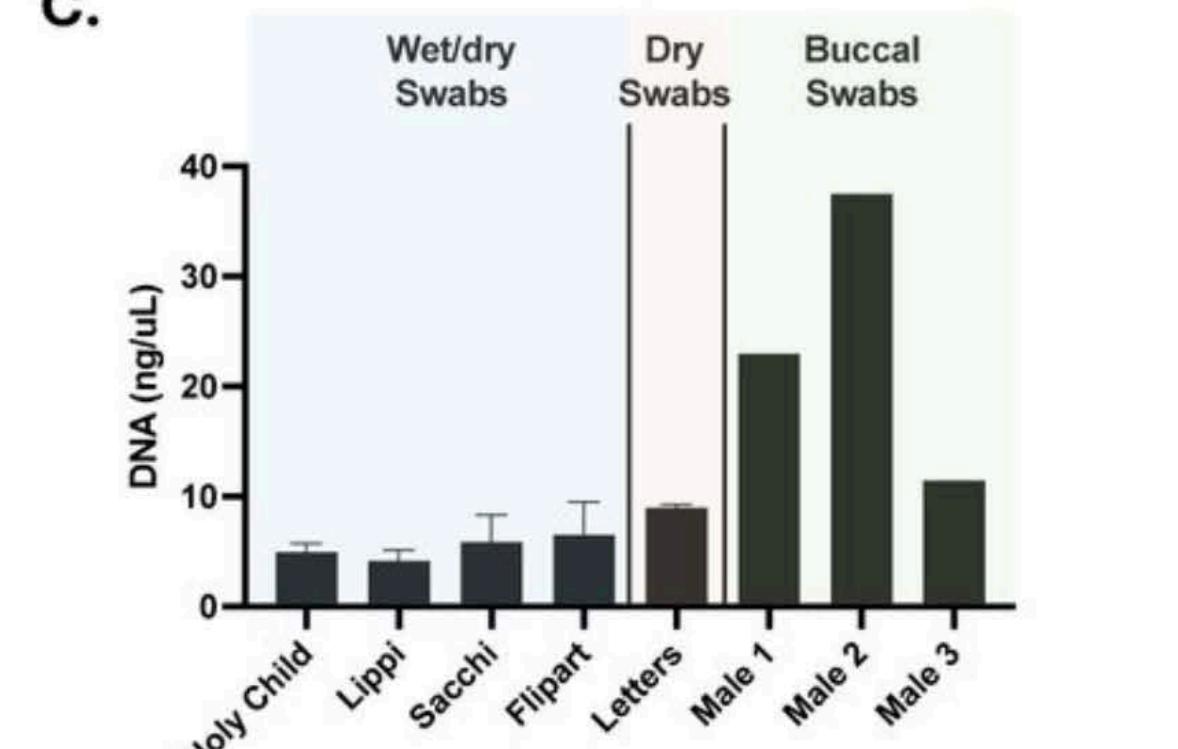


Title or unique ID:	Holy Child	Letters	St. John in a Classical Robe by a Pillar	Man Caught in the Wind	Seated Gentleman at Writing Table
Style:	Red chalk	Handwritten letter	Silver point and ink with blue wash	Red chalk	Red chalk
Paper:	Laid paper	Linen-rag paper	Laid paper	Laid paper	Laid paper
Measurement:	133 x 102 mm	not taken	133 x 102 mm	296 x 212 mm	260 x 172 mm
Sampling Technique:	Double swab (wet/dry)	Dry swab	Double swab (wet/dry)	Double swab (wet/dry)	Double swab (wet/dry)
Country:	Italy	Spain/Italy	Italy	Italy	France
Date or period:	c. 1470 CE	c. 1392-1403 CE	c. 1501 CE	c. 1625 CE	c. 1740 CE
Maker:	Leonardo Da Vinci (1452-1519)	Frosino di Ser Giovanni da Vinci (1400s)	Filippino Lippi (1457-1504)	Andrea Sacchi (1599-1661)	Charles J. Flippart (1721-1796)

B.



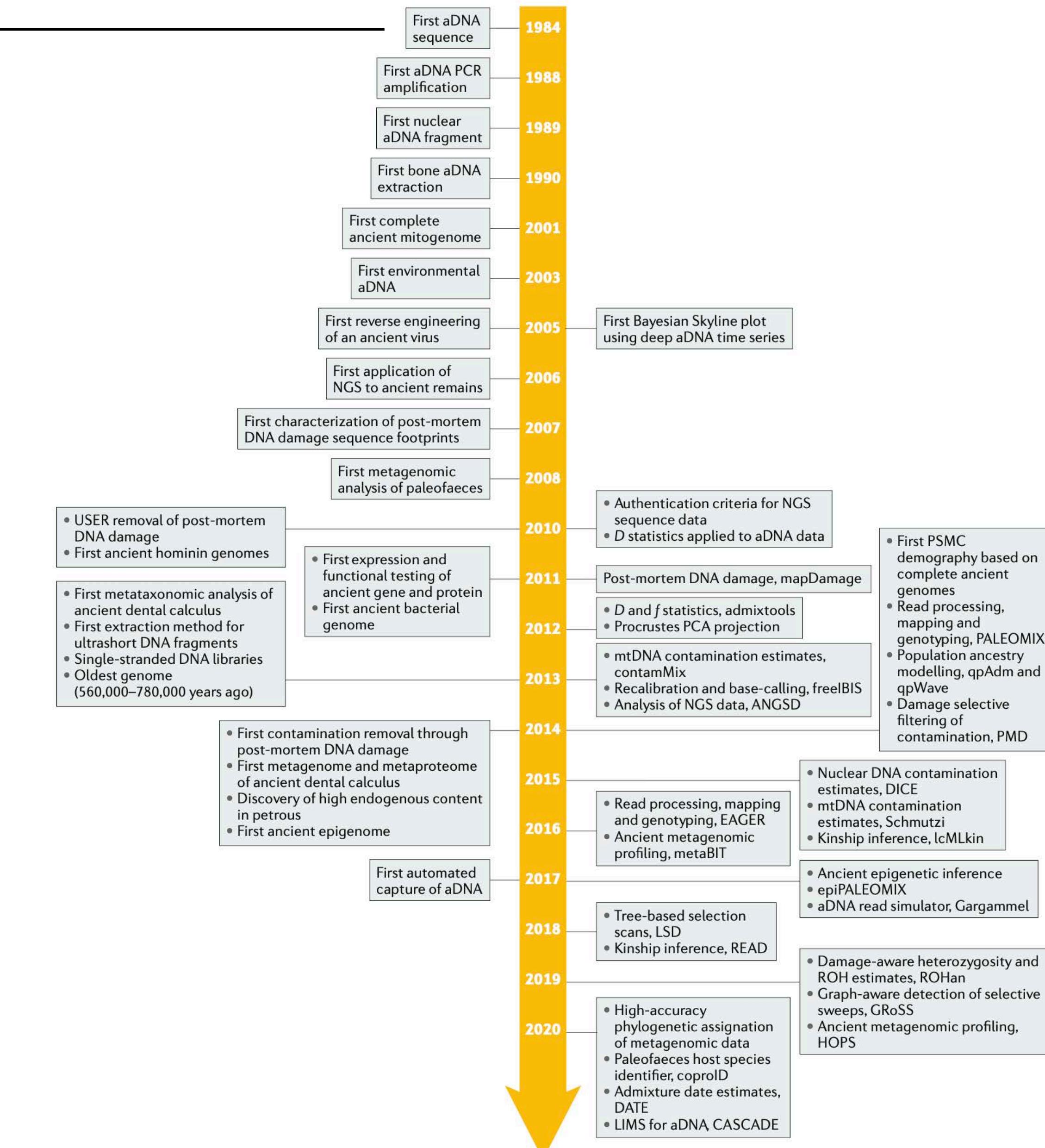
C.



Sample	Swab Type	Approx. DNA (ng/uL)
Holy Child	Wet/dry Swabs	~5
Lippi	Wet/dry Swabs	~4
Sacchi	Wet/dry Swabs	~6
Filippart	Dry Swabs	~7
Letters	Dry Swabs	~10
Male 1	Buccal Swabs	~23
Male 2	Buccal Swabs	~38
Male 3	Buccal Swabs	~12

Milestones in ancient DNA

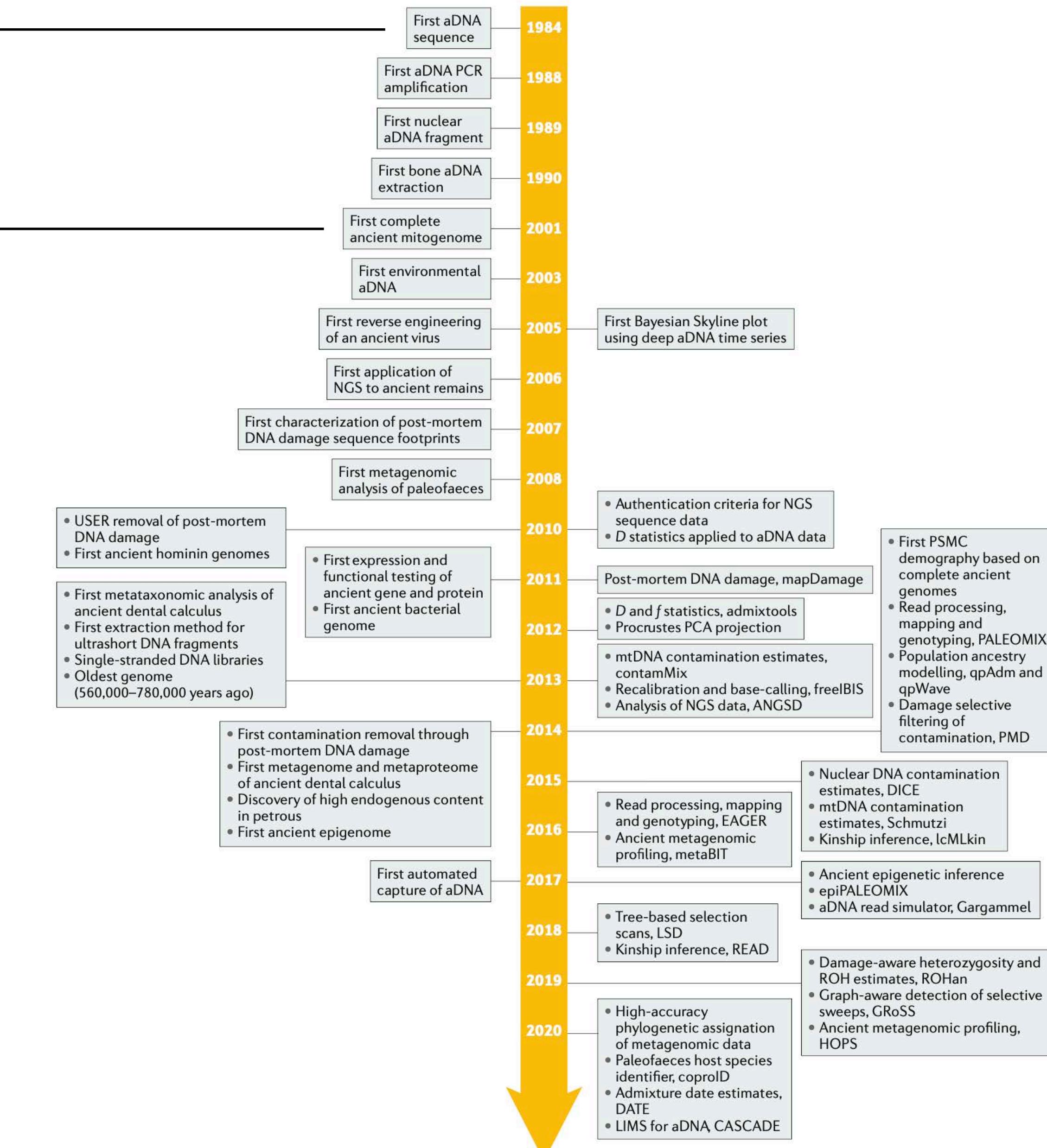
1984 - First aDNA sequence



Milestones in ancient DNA

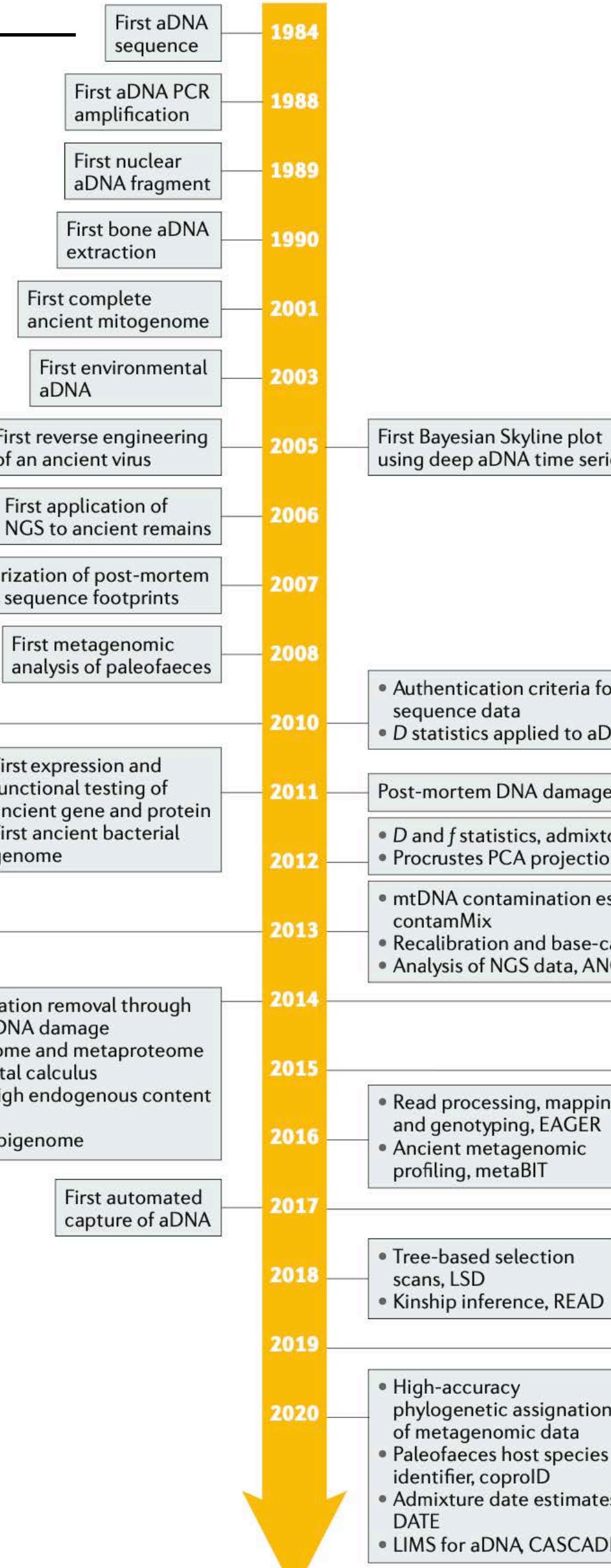
1984 - First aDNA sequence

2001 - First ancient mitogenome



Milestones in ancient DNA

1984 - First aDNA sequence



2001 - First ancient mitogenome

2010 - First ancient human genome

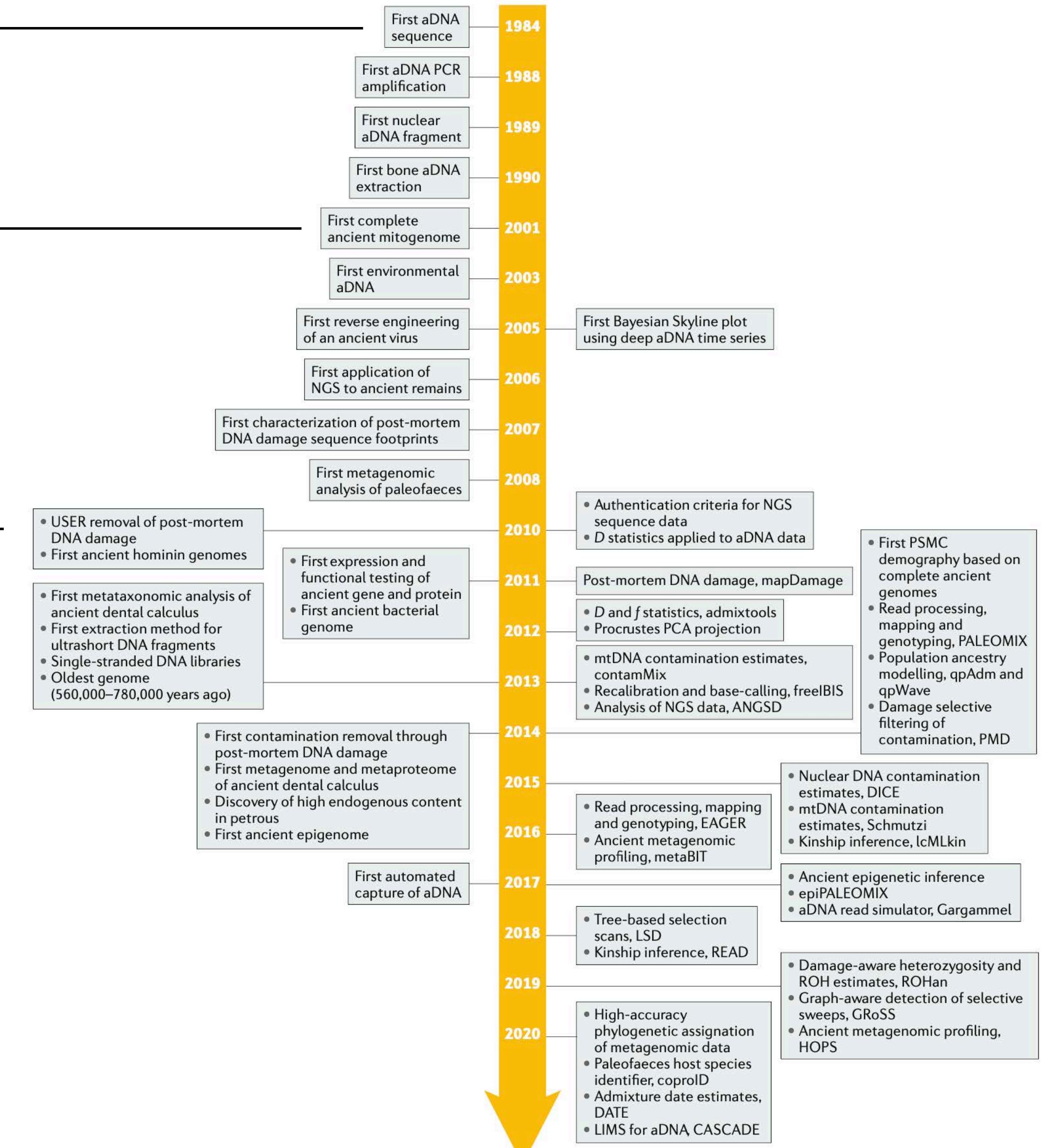
Milestones in ancient DNA

1984 - First aDNA sequence

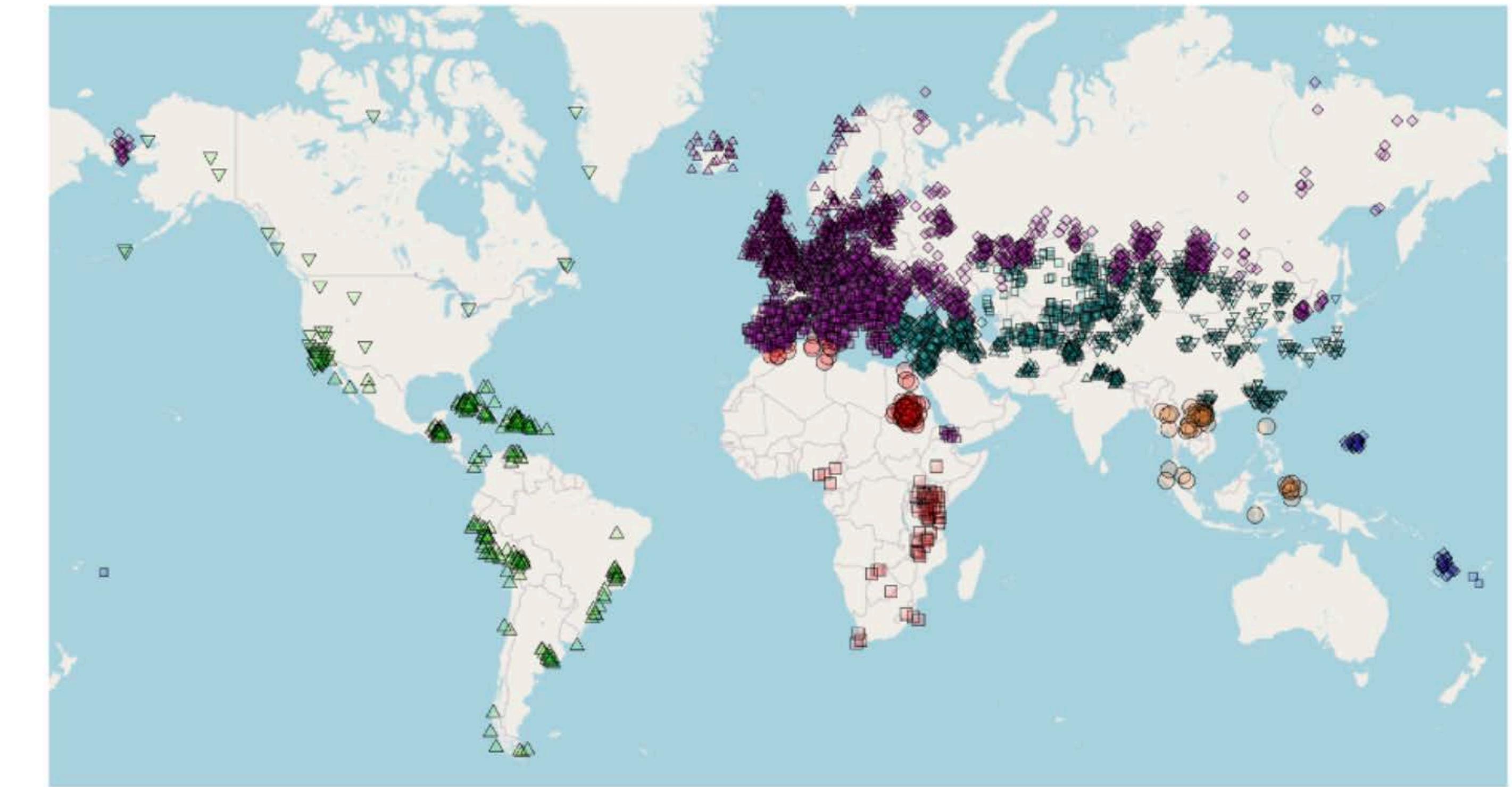
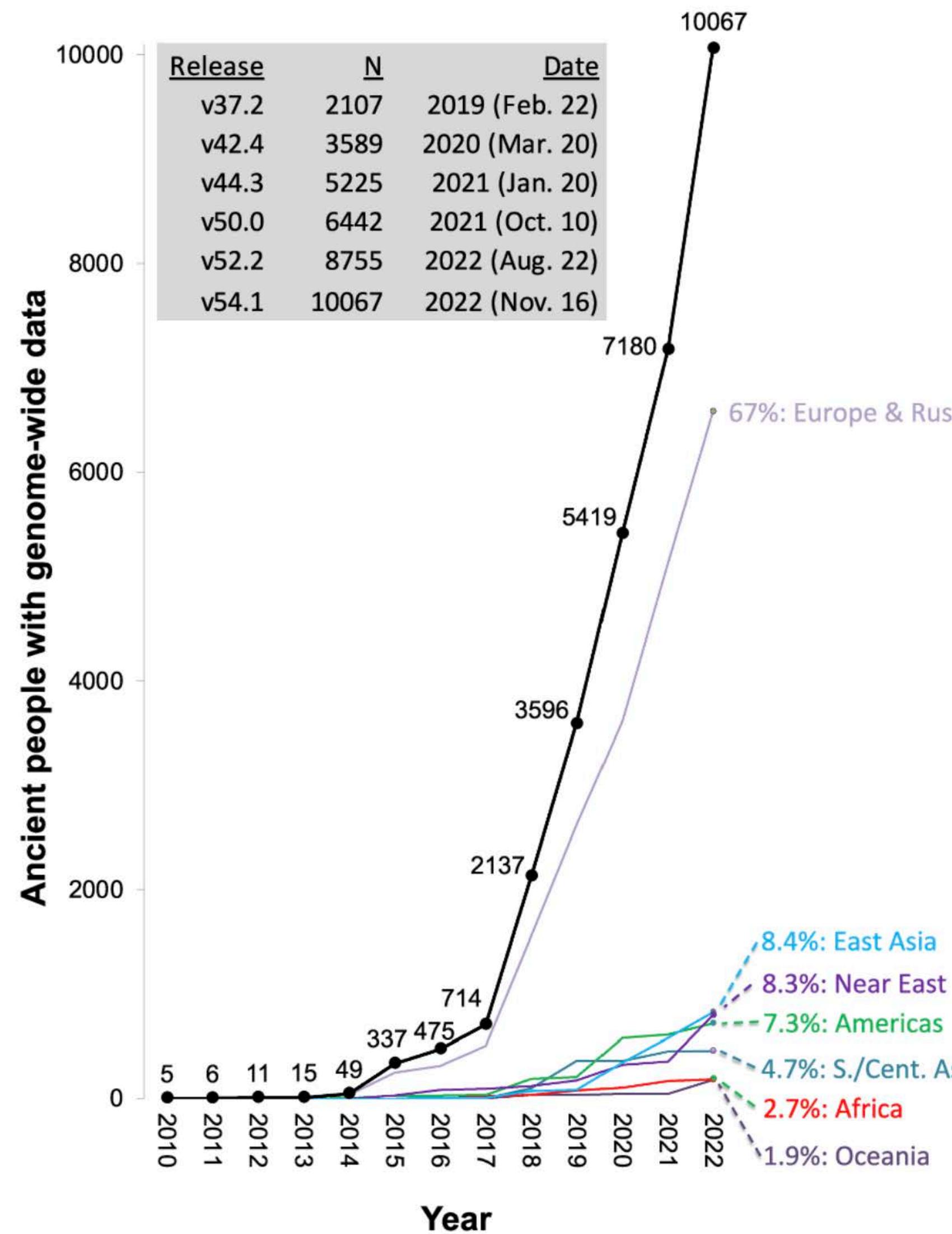
2001 - First ancient mitogenome

2010 - First ancient human genome

The “big data” era of ancient genomics

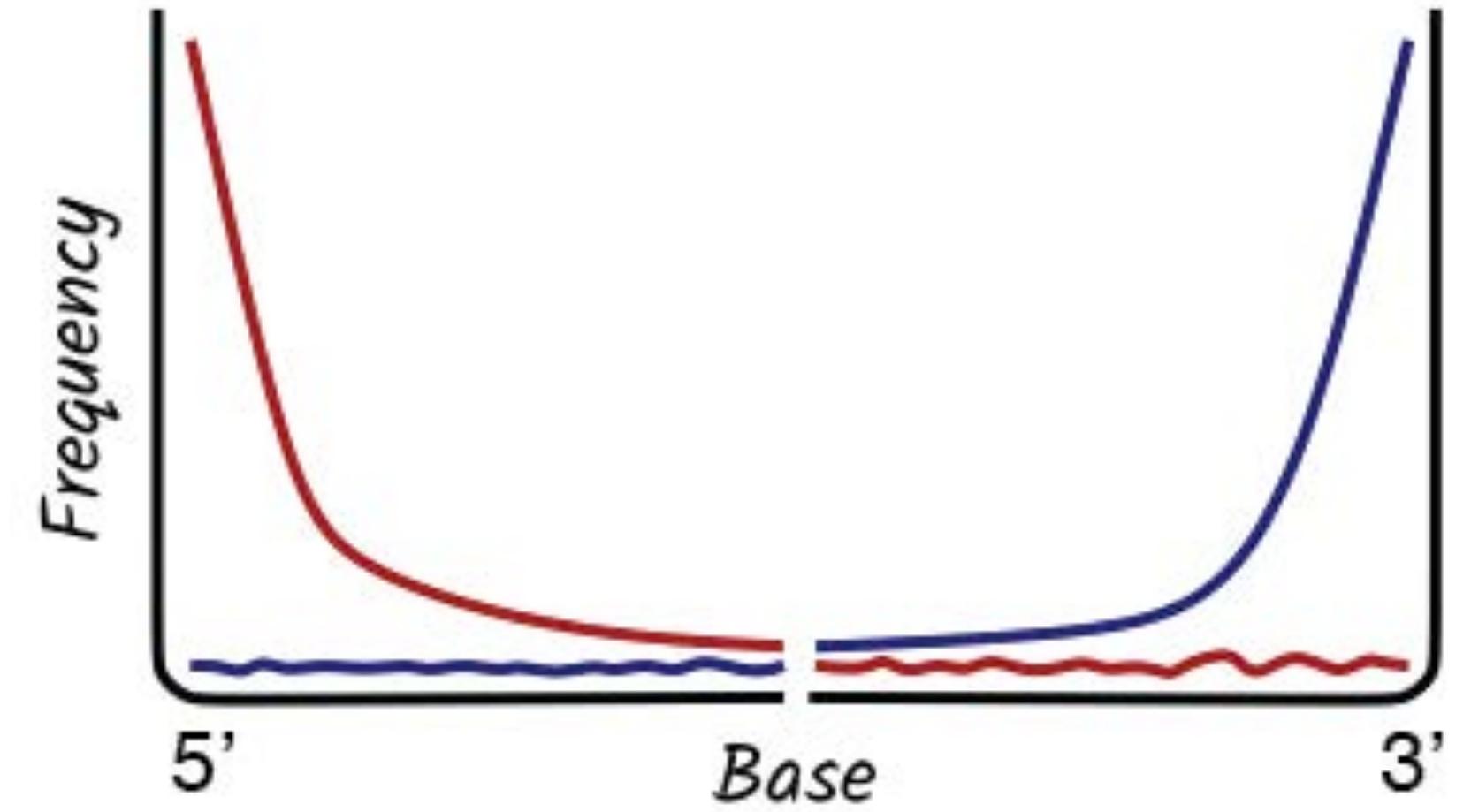


The ancient human DNA revolution



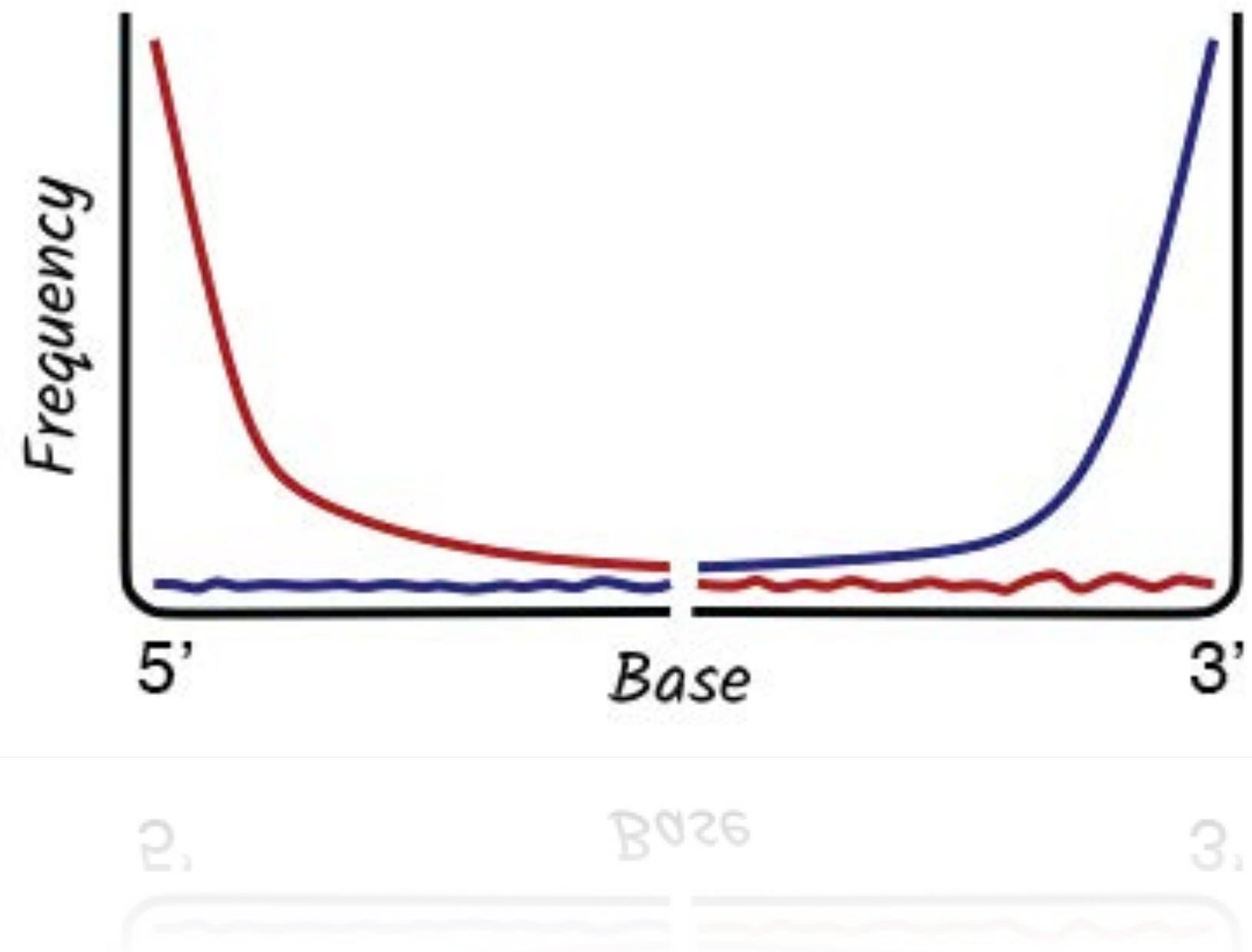
Over 10,000 ancient humans with genome-wide data since 2010

Topics for today's lecture

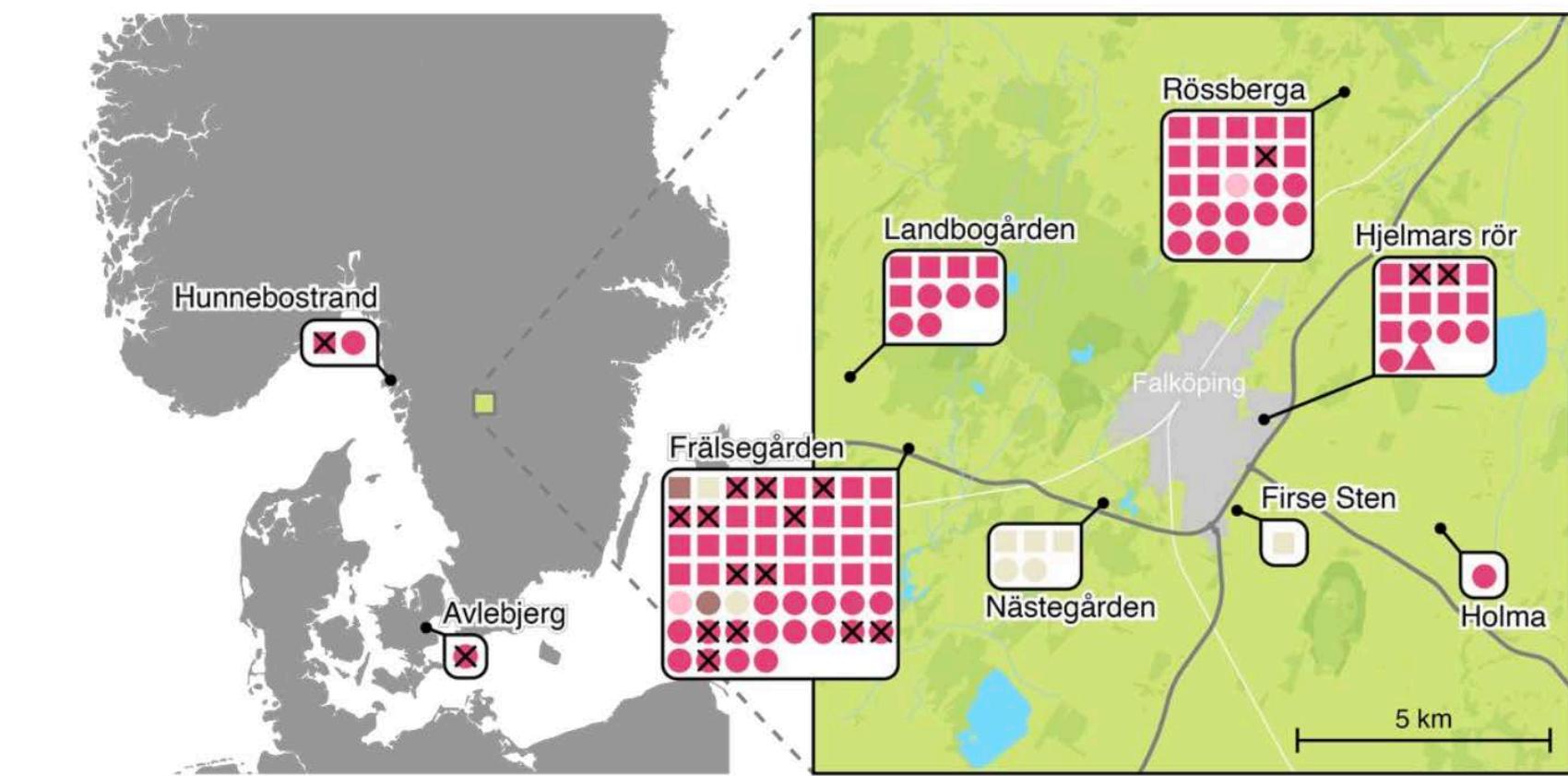


Ancient DNA primer
Characteristics and challenges

Topics for today's lecture

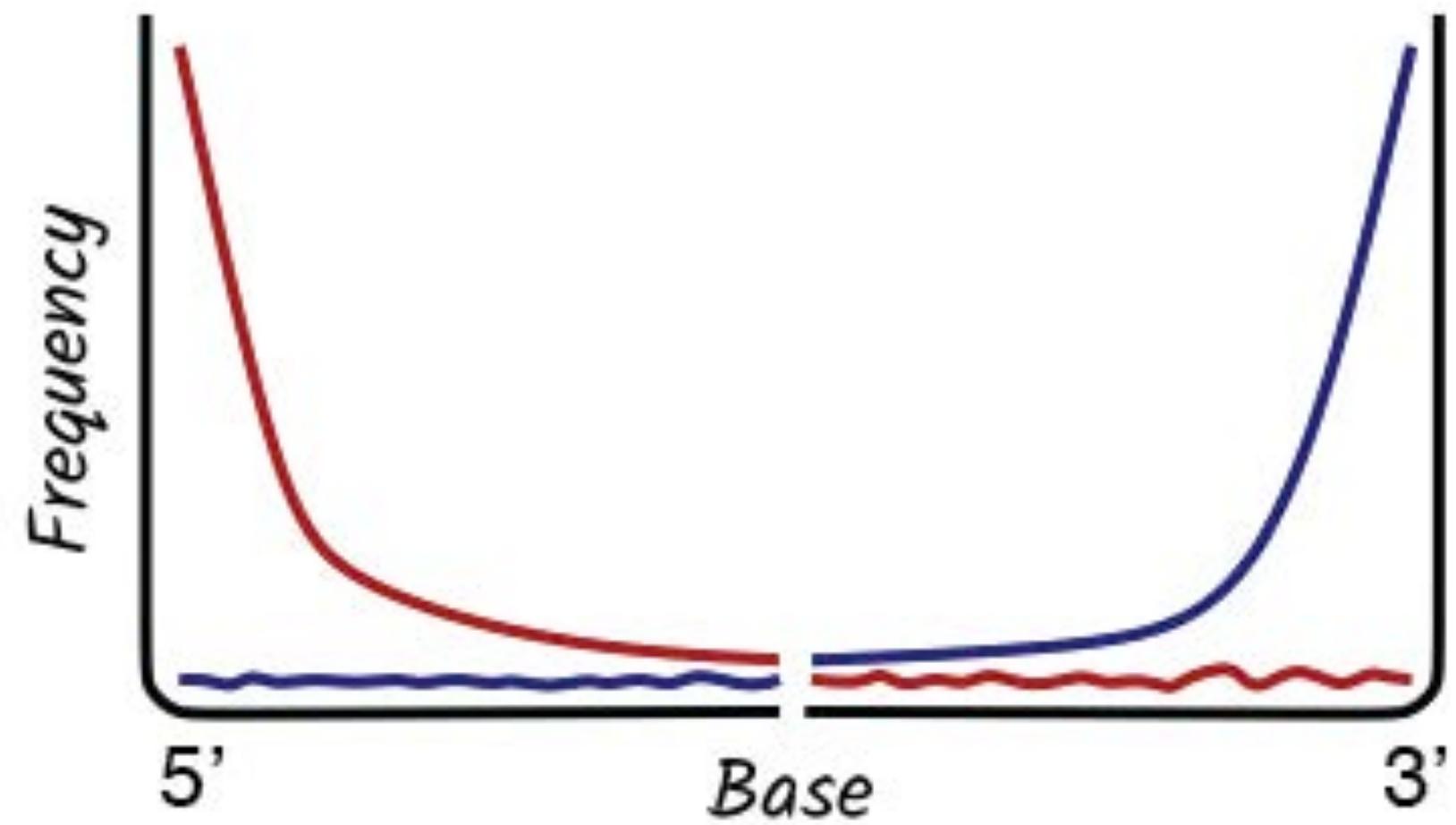


Ancient DNA primer
Characteristics and challenges

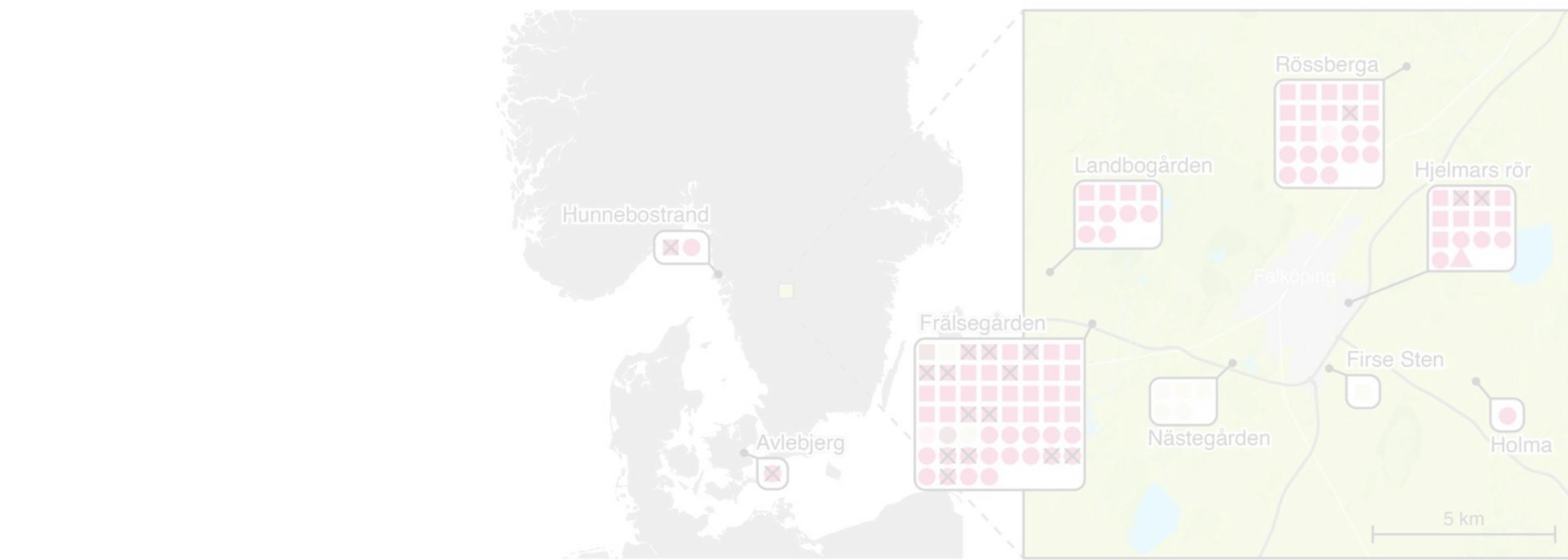


Ancient DNA showcase
Kinship and plague in Stone Age Scandinavia

Topics for today's lecture



Ancient DNA primer
Characteristics and challenges



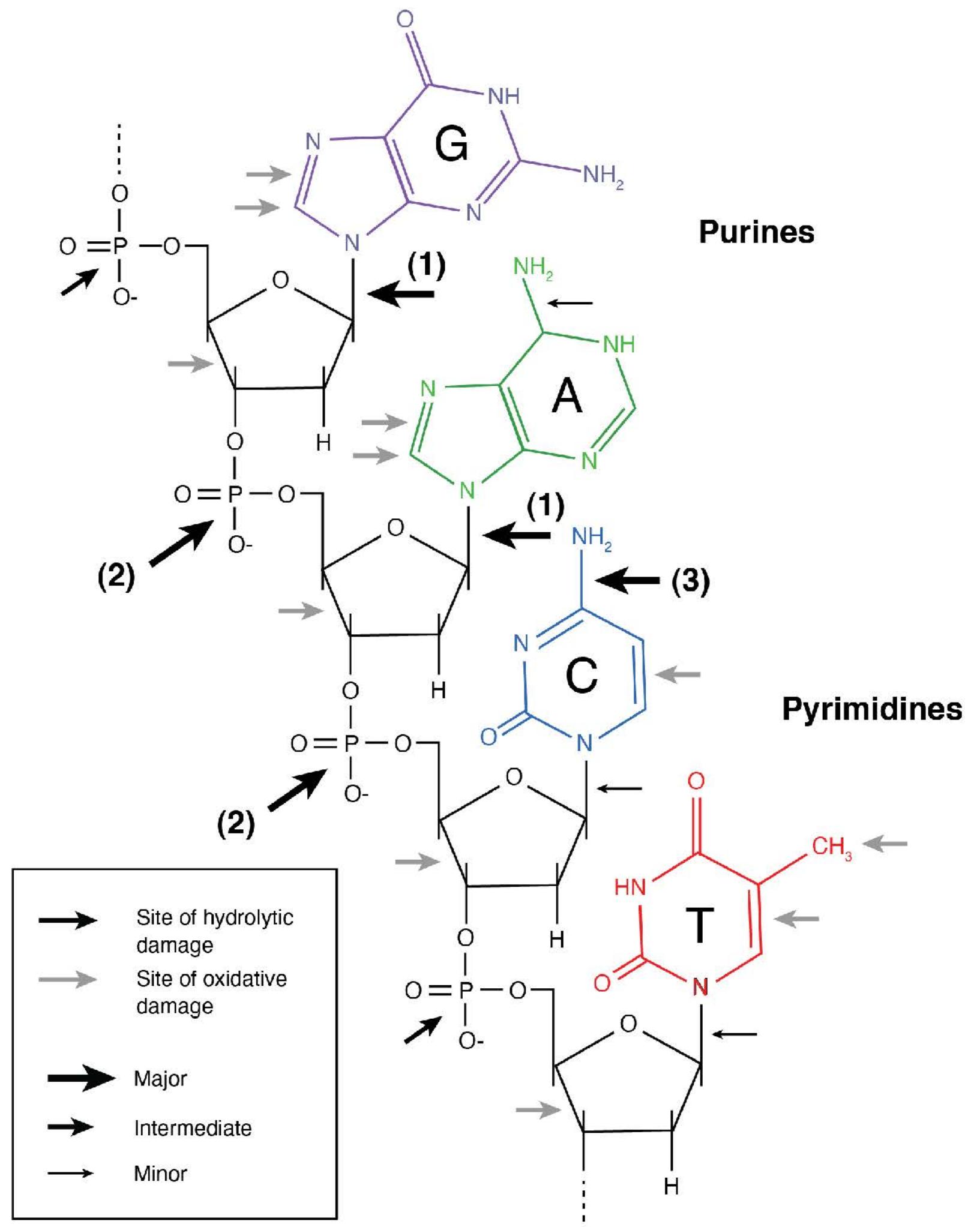
Ancient DNA showcase
Kinship and plague in Stone Age Scandinavia

Characteristics of ancient DNA

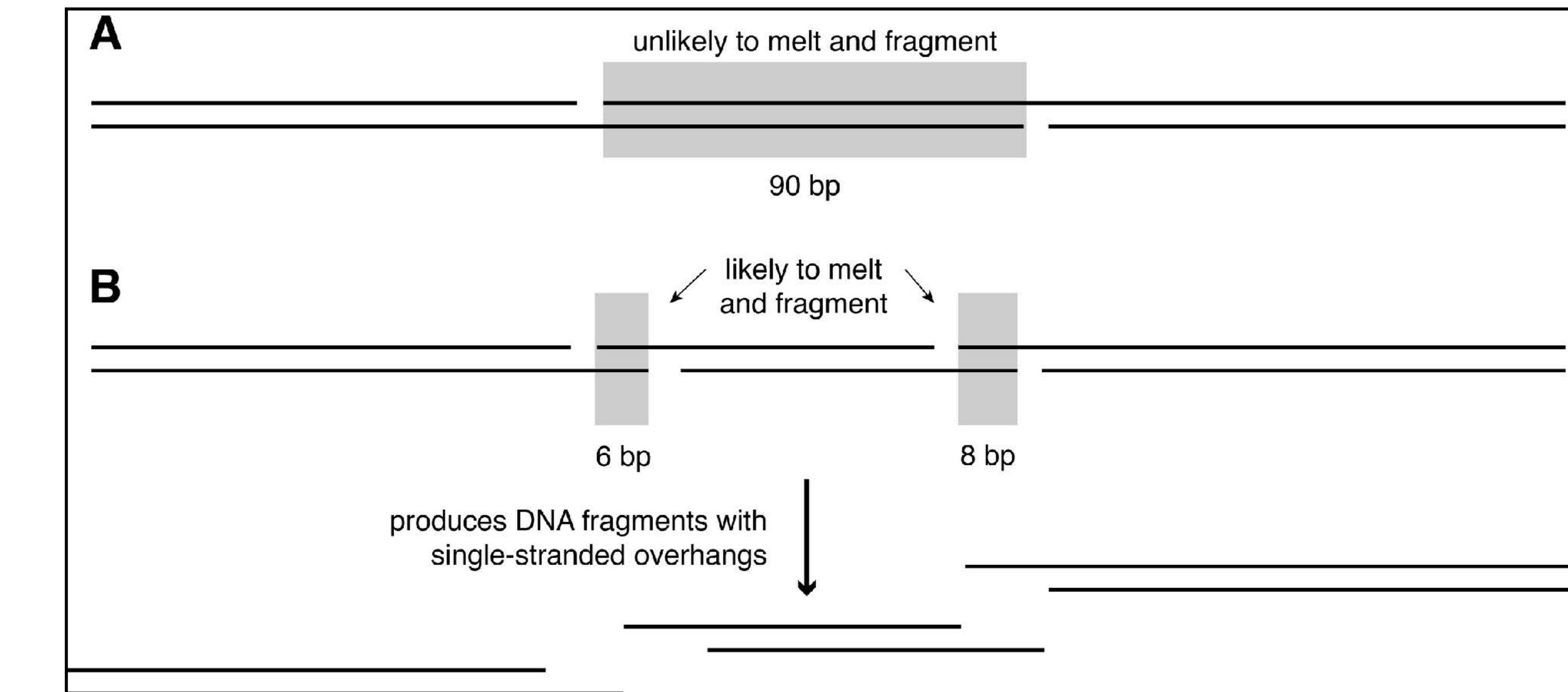
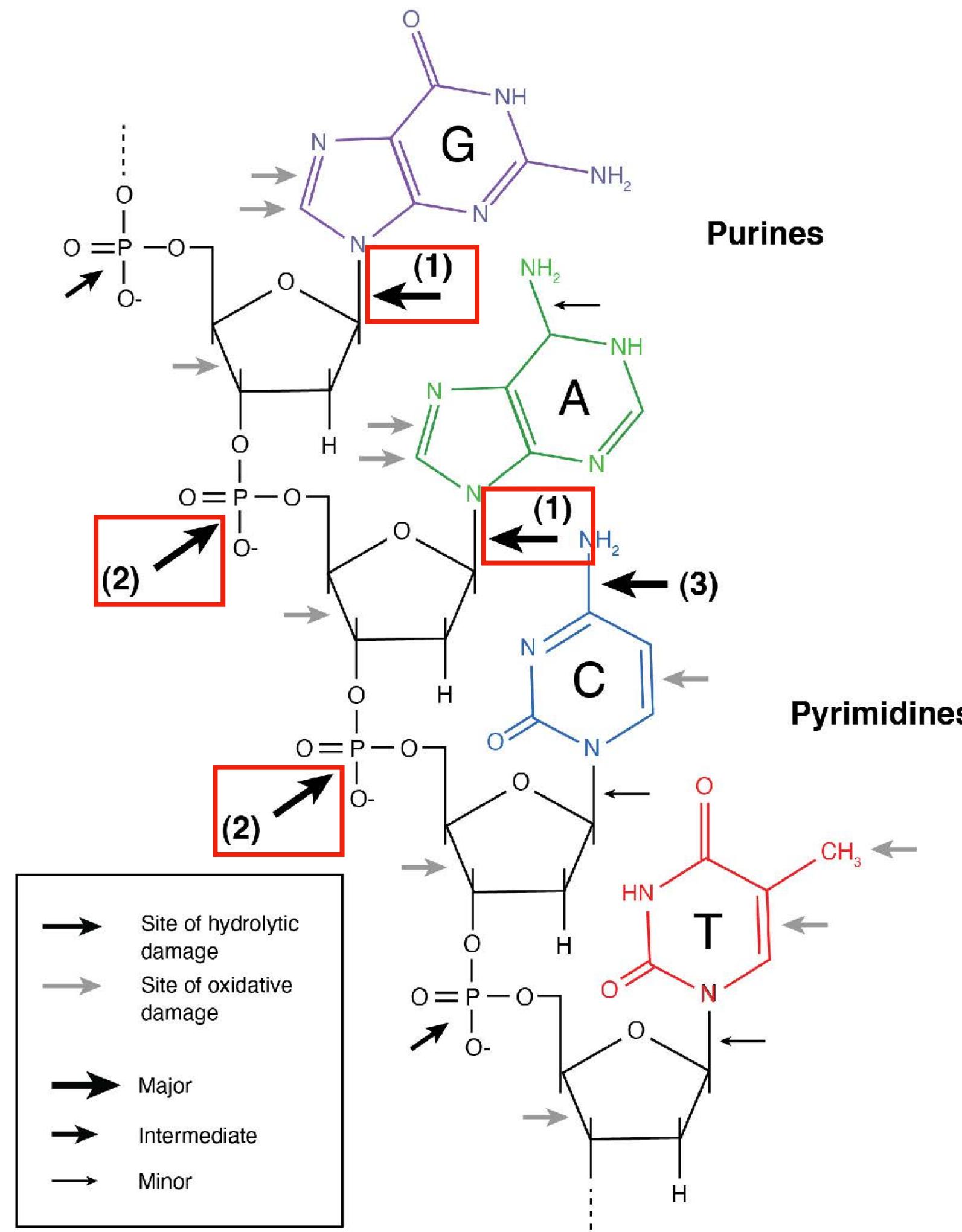


5,000 year-old human remains from the Provadia salt mine, Bulgaria

How does DNA degrade?

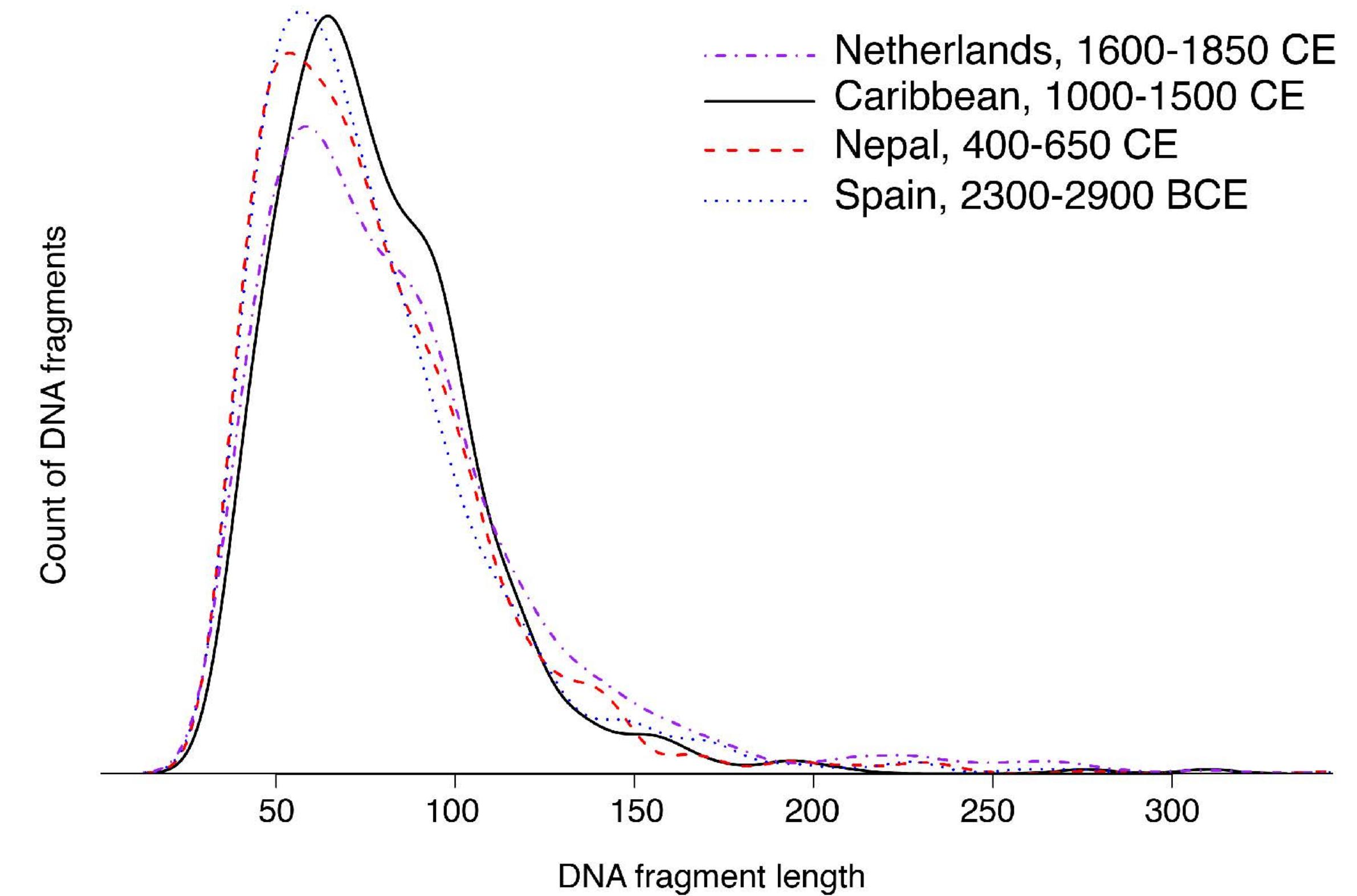


Fragmentation of DNA



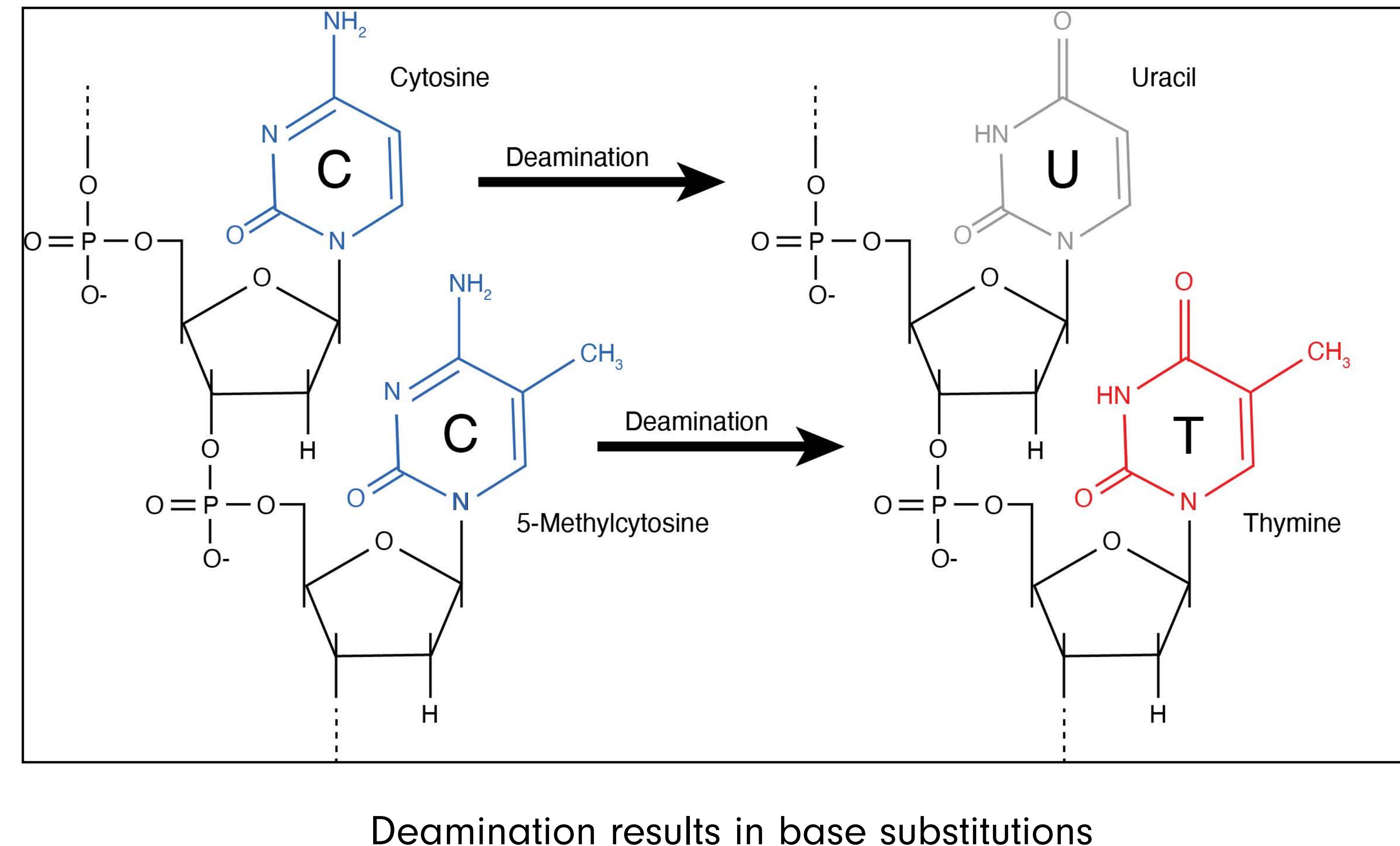
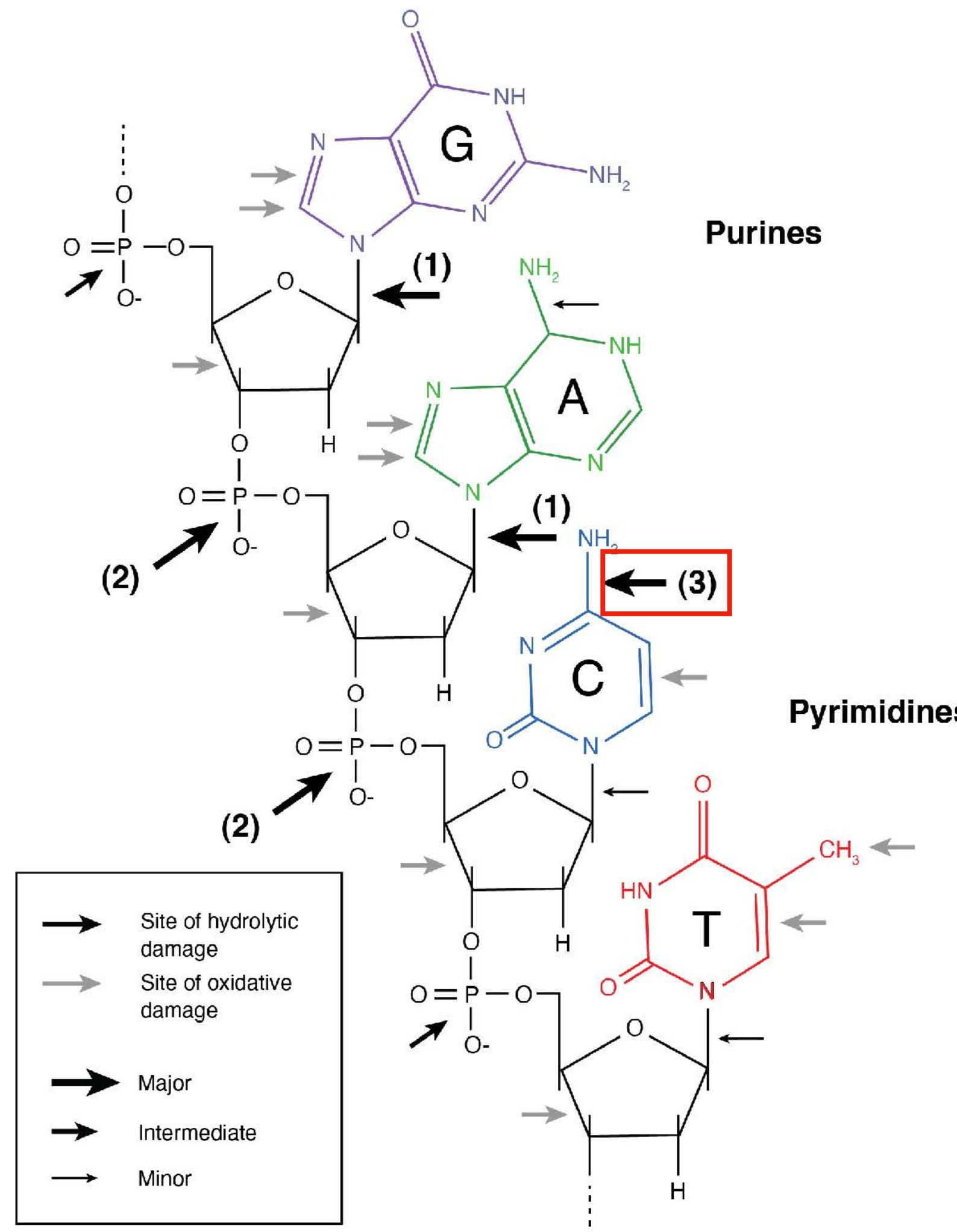
Depurination and nicking results in DNA fragmentation

Characteristics of ancient DNA

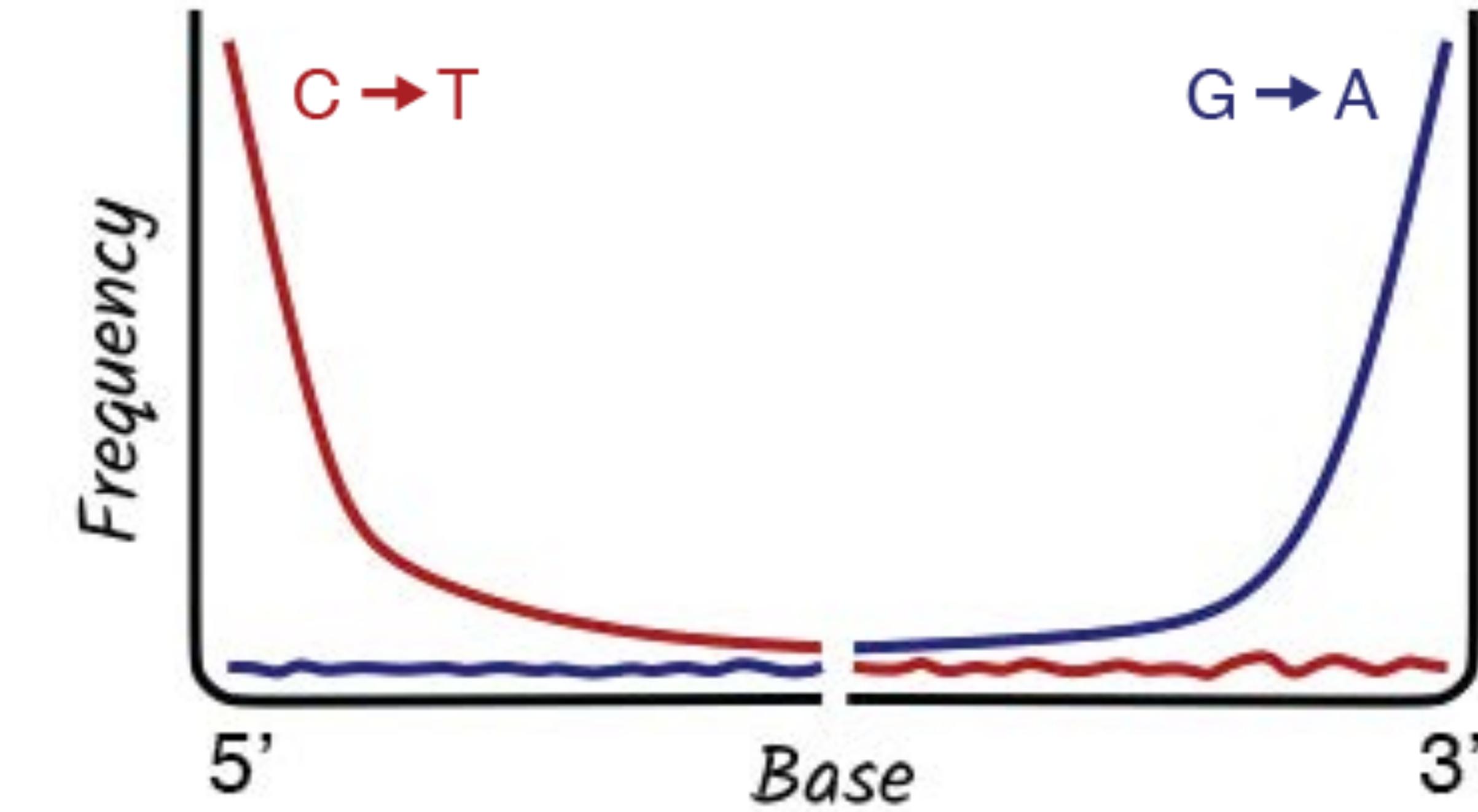


Ancient DNA is short and fragmented

Accumulation of DNA damage

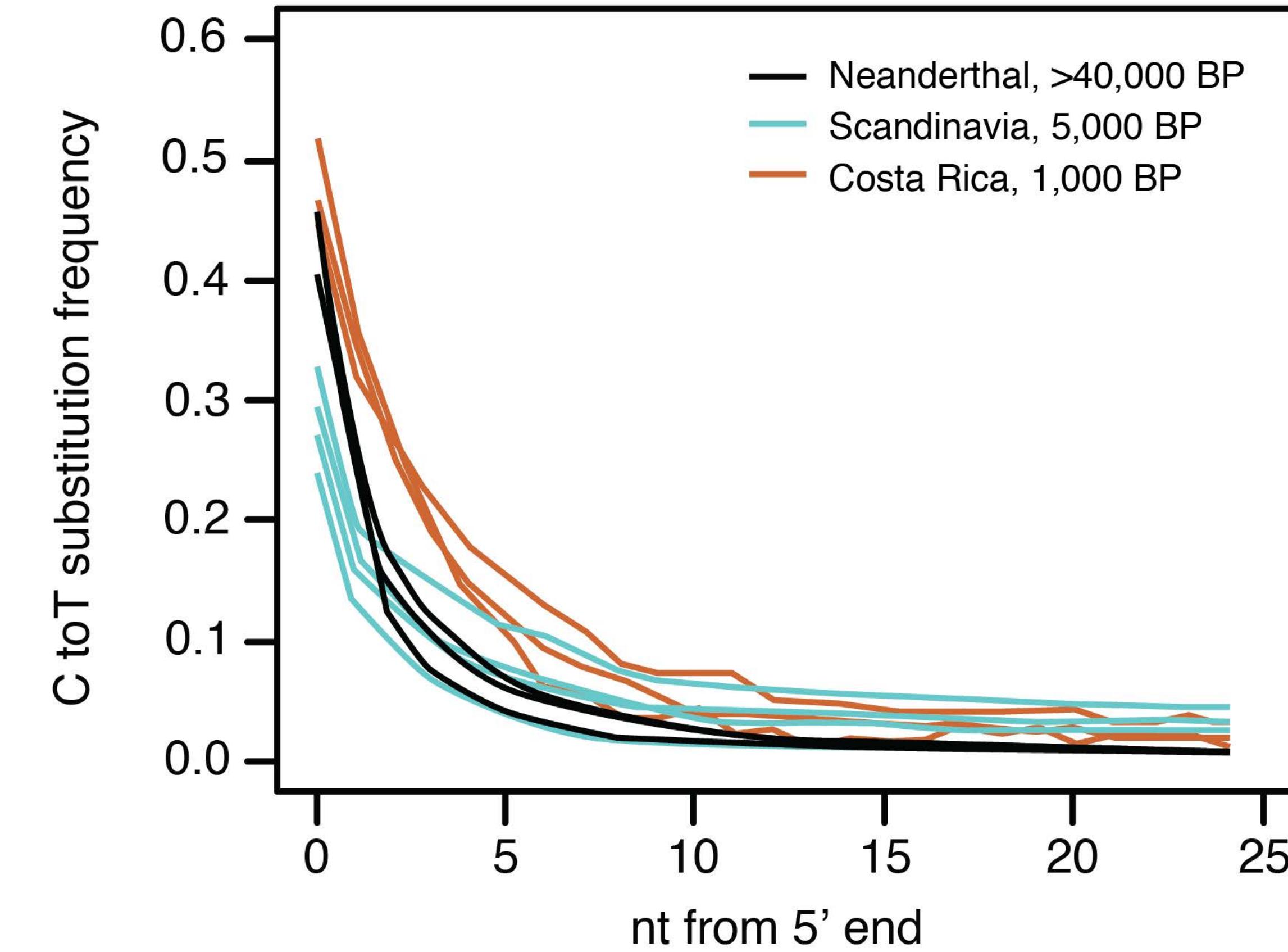
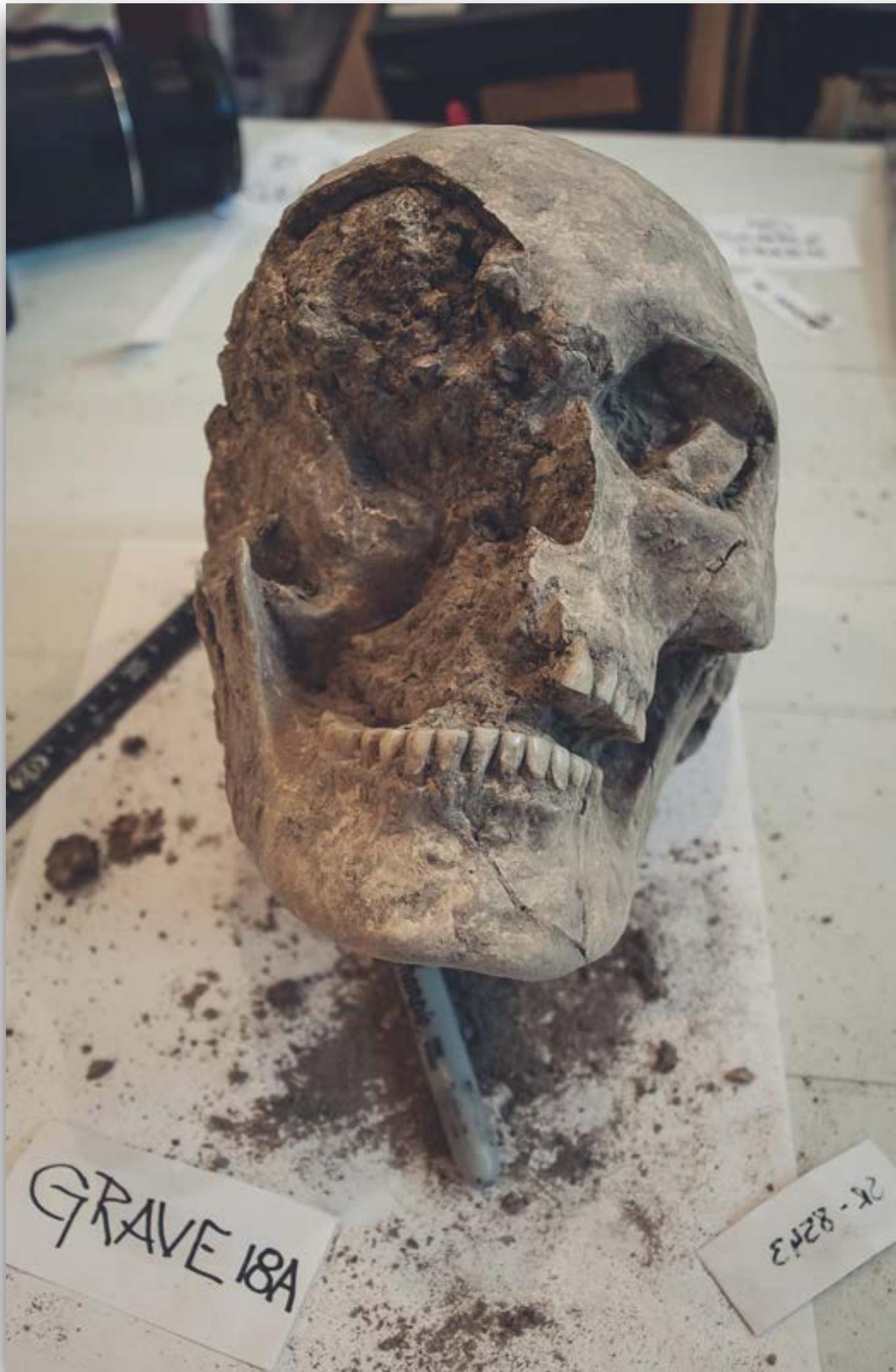


Characteristics of ancient DNA



Ancient DNA is damaged with characteristic substitution patterns

Characteristics of ancient DNA



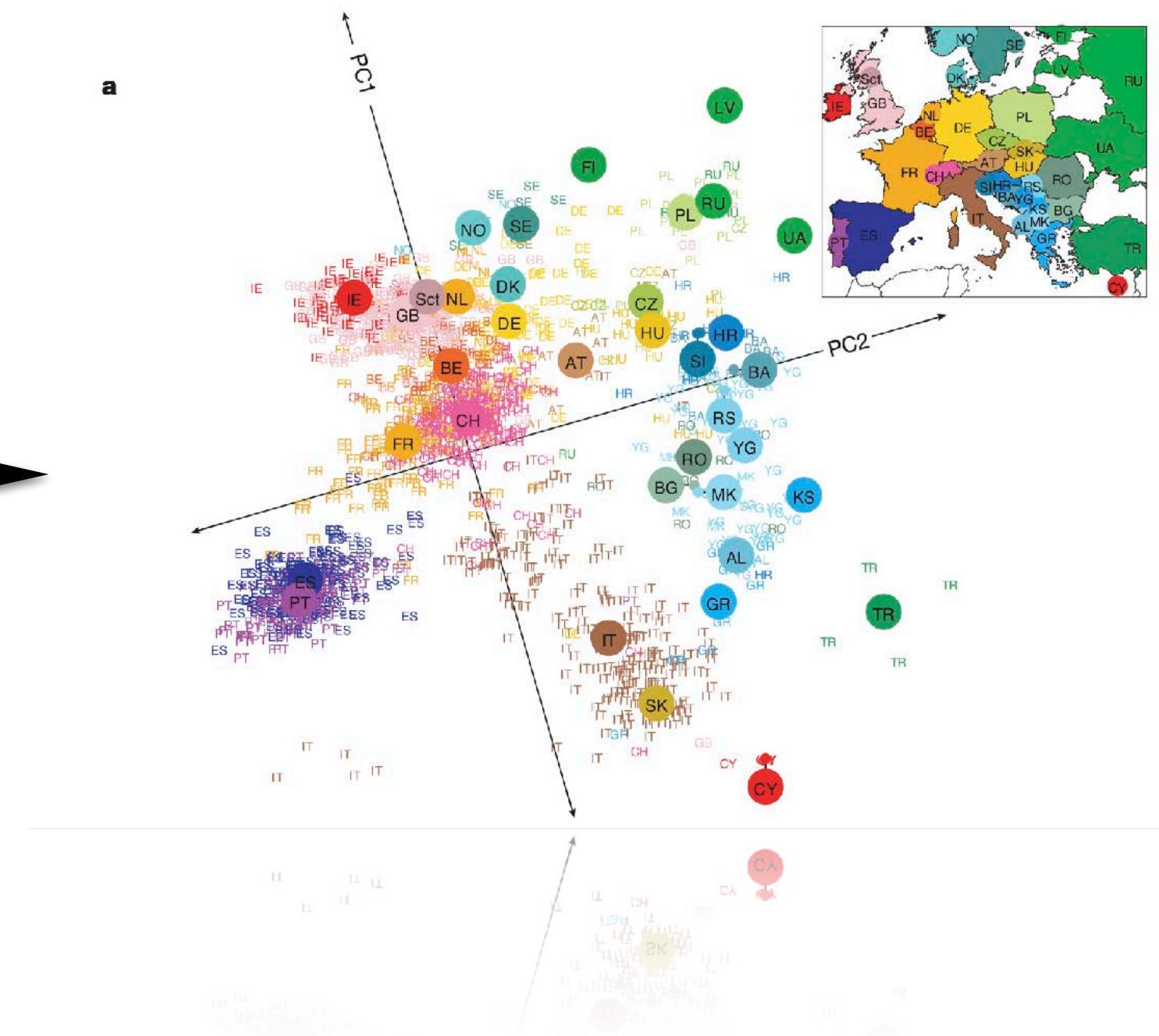
Ancient DNA damage rates depend on microenvironmental conditions

Ancient DNA challenges

Sample

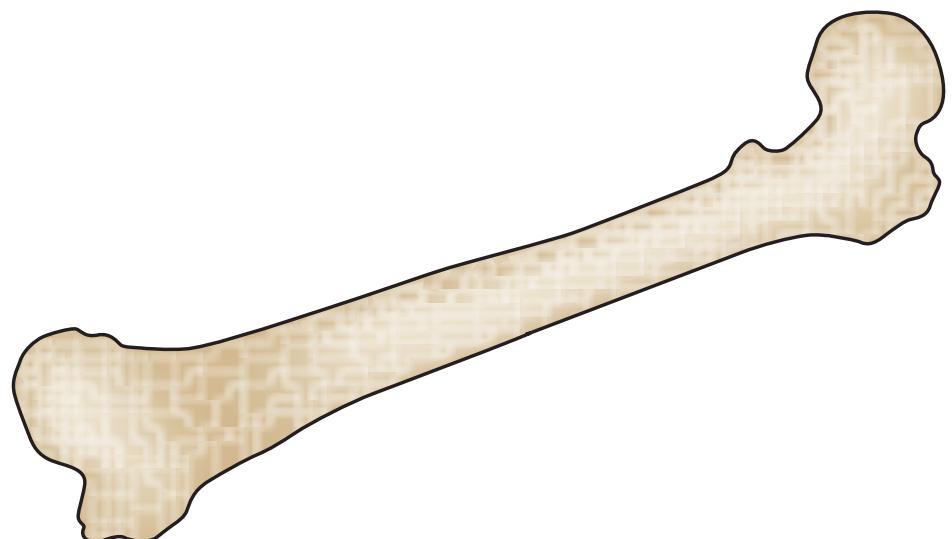
DNA sequences

Analysis results



Ancient DNA challenges

Sample



DNA recovery

Preservation
Sample material

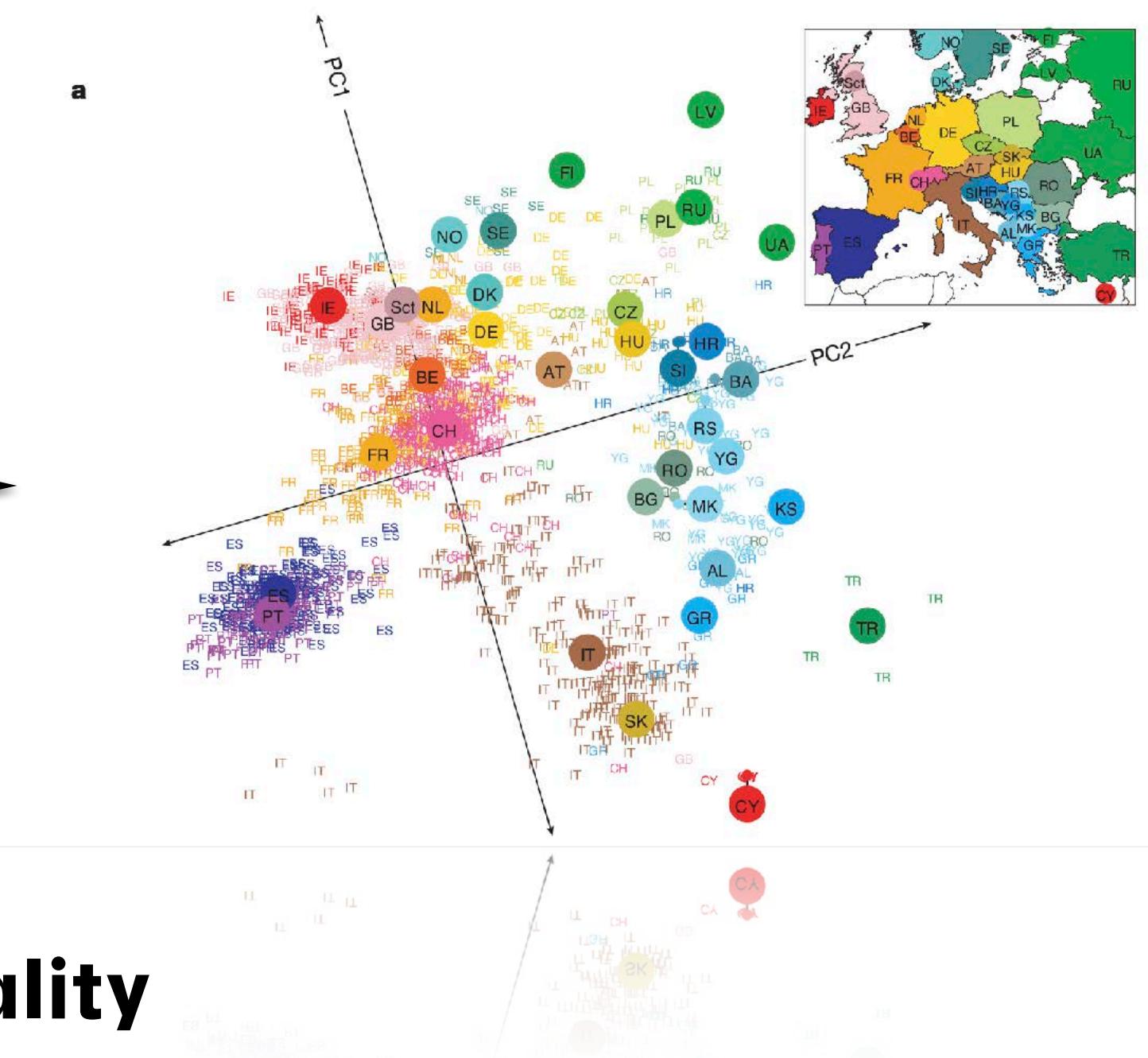
DNA sequences



Data quality

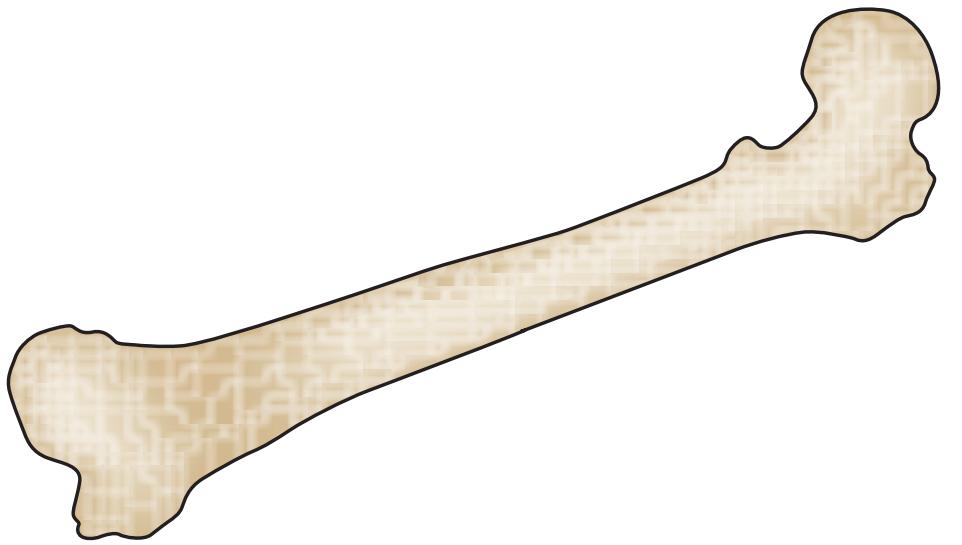
Contamination
Genome coverage
Error rates

Analysis results



Ancient DNA challenges

Sample



DNA recovery

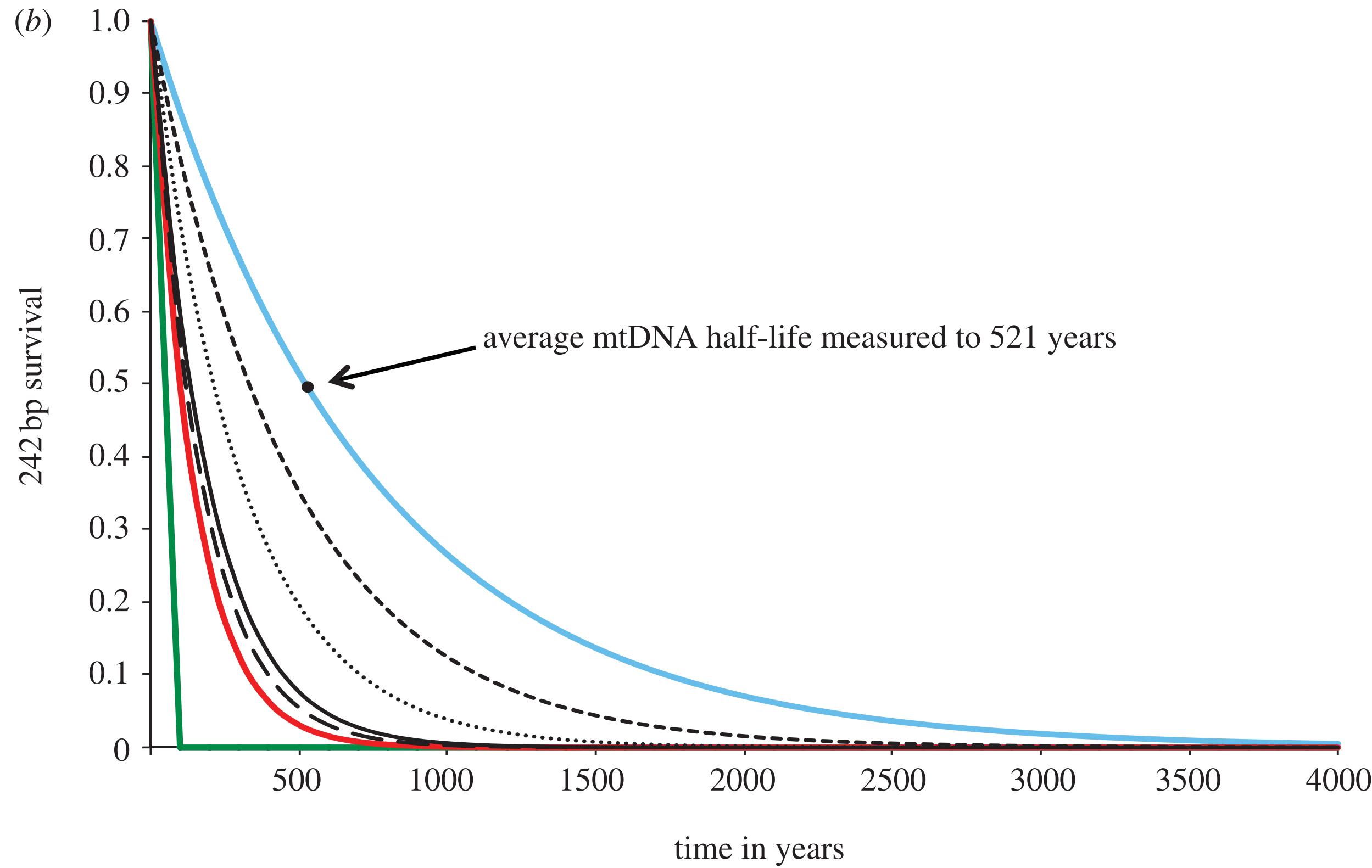
Preservation Sample material

DNA sequence



Analysis results

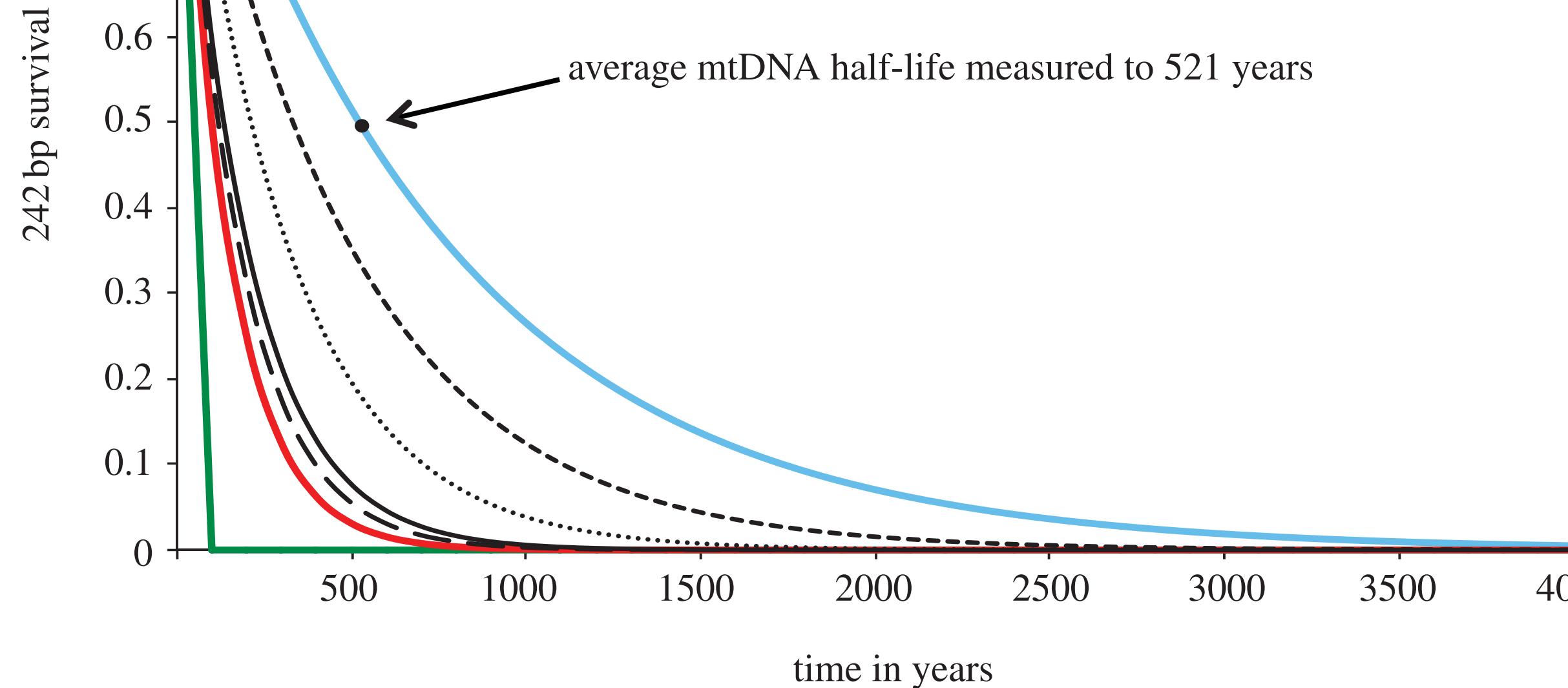
Challenges - Ancient DNA preservation



Ancient DNA fragmentation follows exponential decay dynamics

Challenges - Ancient DNA preservation

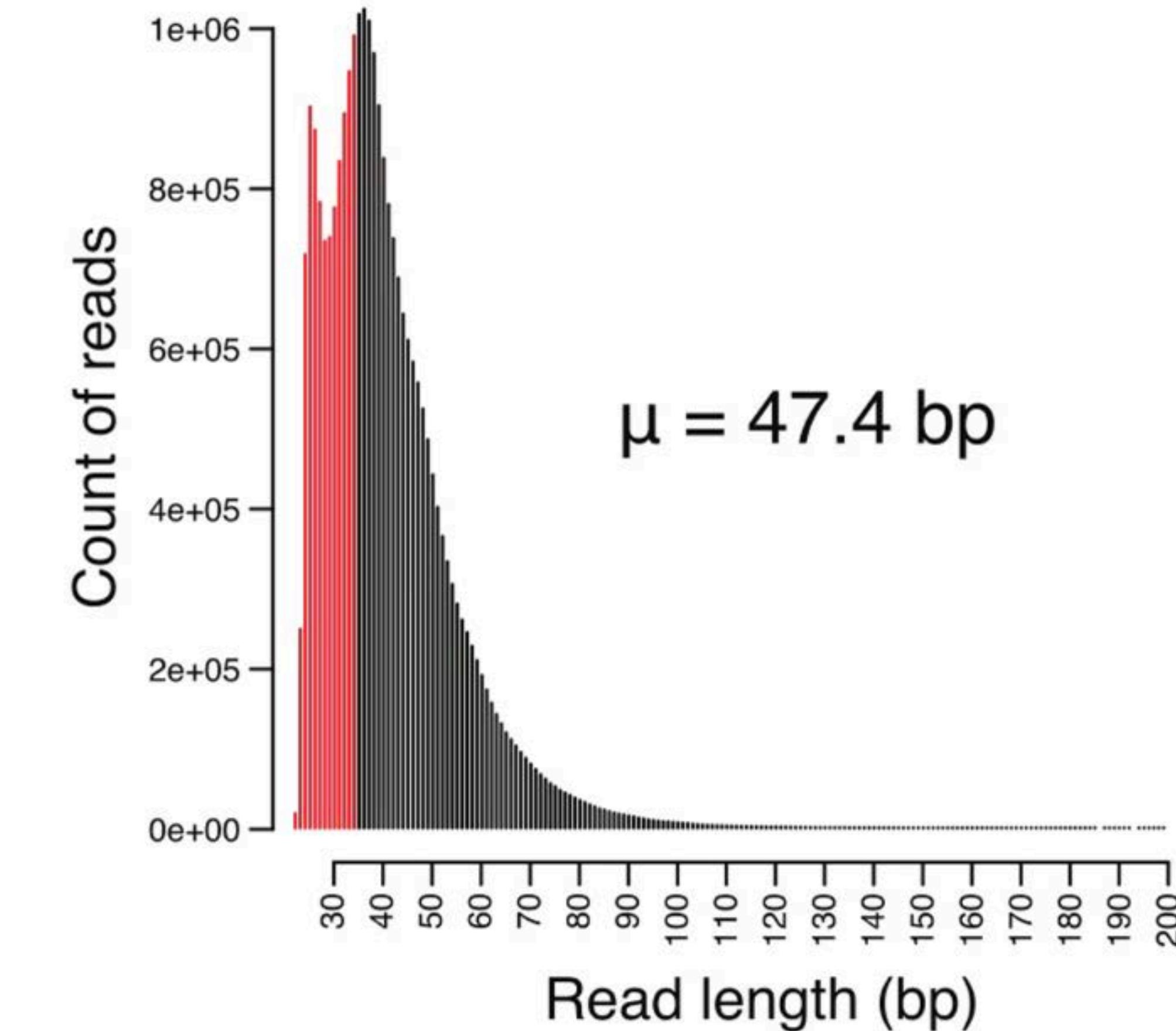
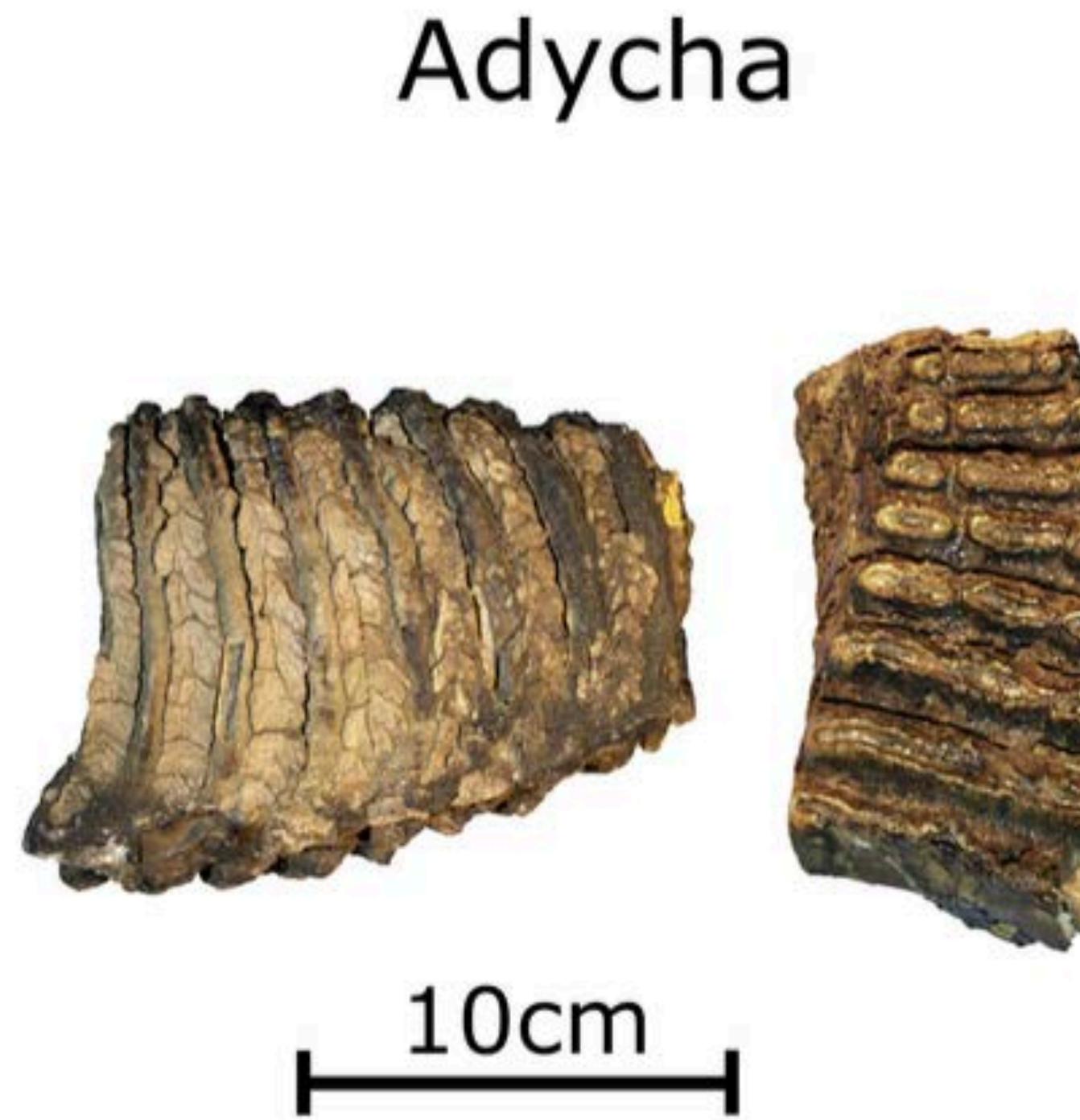
(b)



Average fragment length of 6.8 million-year old DNA
1 base pair

temperature	k per site per year	half-life (years), 30 bp	half-life (years), 100 bp	half-life (years), 500 bp	average length at 10 kyr	time (years) until average length = 1 bp
25°C	4.5×10^{-5}	500	150	30	2 bp	22 000
15°C	7.6×10^{-6}	3000	900	180	13 bp	131 000
5°C	1.1×10^{-6}	20 000	6000	1200	88 bp	882 000
-5°C	1.5×10^{-7}	158 000	47 000	9500	683 bp	6 830 000

Challenges - Ancient DNA preservation



DNA from a 1.2 million year old mammoth tooth is highly fragmented

Challenges - Endogenous DNA content

Low endogenous

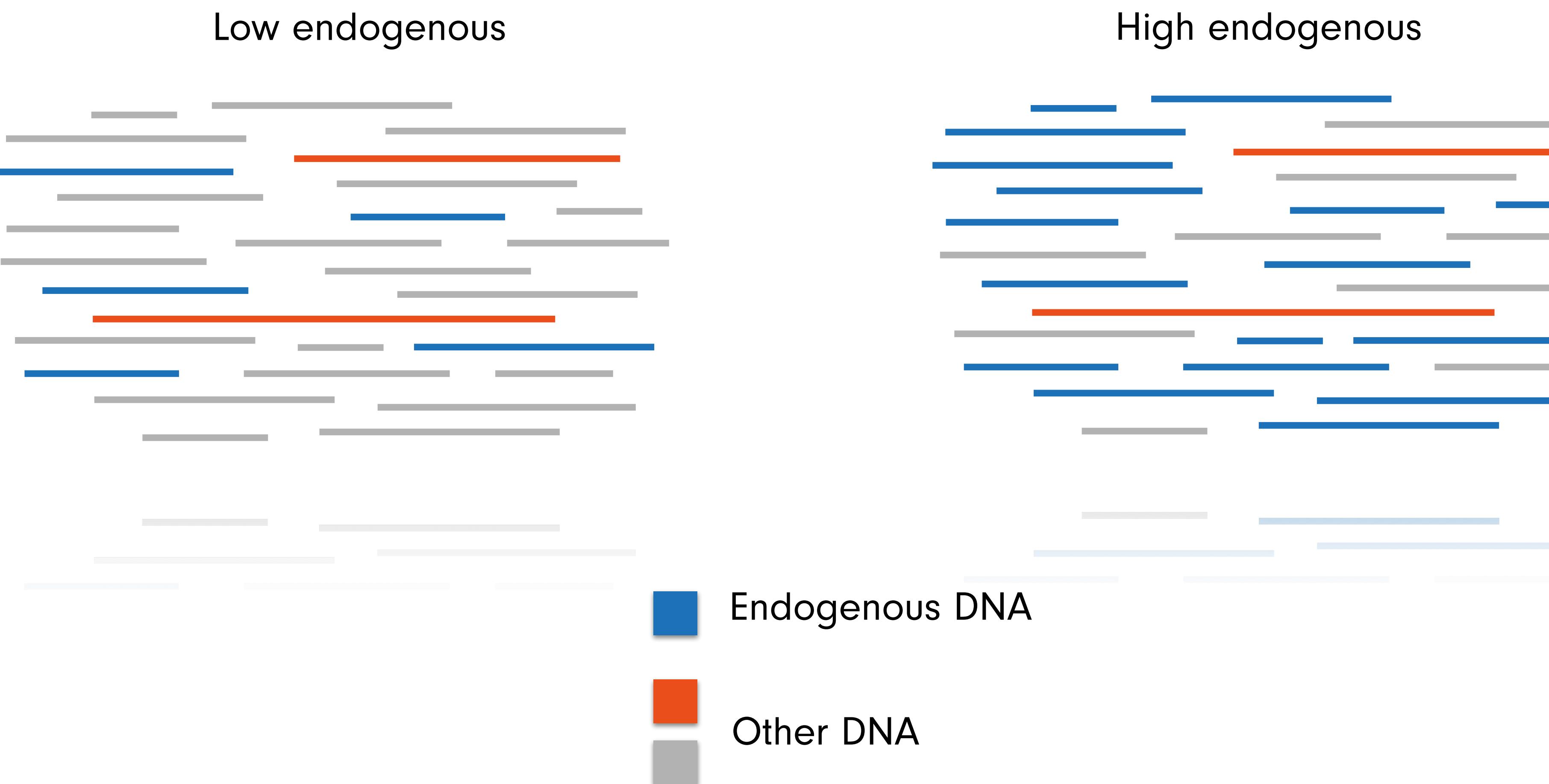


█ Endogenous DNA

█ Other DNA

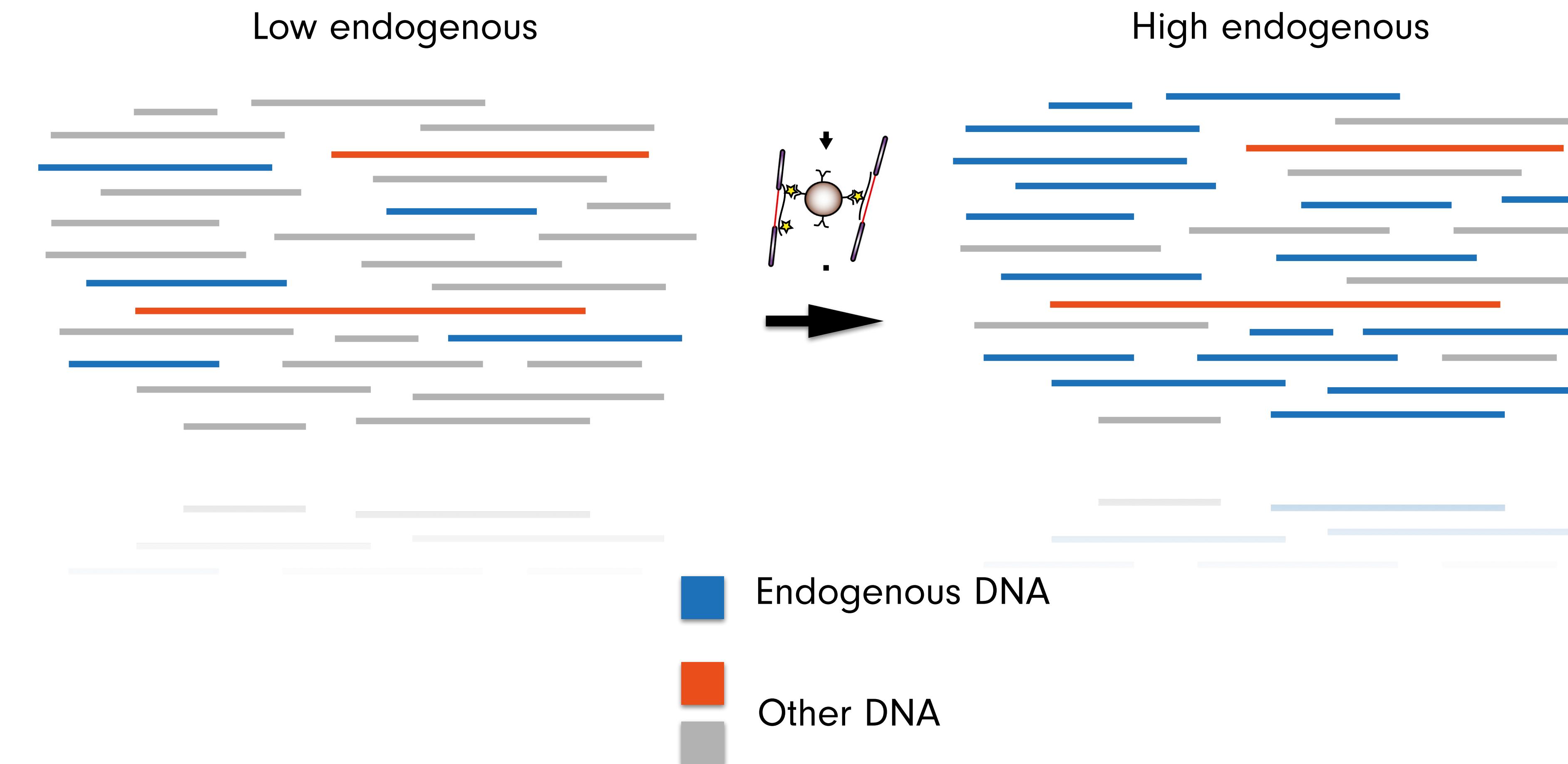


Challenges - Endogenous DNA content



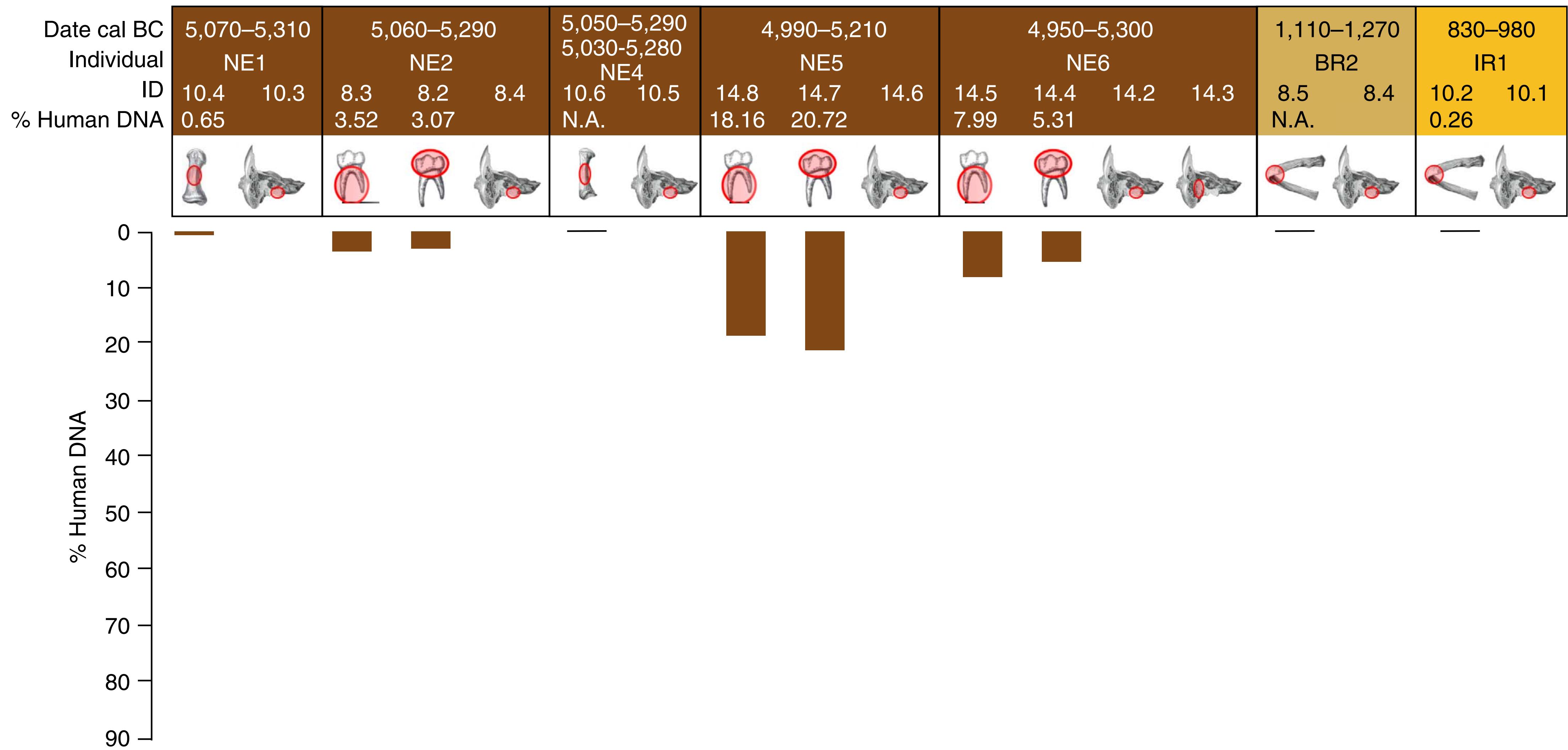
Endogenous DNA content varies substantially between samples

Challenges - Endogenous DNA content

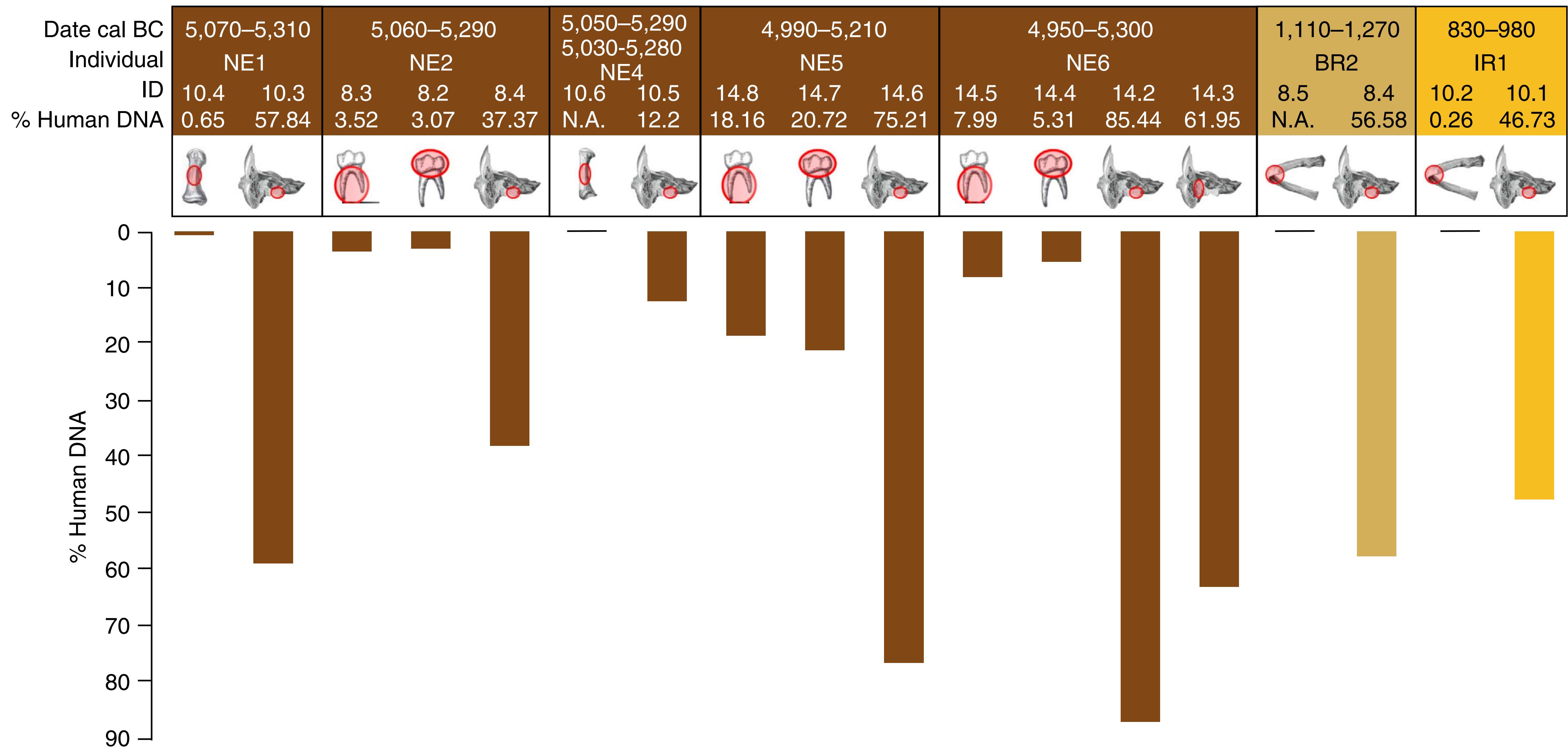


DNA capture enrichment (targeted or whole genome) to increase endogenous DNA

Impact of sample material



Impact of sample material



Ancient DNA challenges

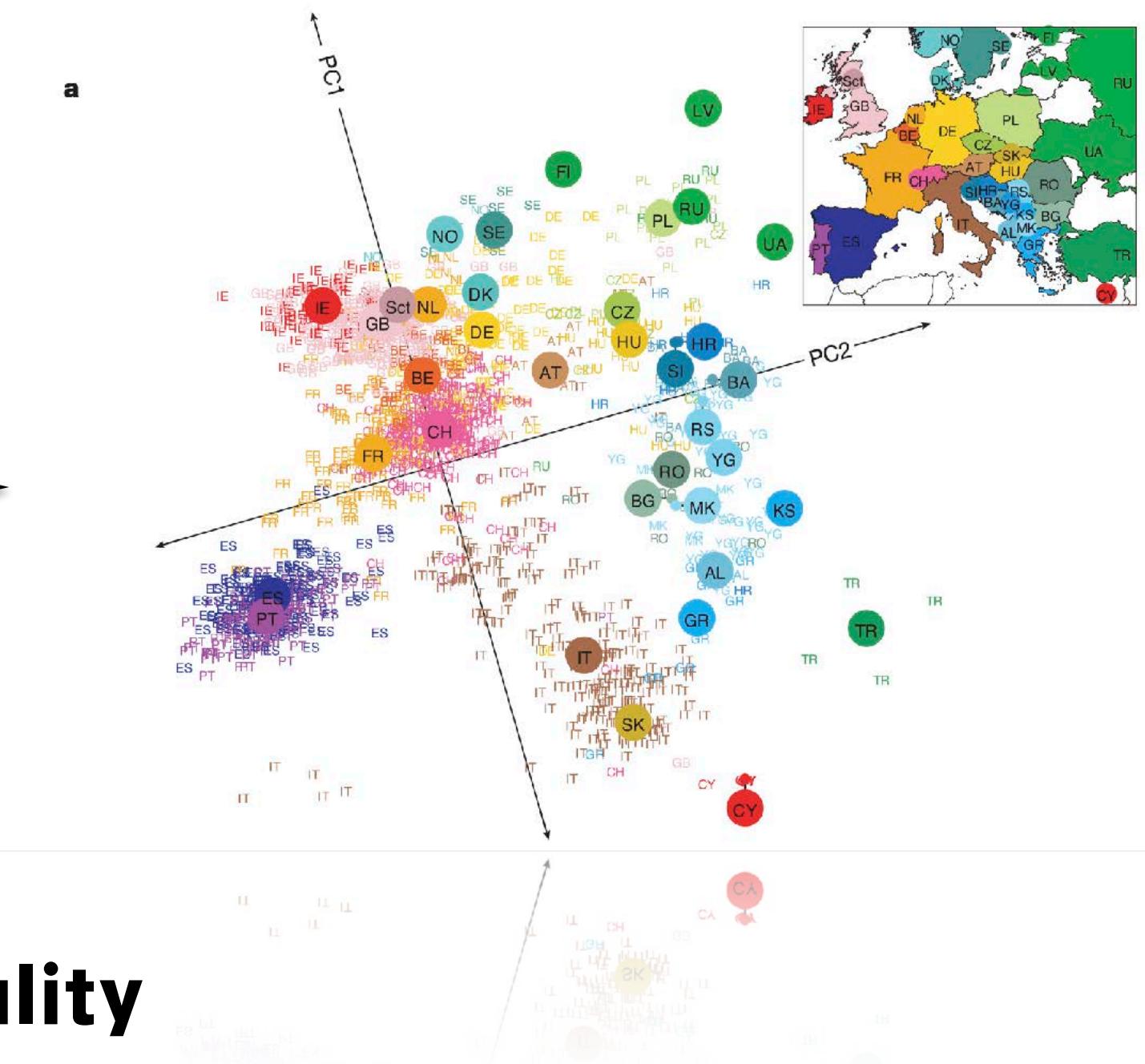
Sample



DNA sequences



Analysis results



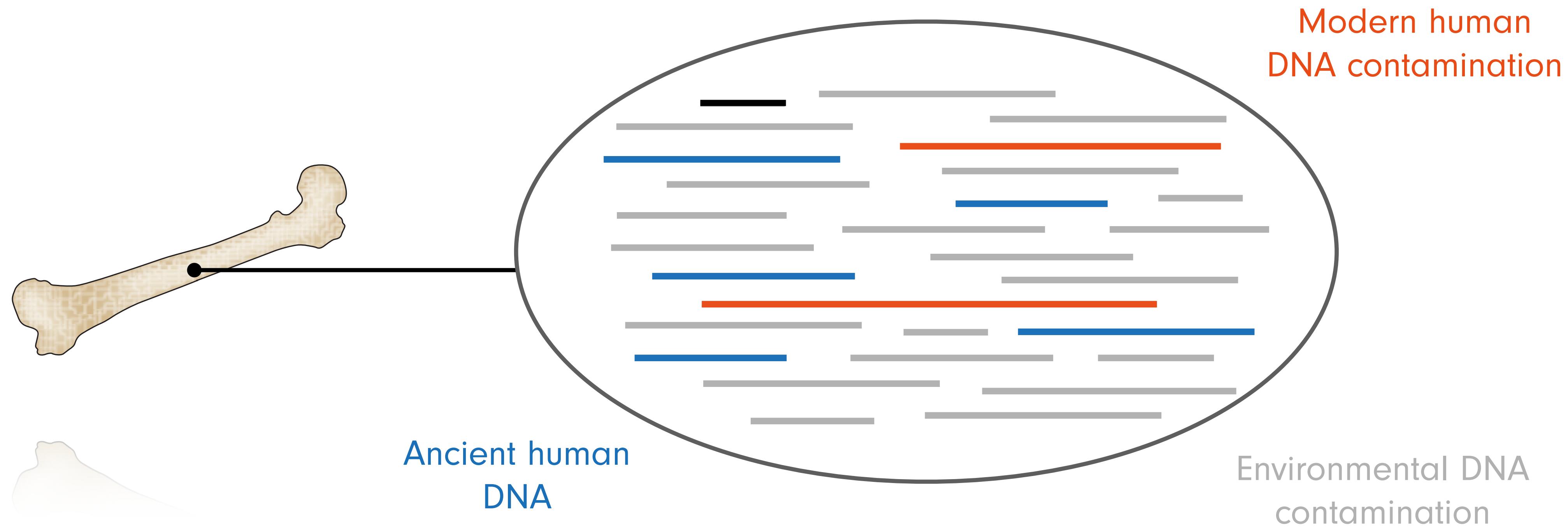
Data quality

Contamination
Genome coverage
Error rates

Challenges - Contamination



Challenges - Contamination



Ancient DNA data is a complex mixture of DNA from different sources

Challenges - Contamination

Barcelona lab

Monti Lessini (L906-H924)

- B.2.1
- B.2.2
- B.2.3
- B.2.4
- B.2.5
- B.2.6
- B.2.7
- B.2.8
- B.2.9
- B.2.10
- B.2.11
- B.2.12

TCATCTACGCCTCCACAGCCAGGAGCTCCGCAGGACGCTCAAGGAGGTGCTGACA

A handwriting practice sheet featuring ten rows of dotted lines. The first three rows are labeled with a large red 'G' at the right end of the top line. The remaining seven rows are blank for independent practice.

Florence lab

Monti Lessini (L884-H93)

CTCCTGGTGA
F.1.1
F.1.2
F.1.3
F.1.4
F.1.5
F.1.6
F.1.7
F.1.8
F.1.9
F.1.10
F.1.11
F.1.12
F.1.13
F.1.14
F.1.15
F.1.16
F.1.17
F.1.18
F.1.19
F.1.20
F.1.21
F.1.22
F.1.23

TGCAATGCCATCATCGACCCCTCATCTACGCCCTCACAGCCAGGAGCTCCGCAGGACGCTAAGGAGGTGCTGACA'

A handwriting practice sheet featuring a grid of dotted lines. The grid consists of 10 horizontal rows and 3 vertical columns. The first column contains the uppercase letter 'G' in black at the top and the lowercase letter 'g' in red below it. The second column contains the uppercase letter 'G' in red at the top and the lowercase letter 'g' in black below it. The third column contains only red dots. The rows are separated by horizontal dotted lines, and the columns are separated by vertical dotted lines.

MC1R gene fragments amplified from Monti Lessini Neandertal DNA extract

Challenges - Contamination

Barcelona lab

Monti Lessini (L906-H924)

- B.2.1
- B.2.2
- B.2.3
- B.2.4
- B.2.5
- B.2.6
- B.2.7
- B.2.8
- B.2.9
- B.2.10
- B.2.11
- B.2.12

TCATCTACGCCTTCCACAGCCAGGAGCTCCGC**AGGACGCTAAGGAGGTGCTGACA**

Neandertal sequences

Florence lab

Monti Lessini (L884-H93)

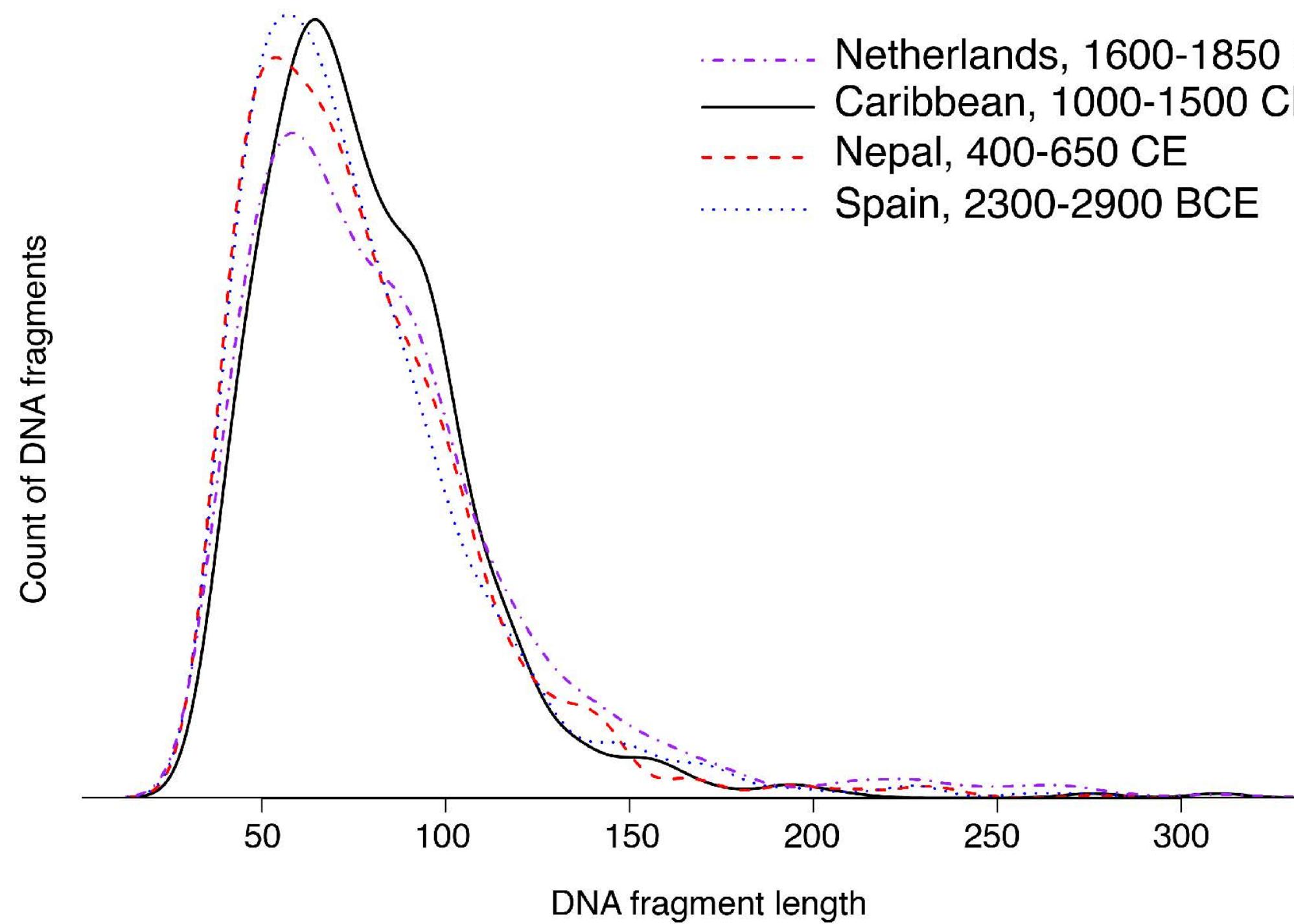
CTCCTGGTGA
F.1.1
F.1.2
F.1.3
F.1.4
F.1.5
F.1.6
F.1.7
F.1.8
F.1.9
F.1.10
F.1.11
F.1.12
F.1.13
F.1.14
F.1.15
F.1.16
F.1.17
F.1.18
F.1.19
F.1.20
F.1.21
F.1.22
F.1.23

TGCAATGCCATCATCGACCCCTCATCTACGCCTCCACAGCCAGGAGCTCCGAGGACGCTCAAGGAGGTGCTGACAT

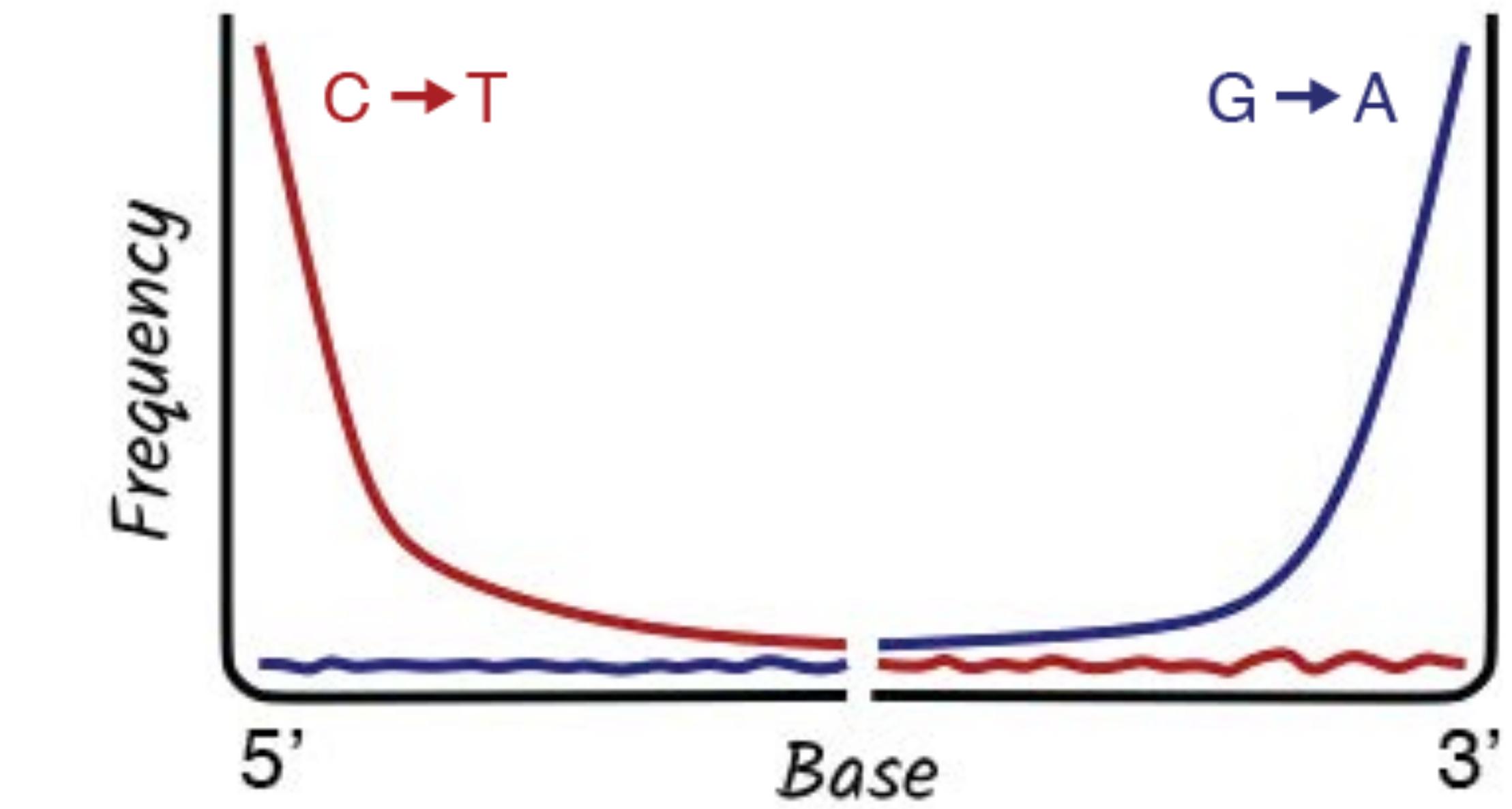
Neandertal sequences

MC1R gene fragments amplified from Monti Lessini Neandertal DNA extract

Authentication of ancient DNA

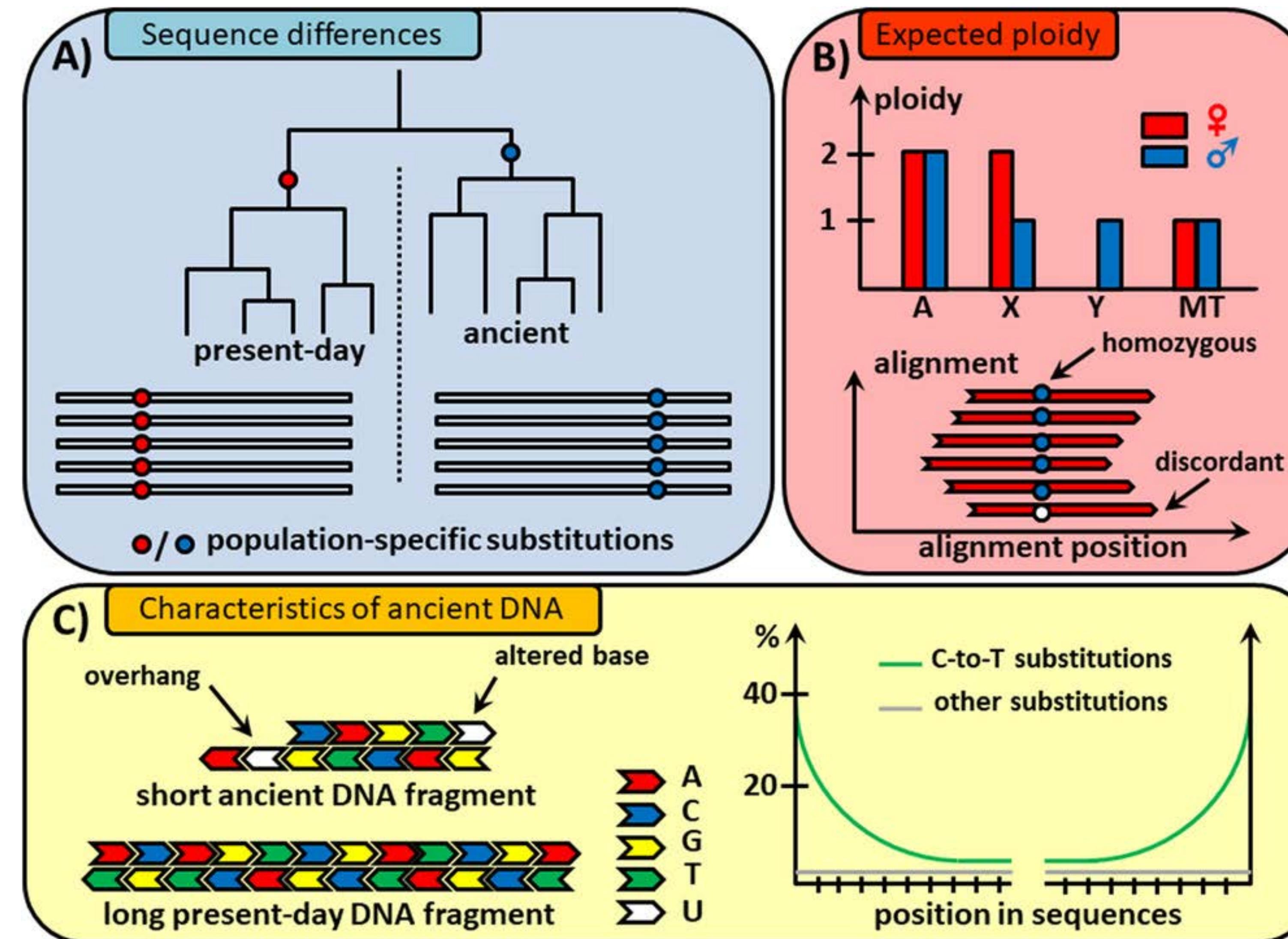


Ancient DNA is short and fragmented



Ancient DNA is damaged with characteristic substitution patterns

Authentication of ancient DNA



Signatures used to estimate contamination levels

Challenges - Genome coverage and errors

* * * * * * * * * *

00000000110000011001100000001010100000111000000001010001
0000100010001000100010000110001010000101010001000010000000

Diploid individual

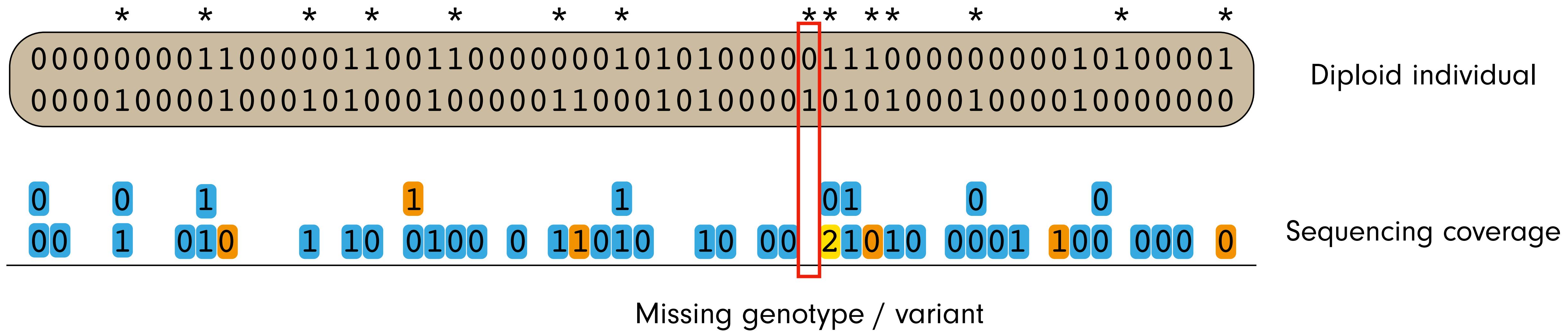
Challenges - Genome coverage and errors

Diploid individual

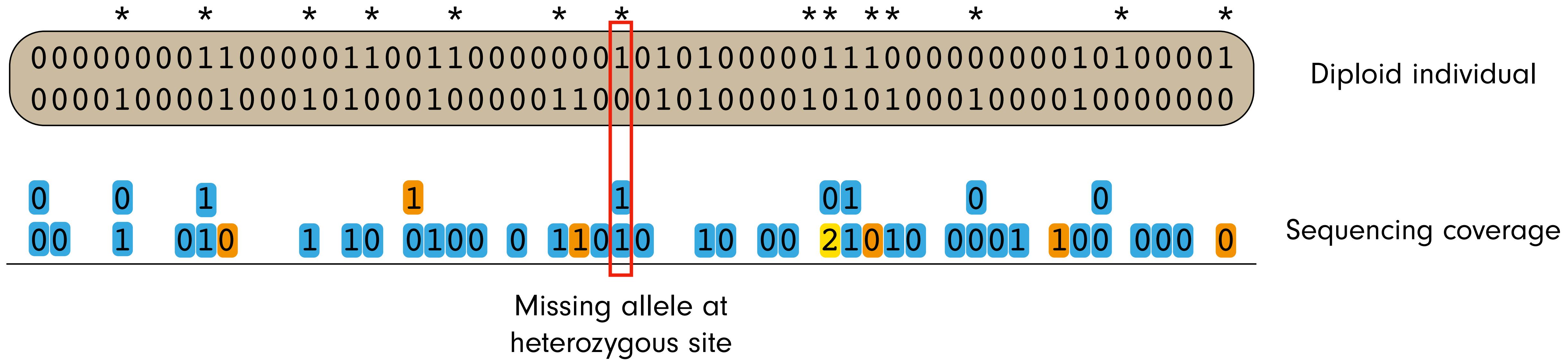
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00	1	010	1	10	0100	0	11010	10	00	21010	0001	100	000	0

Sequencing coverage

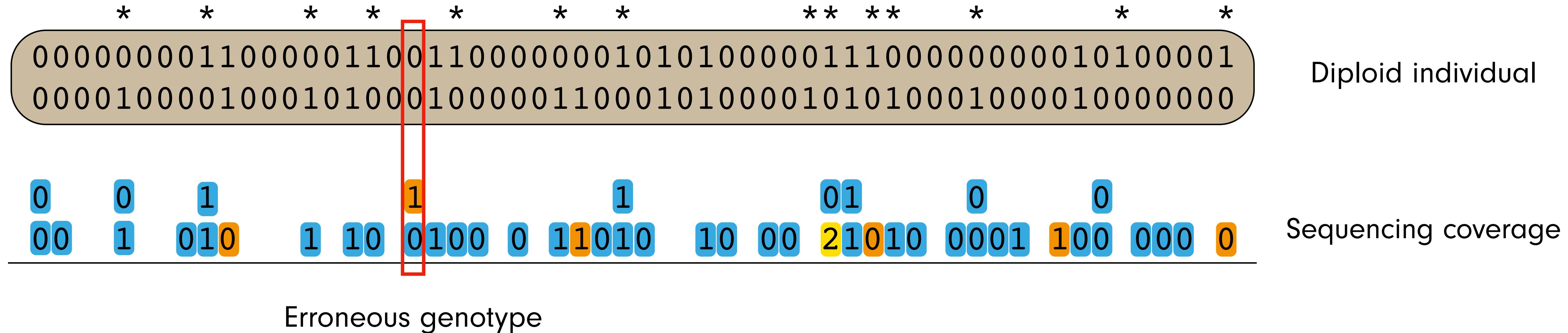
Challenges - Genome coverage and errors



Challenges - Genome coverage and errors



Challenges - Genome coverage and errors



Challenges - Genome coverage and errors

Diploid individual

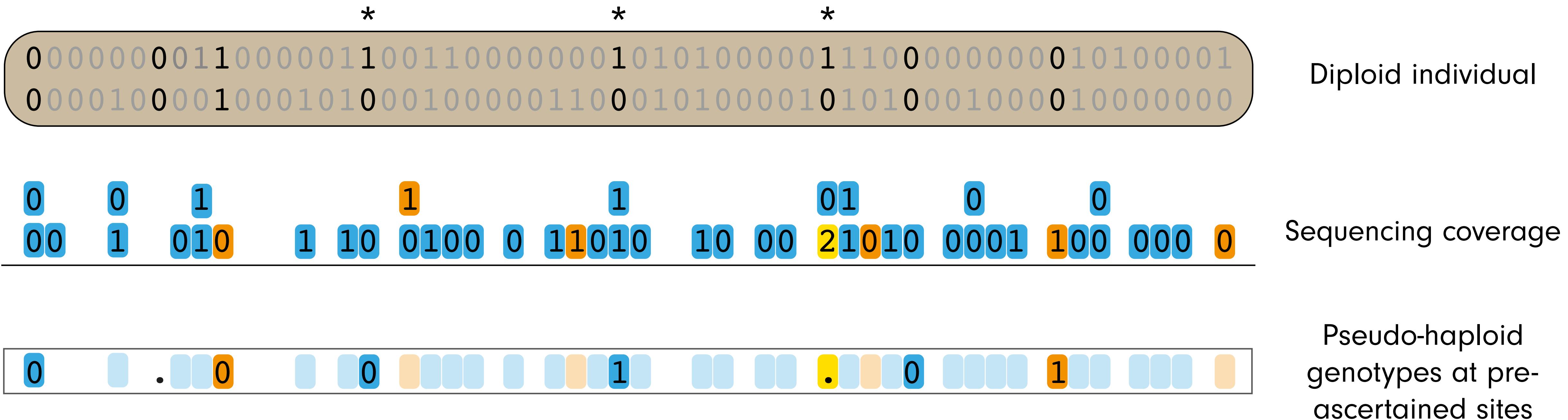
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00	1	010	1	10	0100	0	11010	0	11010	1	10	00	21010	0001	100	000	0

Sequencing coverage

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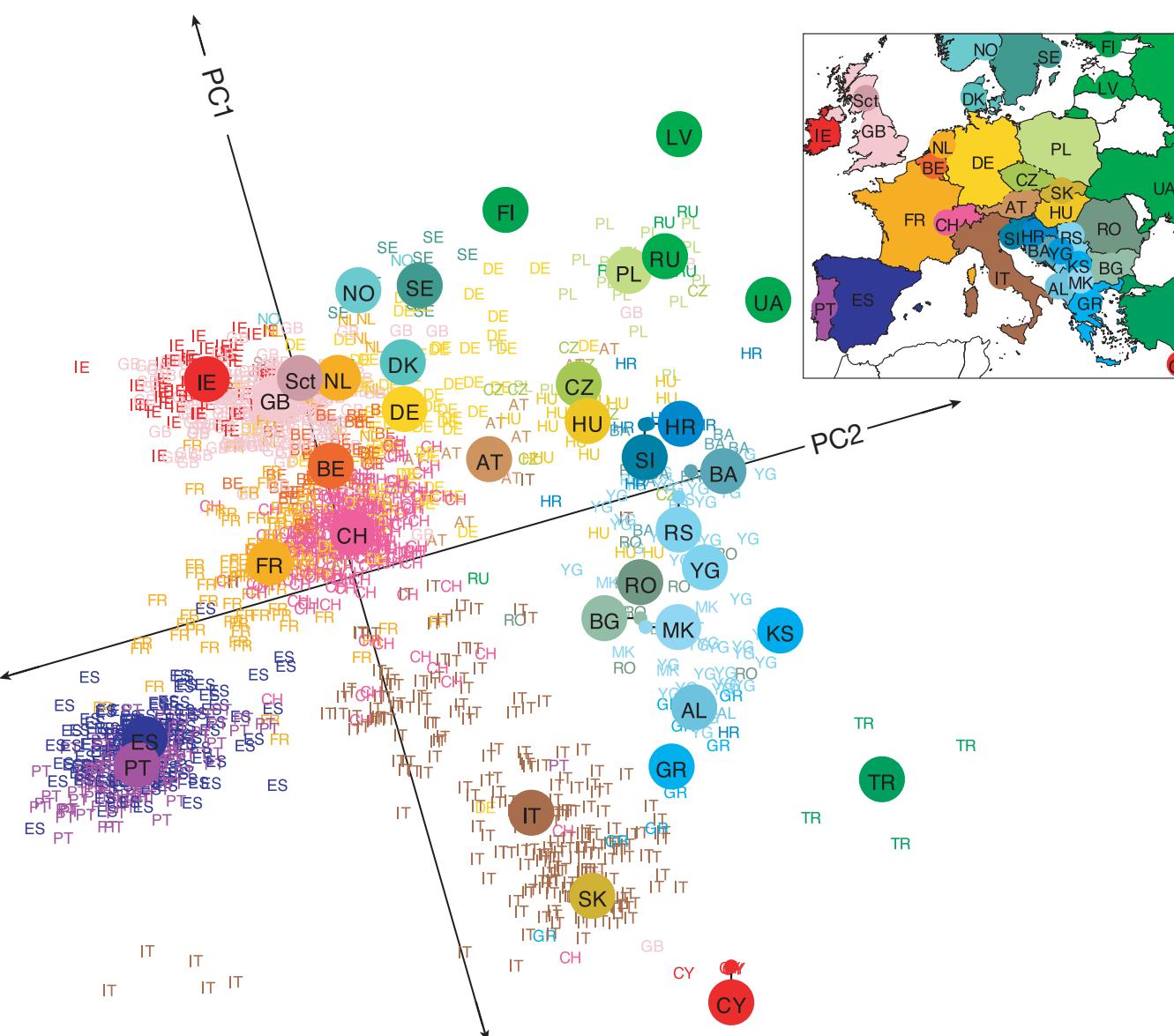
Pseudo-haploid genotypes

Challenges - Genome coverage and errors

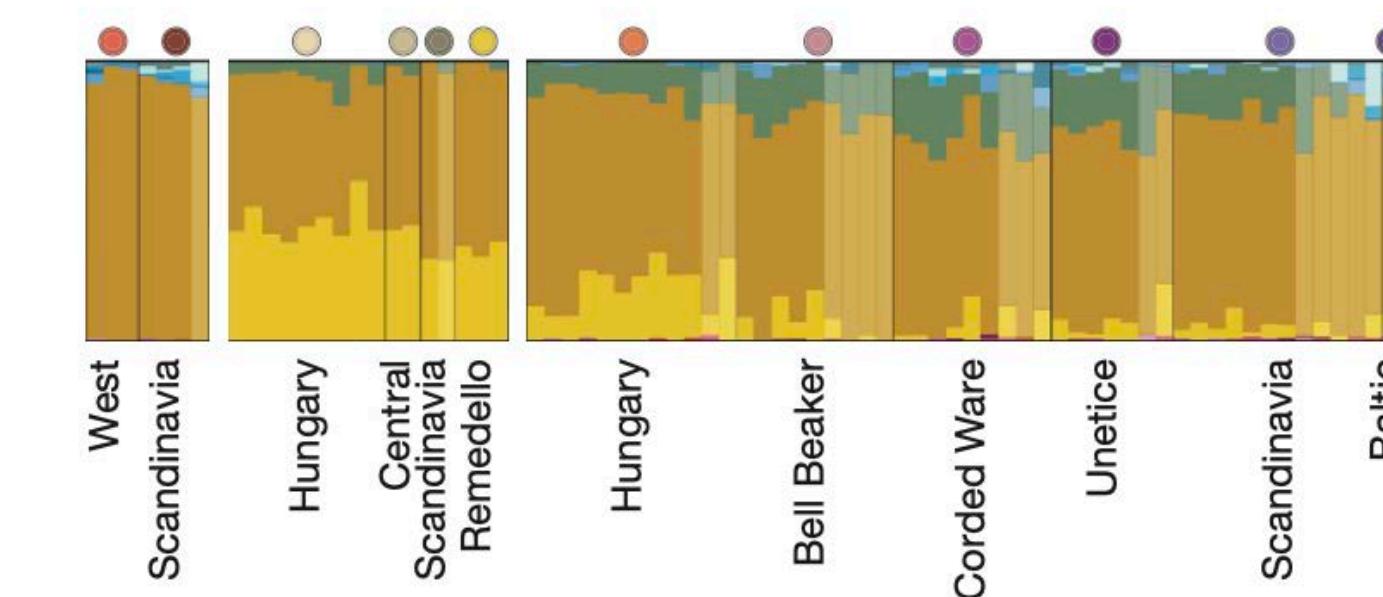


Population genetic analysis of ancient DNA data

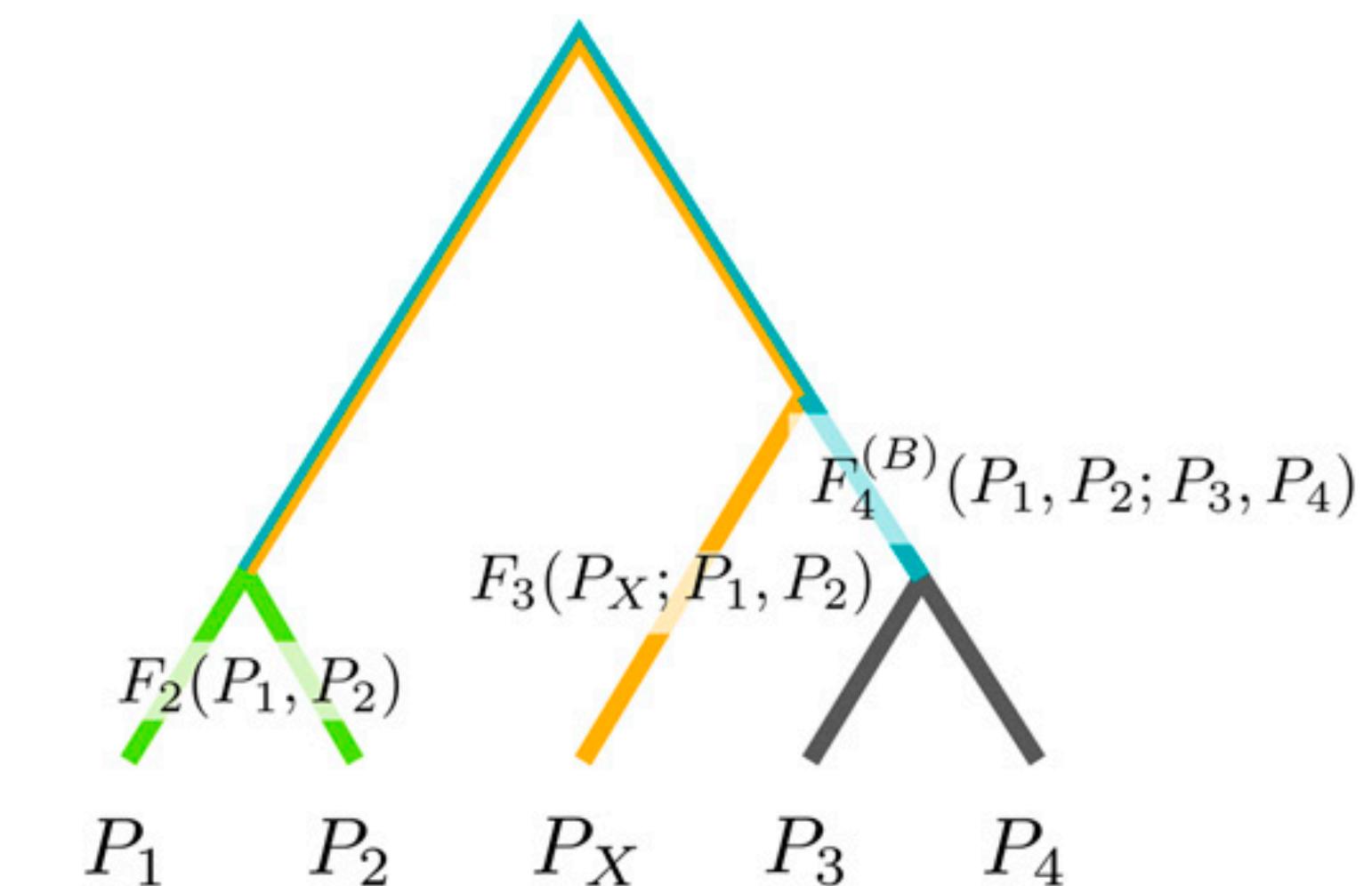
Principal component analysis



Model-based clustering

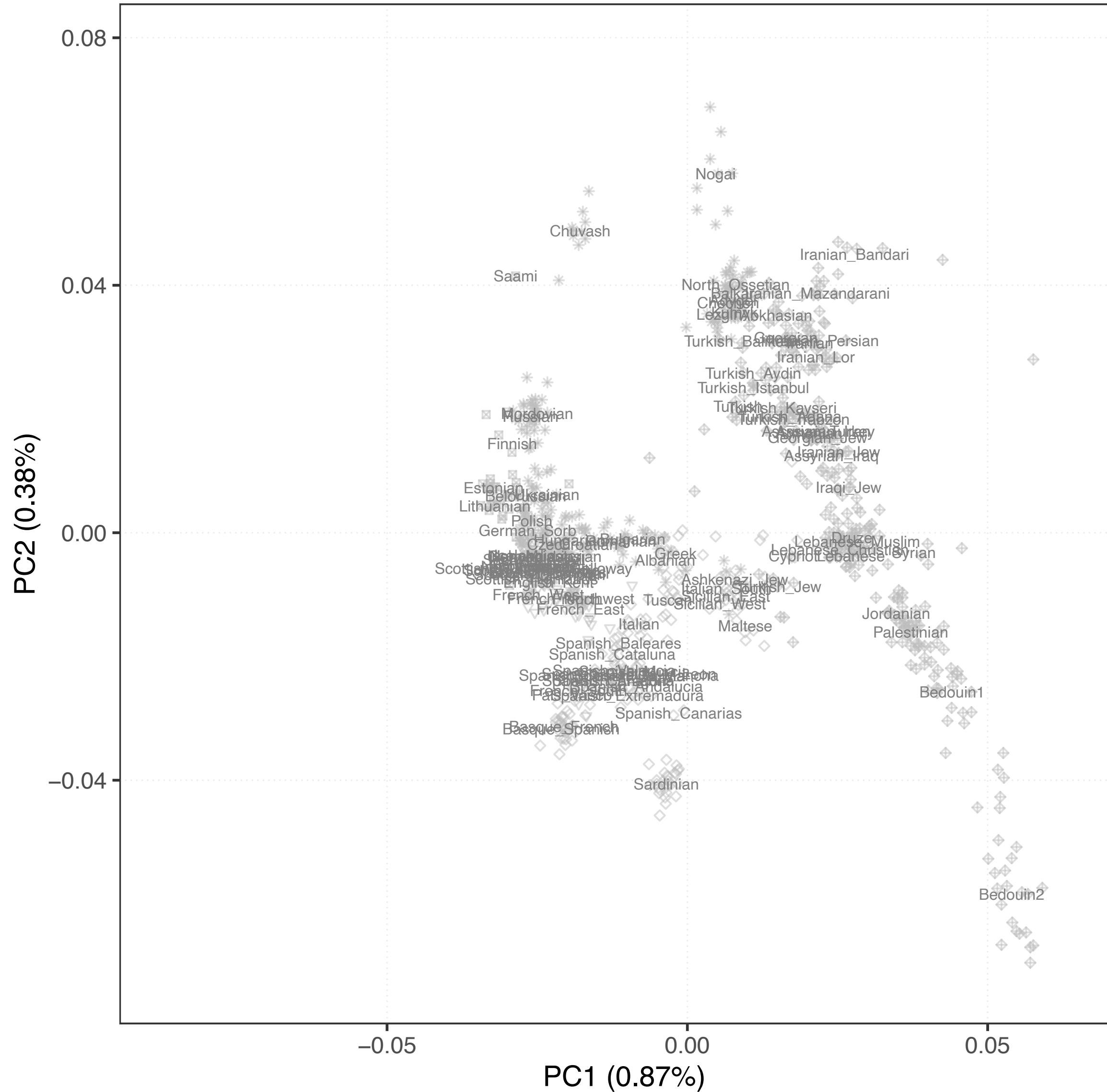


f-statistics

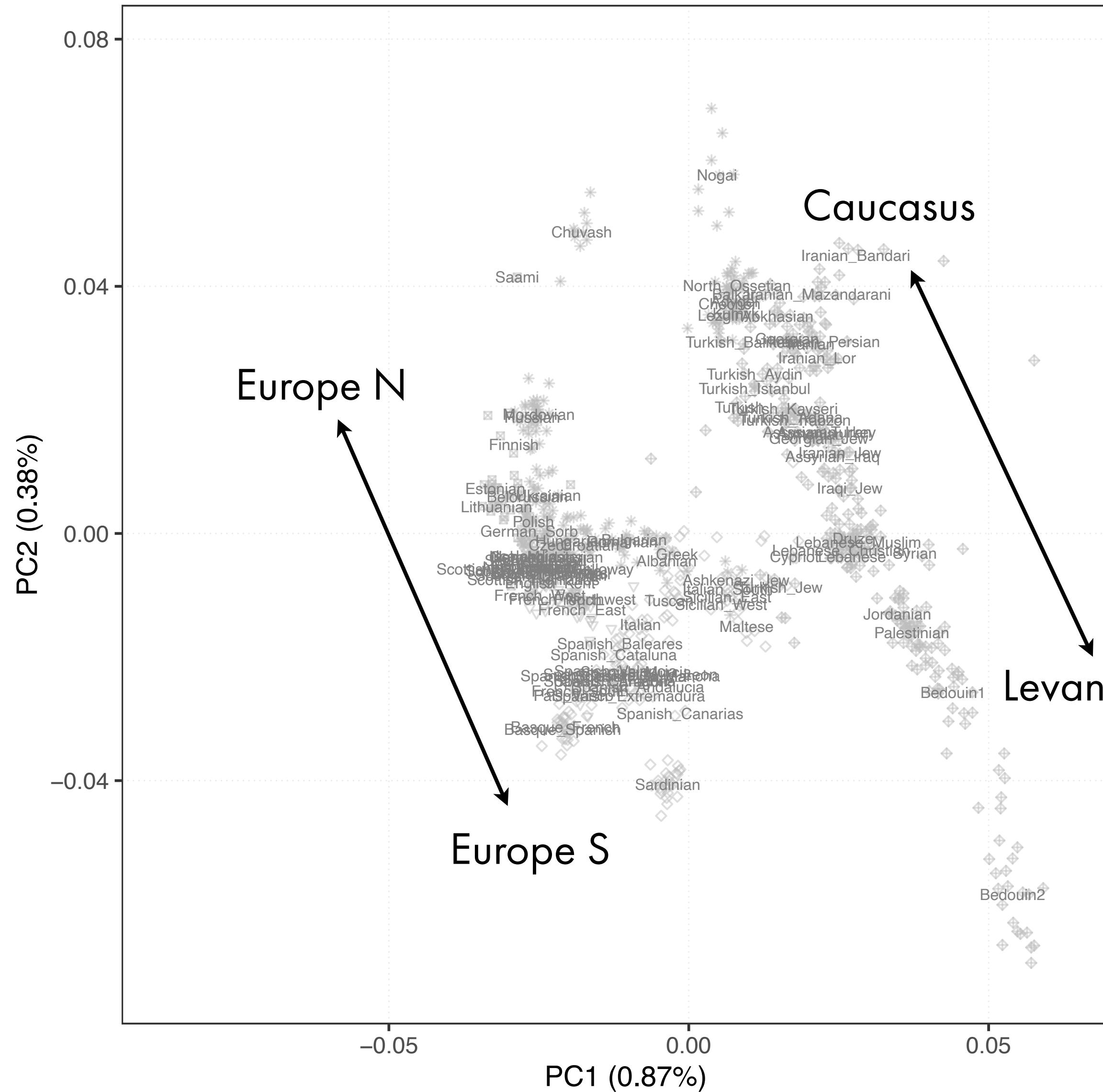


Allele-frequency based methods suited for low coverage / pseudo-haploid data

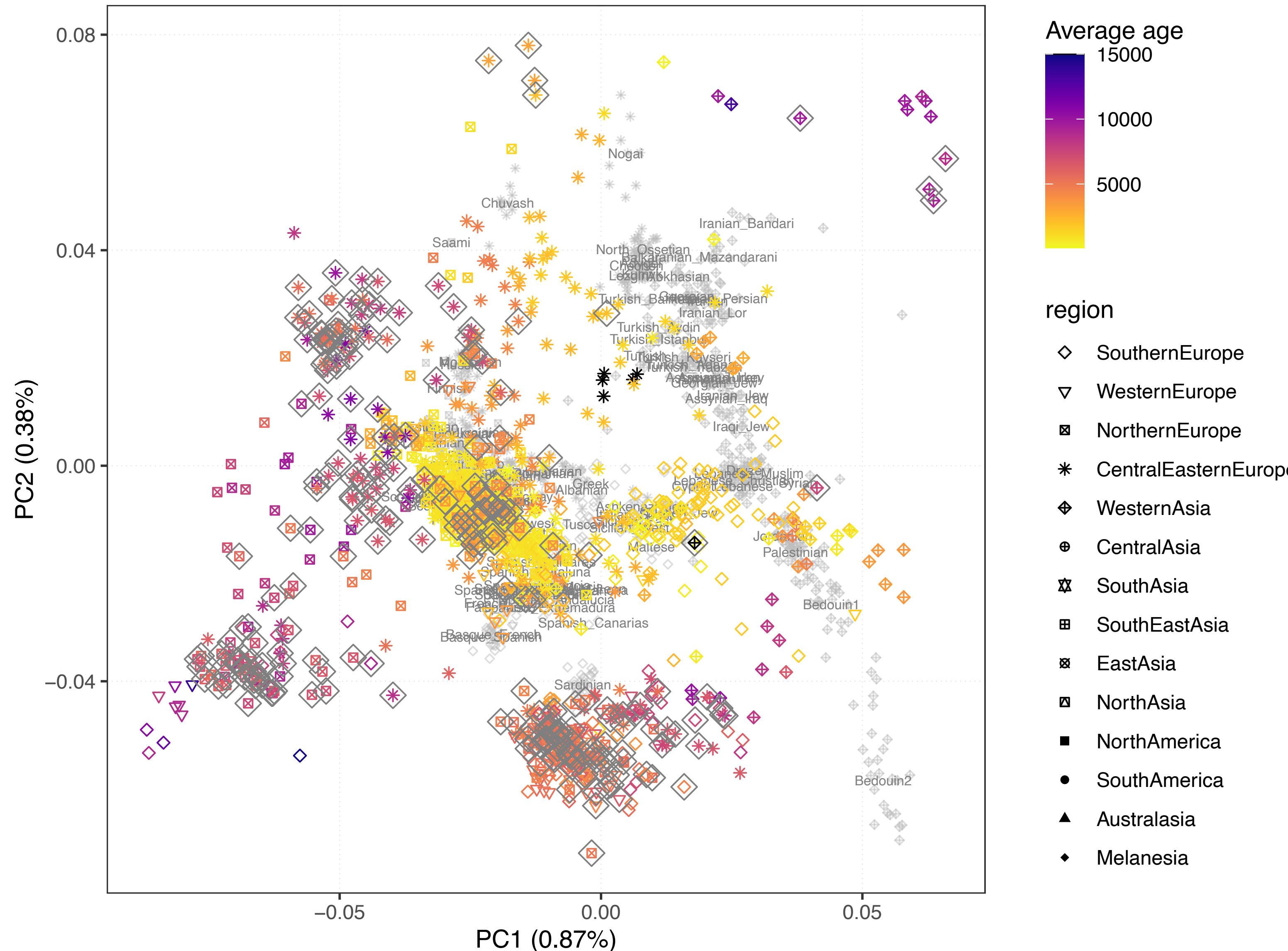
A PCA tour of western Eurasia



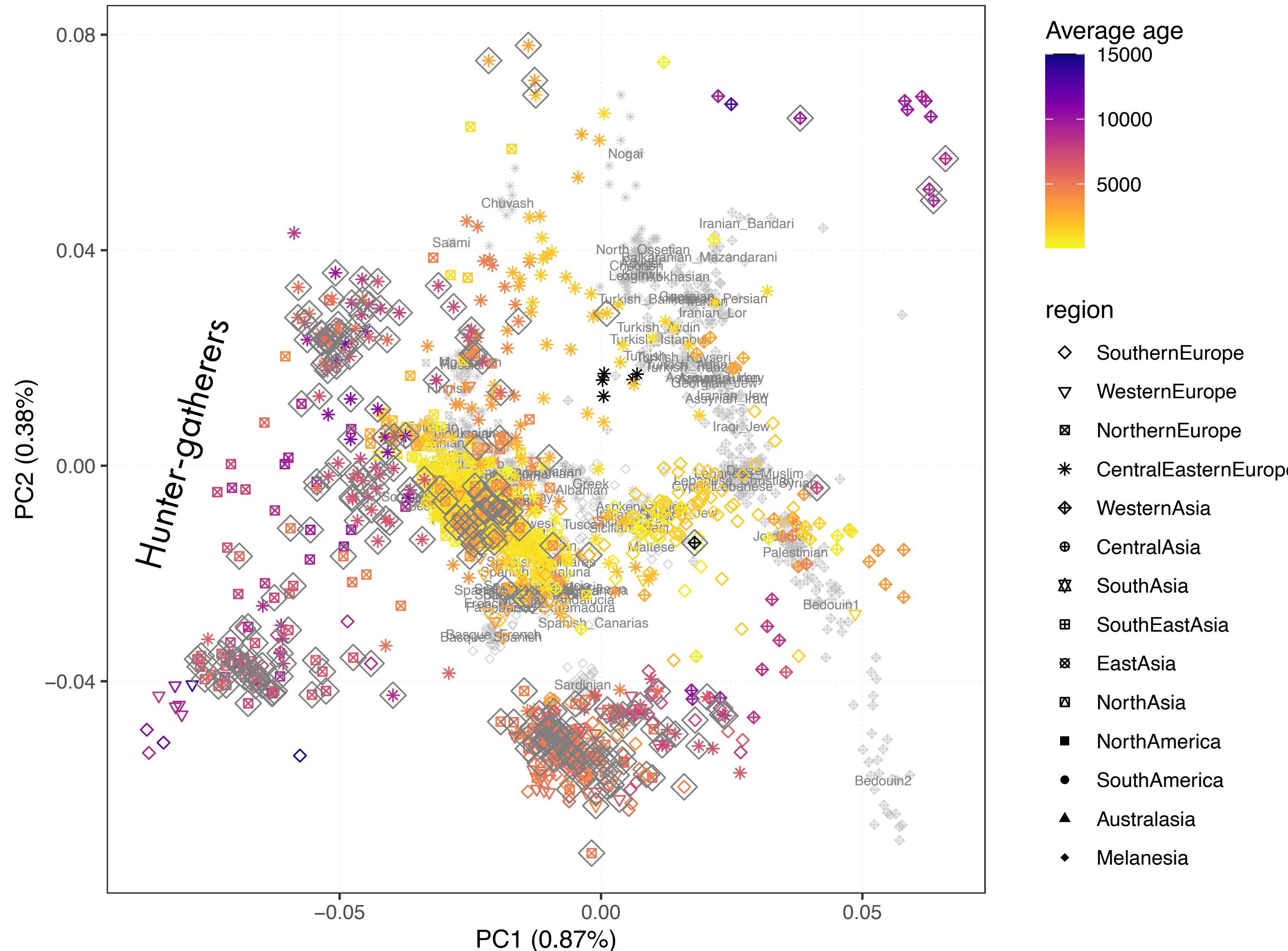
A PCA tour of western Eurasia



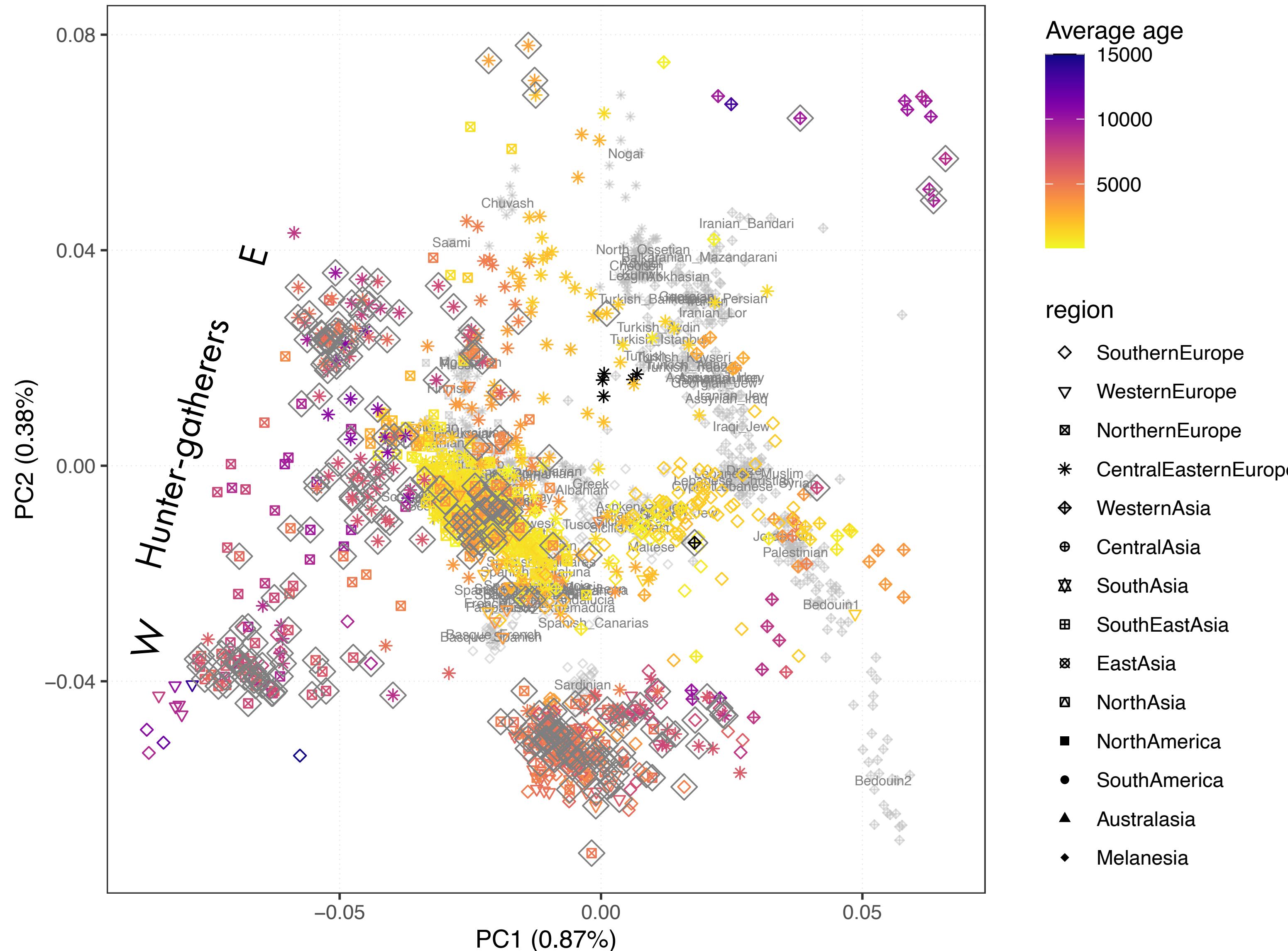
A PCA tour of western Eurasia



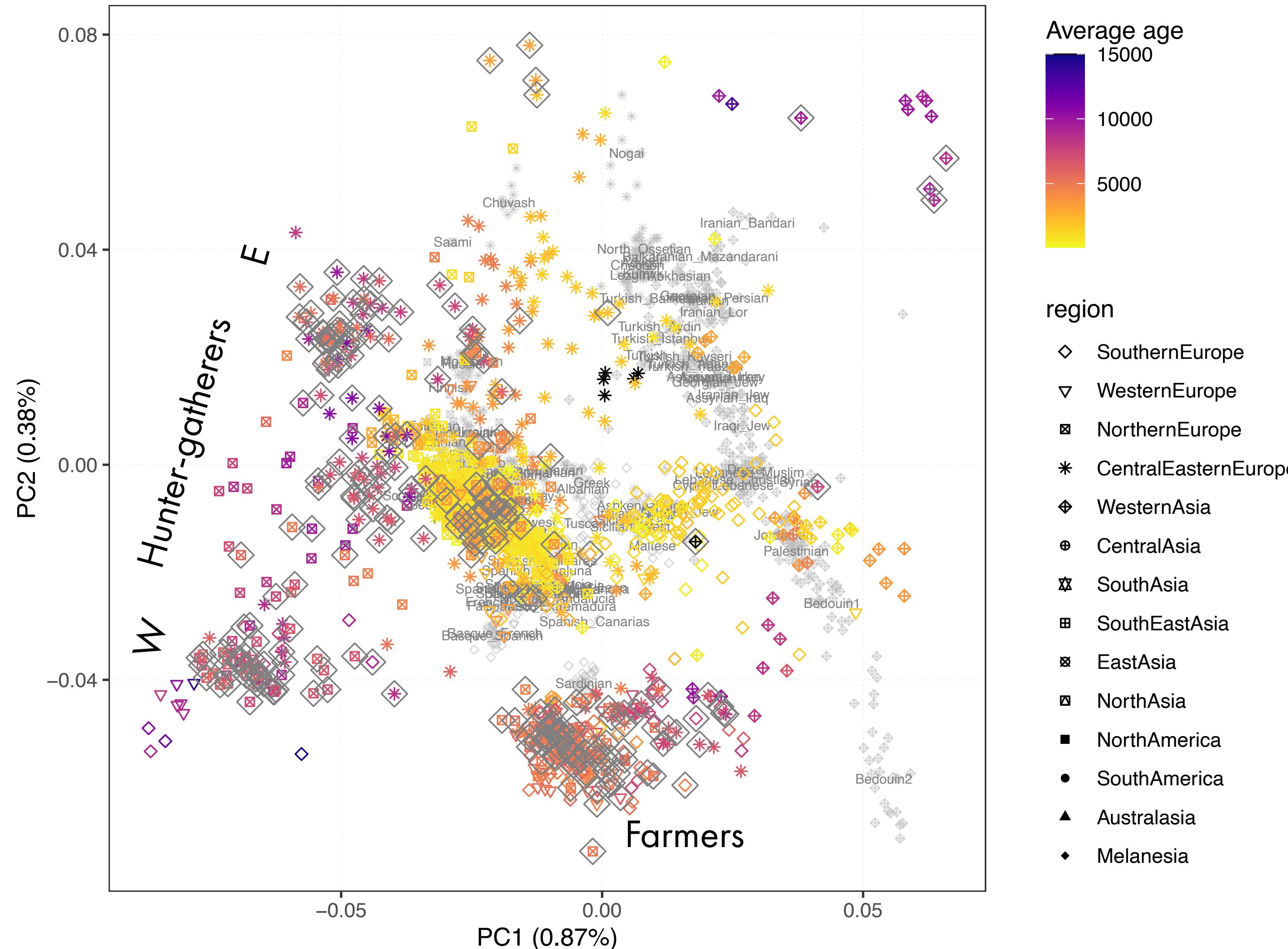
A PCA tour of western Eurasia



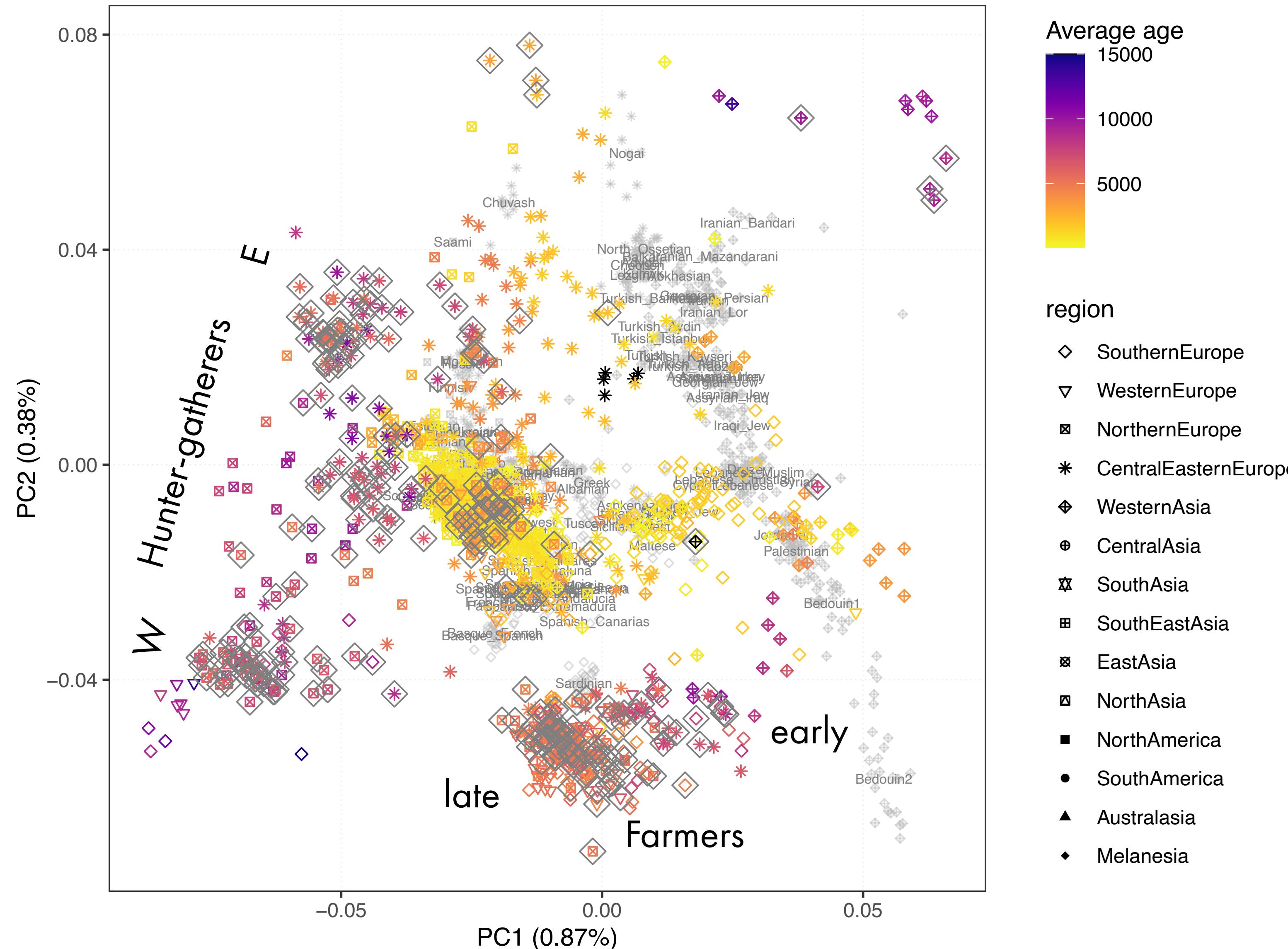
A PCA tour of western Eurasia



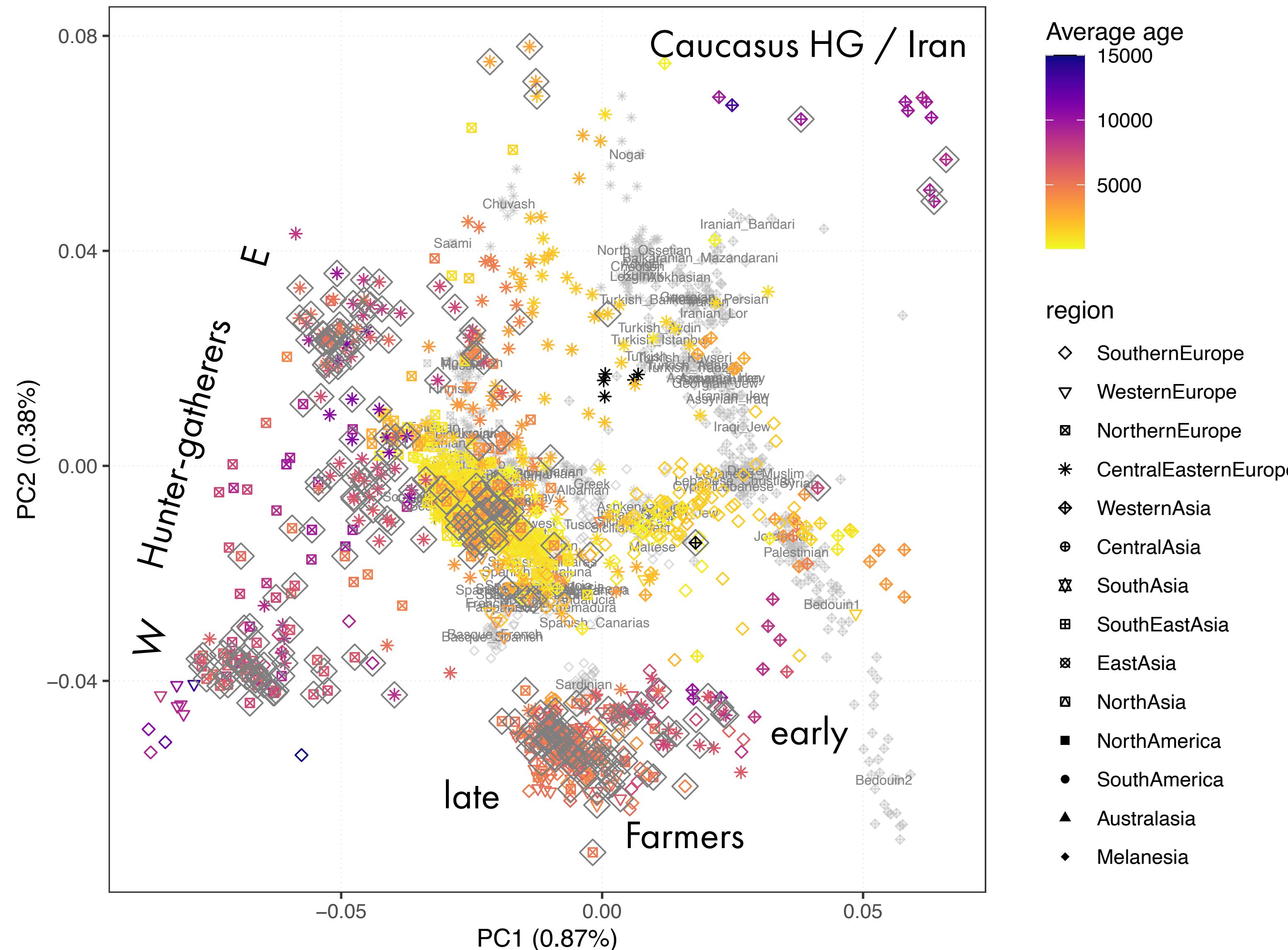
A PCA tour of western Eurasia



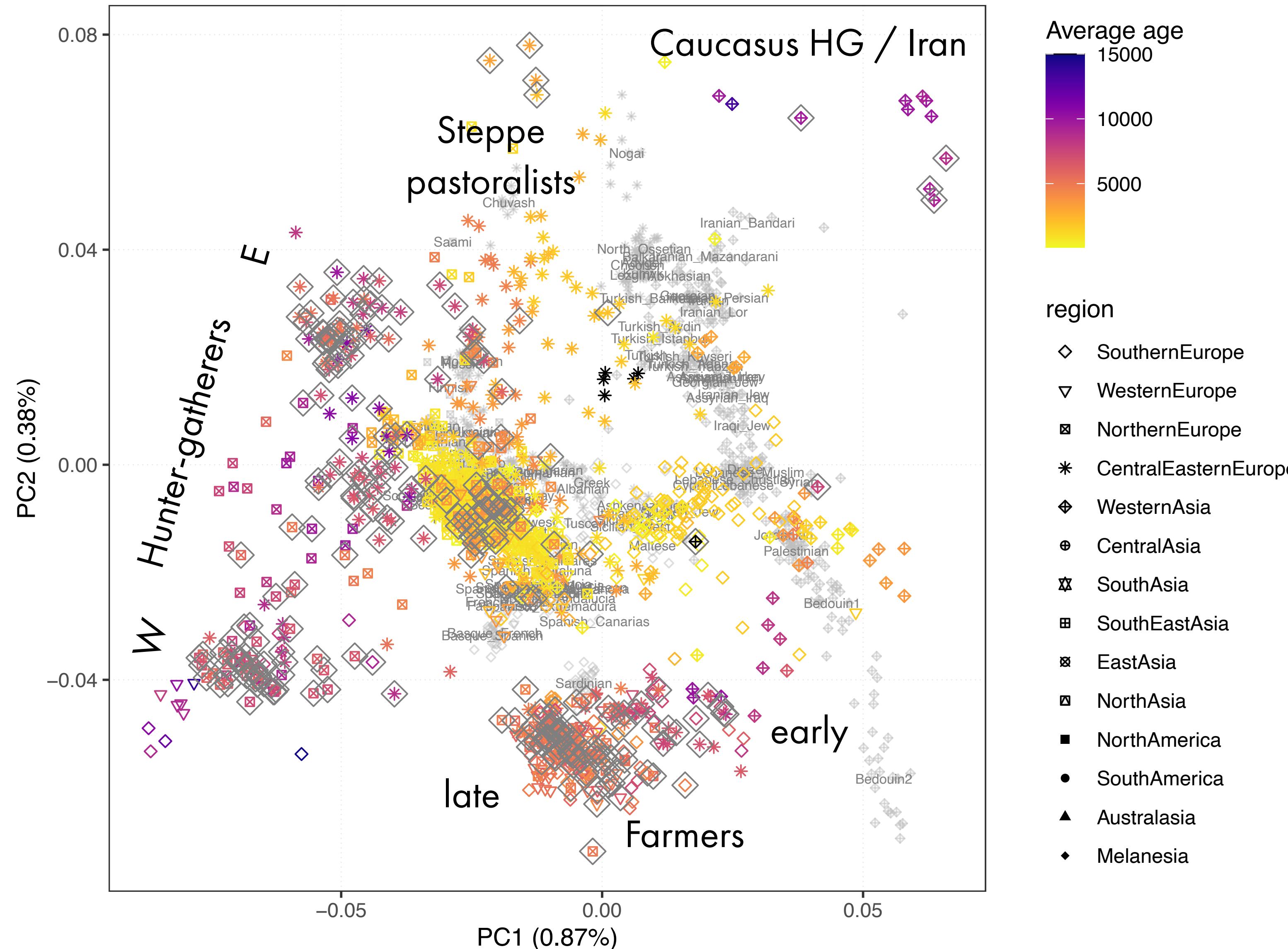
A PCA tour of western Eurasia



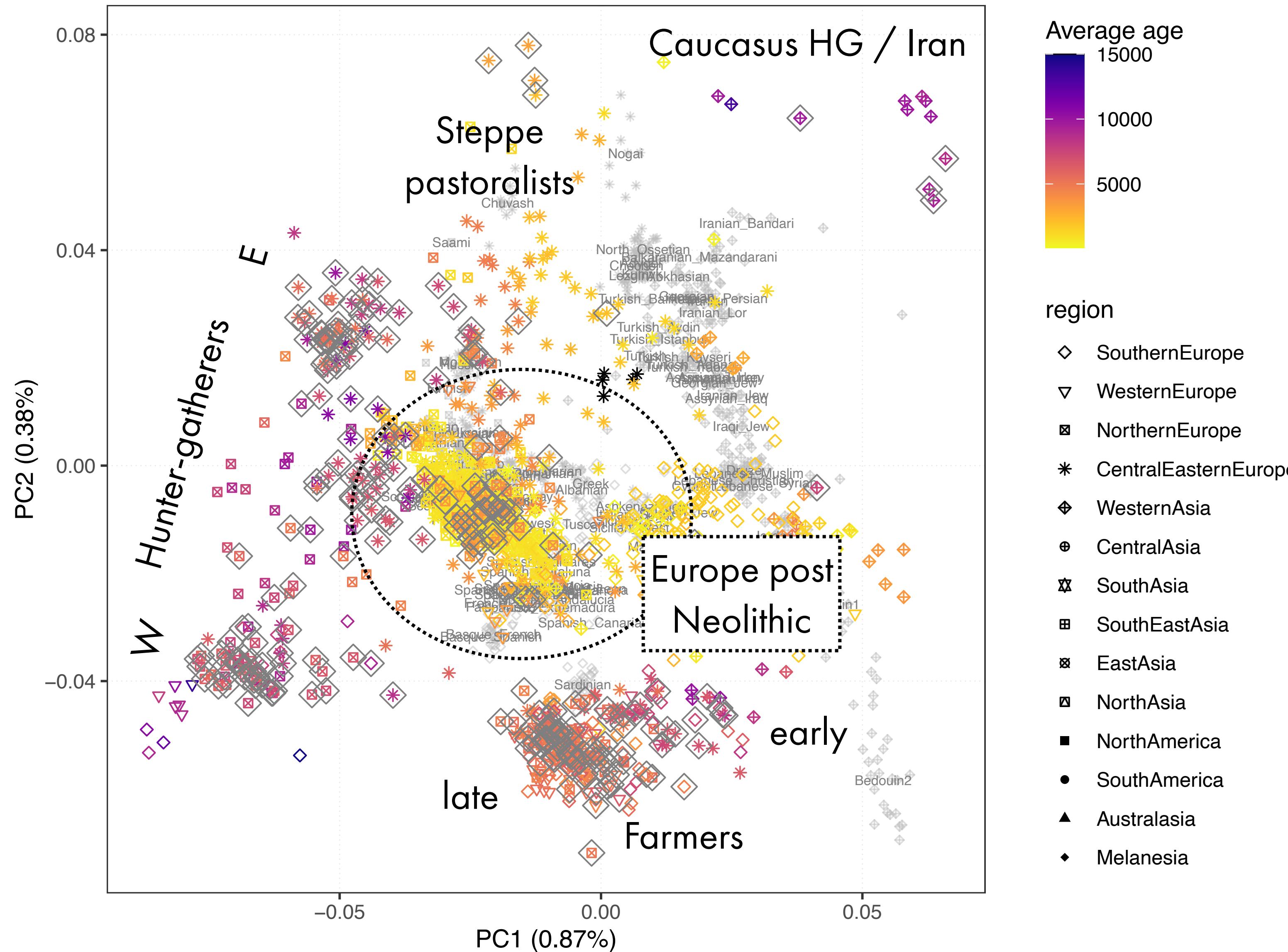
A PCA tour of western Eurasia



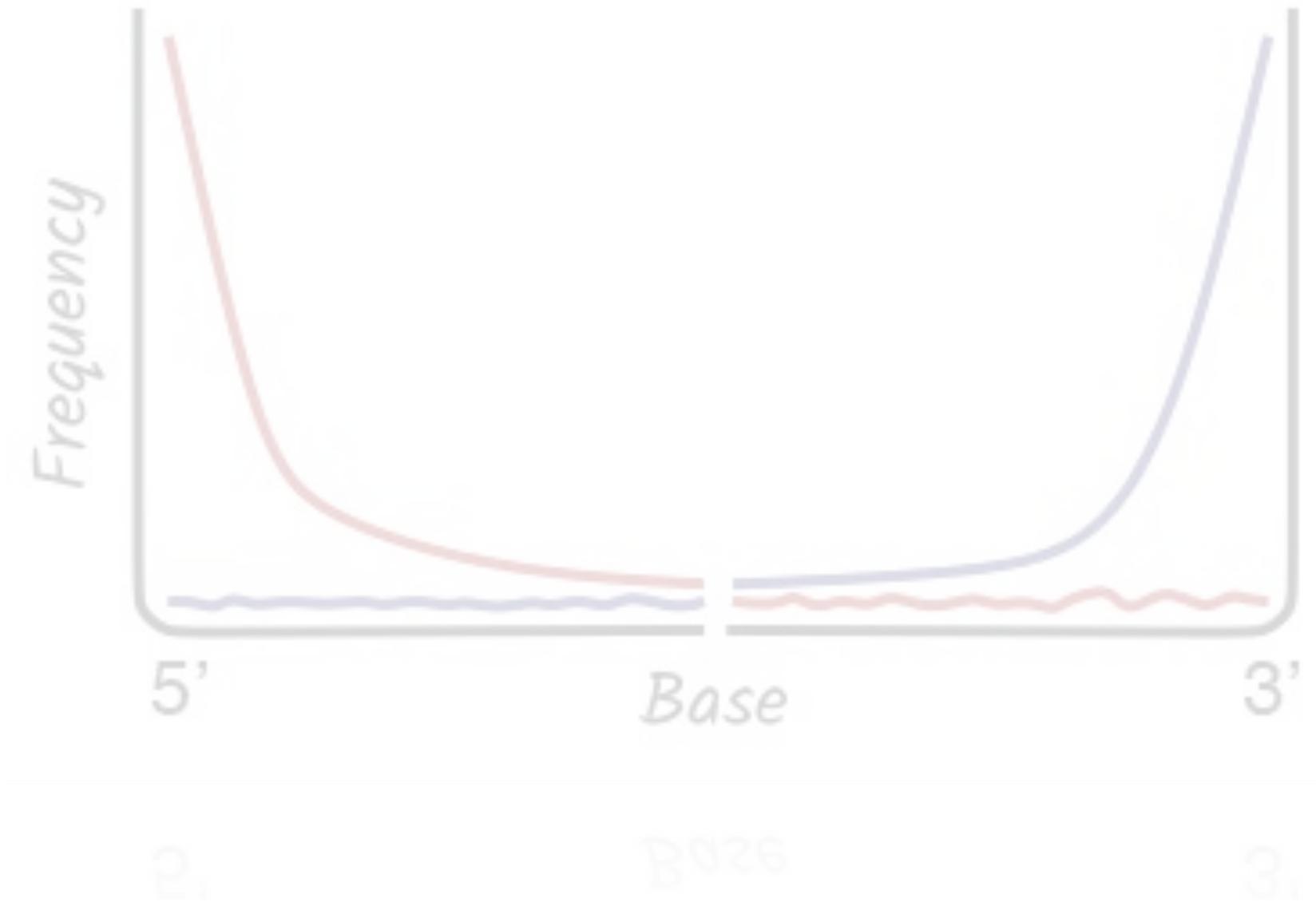
A PCA tour of western Eurasia



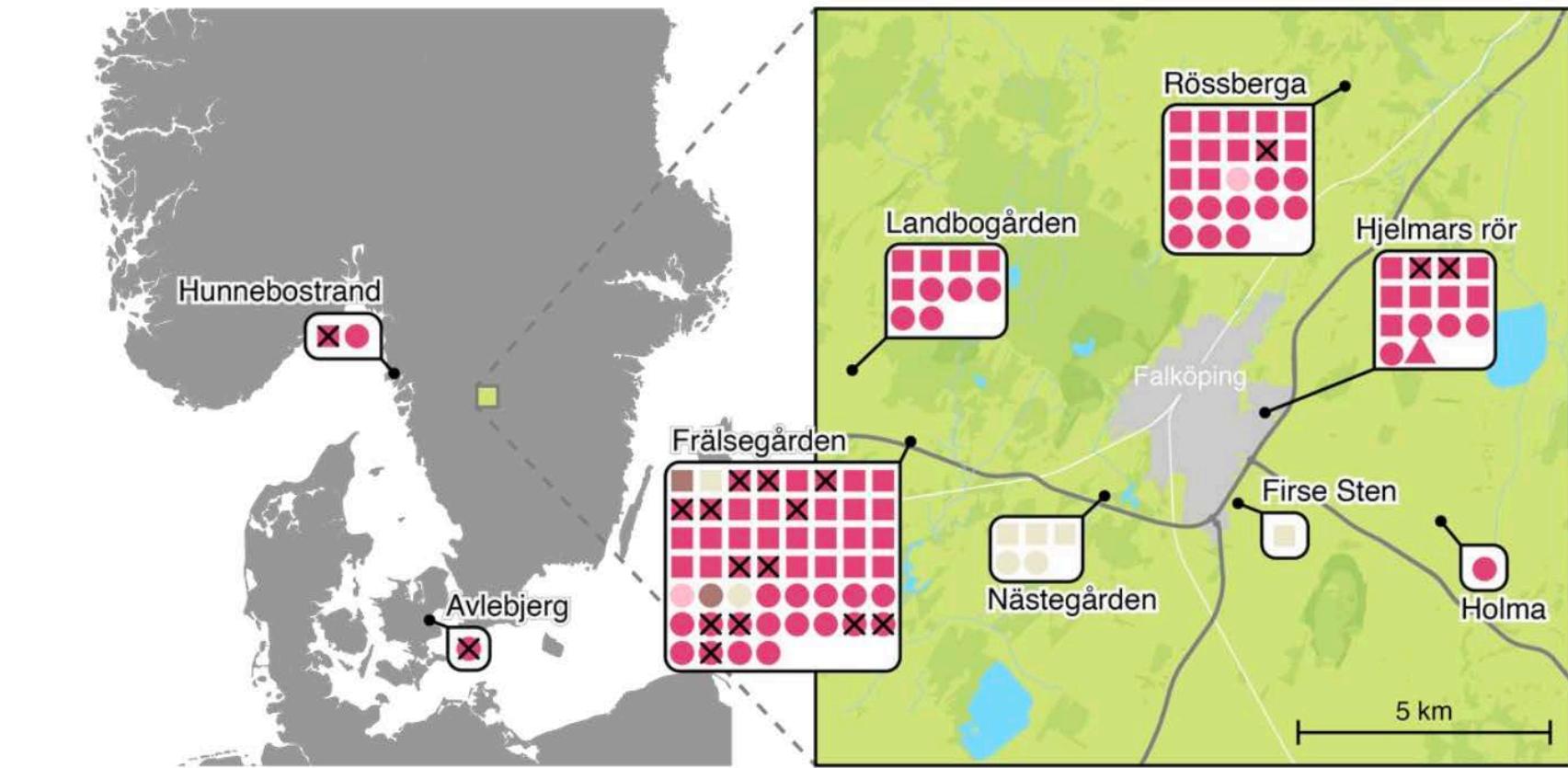
A PCA tour of western Eurasia



Topics for today's lecture



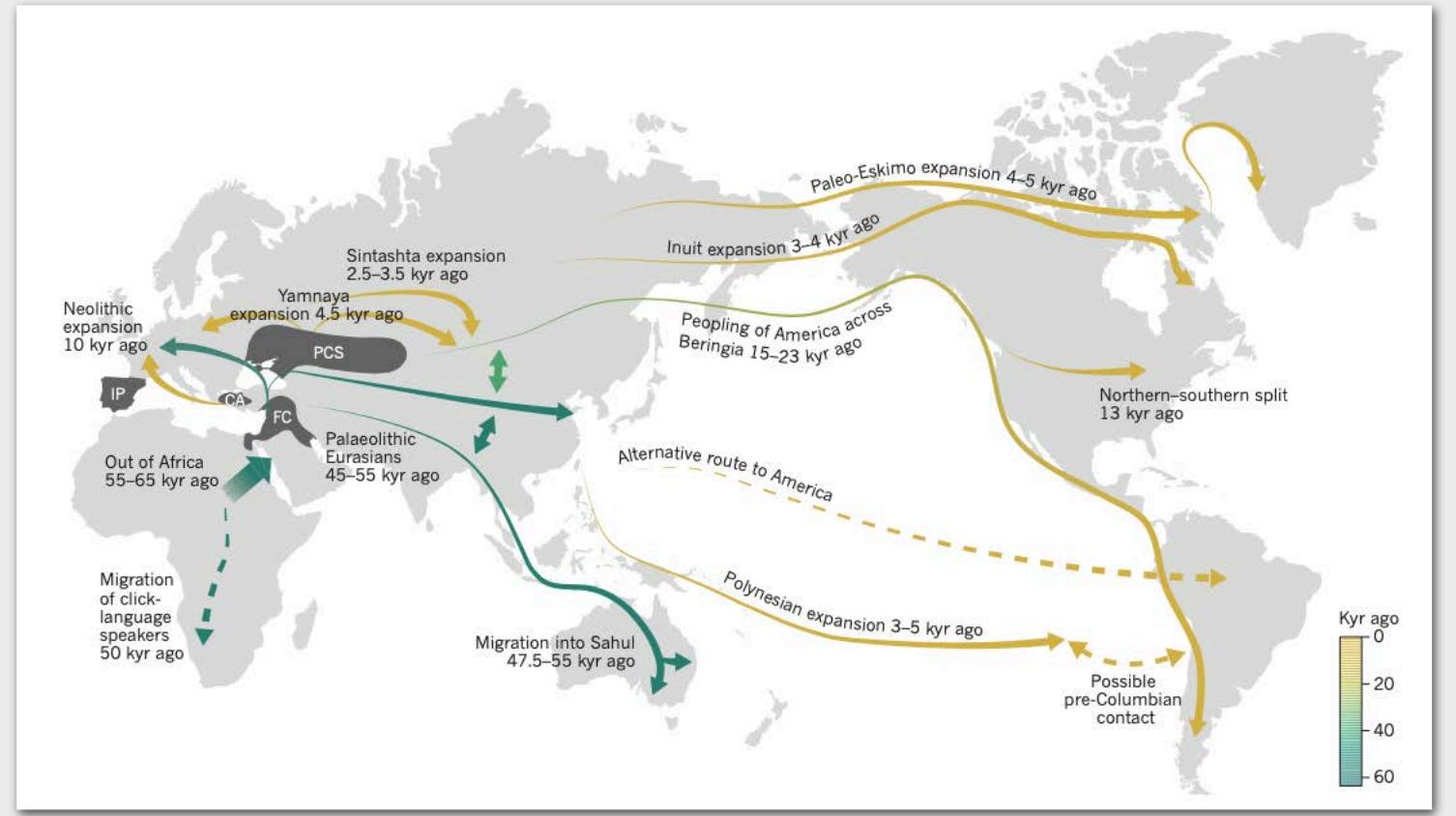
Ancient DNA primer
Characteristics and challenges



Ancient DNA showcase
Kinship and plague in Stone Age Scandinavia



Demography

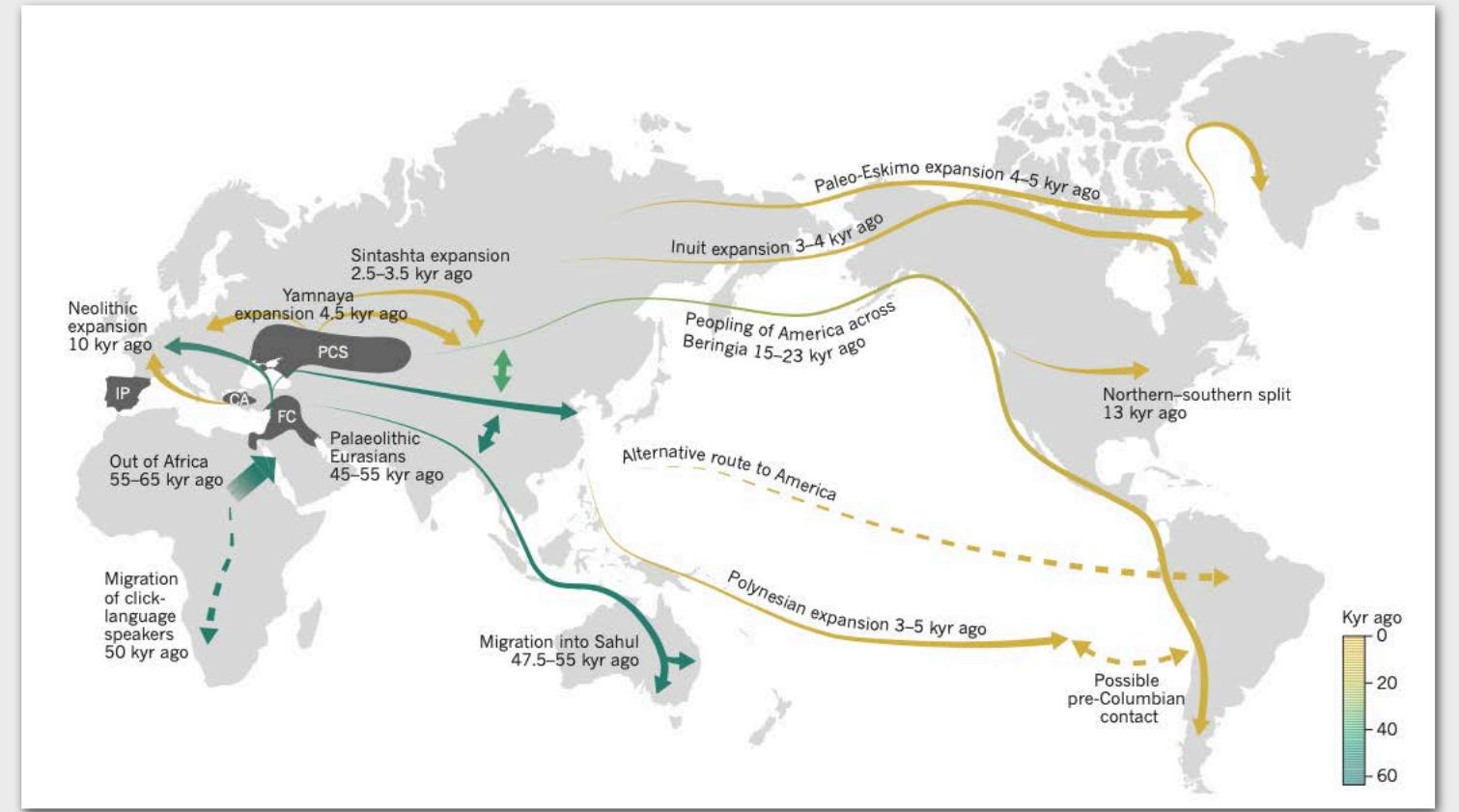


Social organisation

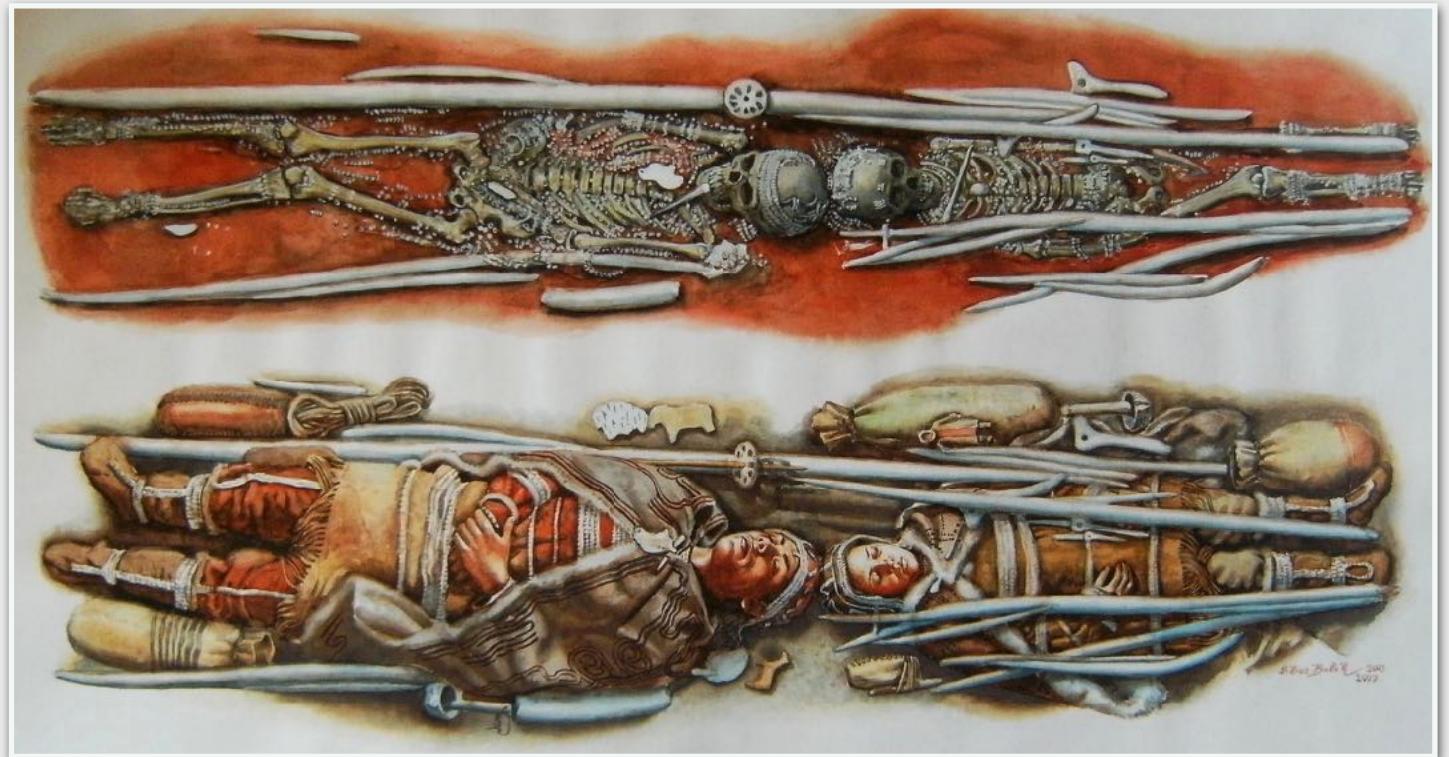


Endogenous host DNA

Demography



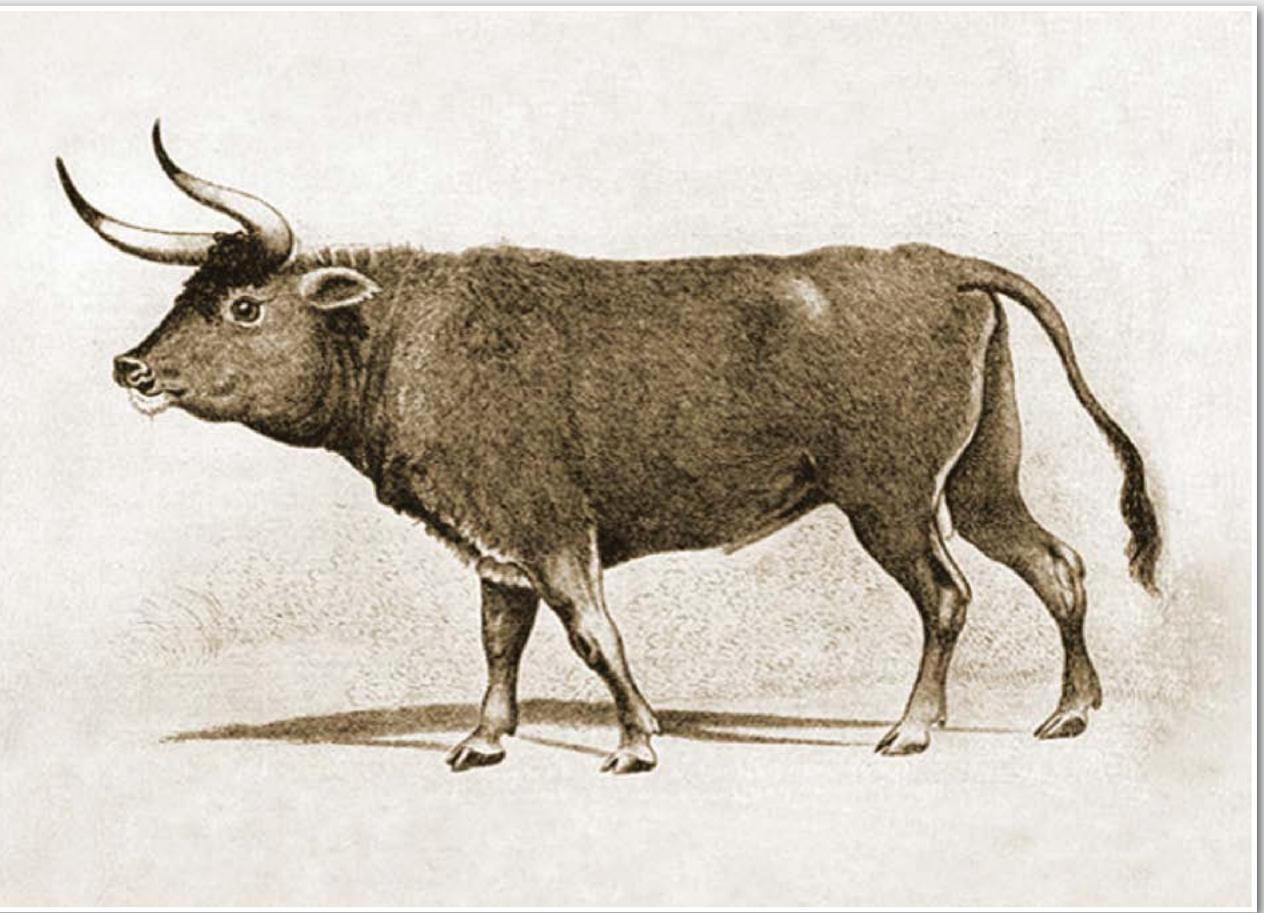
Social organisation



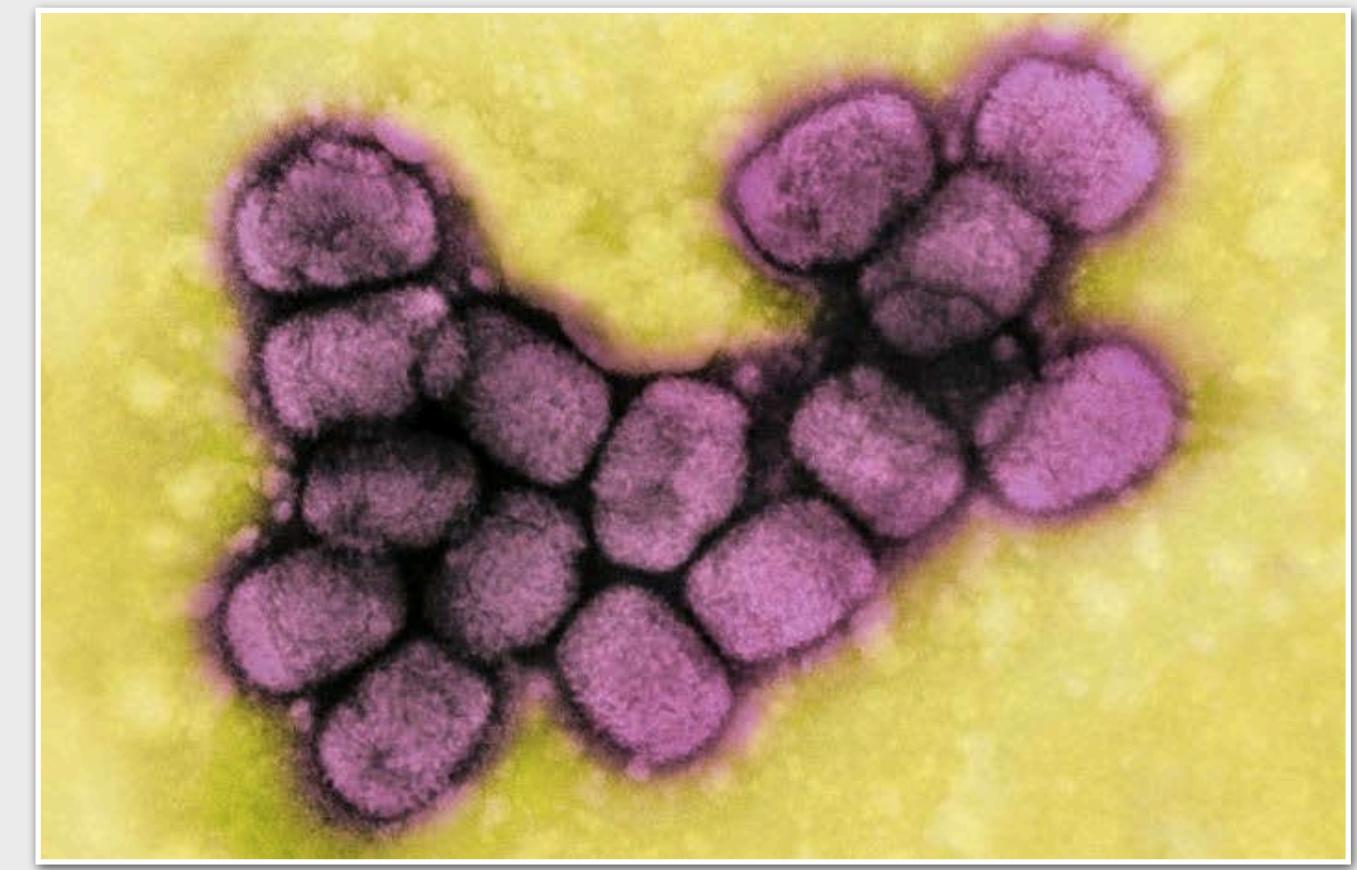
Endogenous host DNA



Non-host DNA



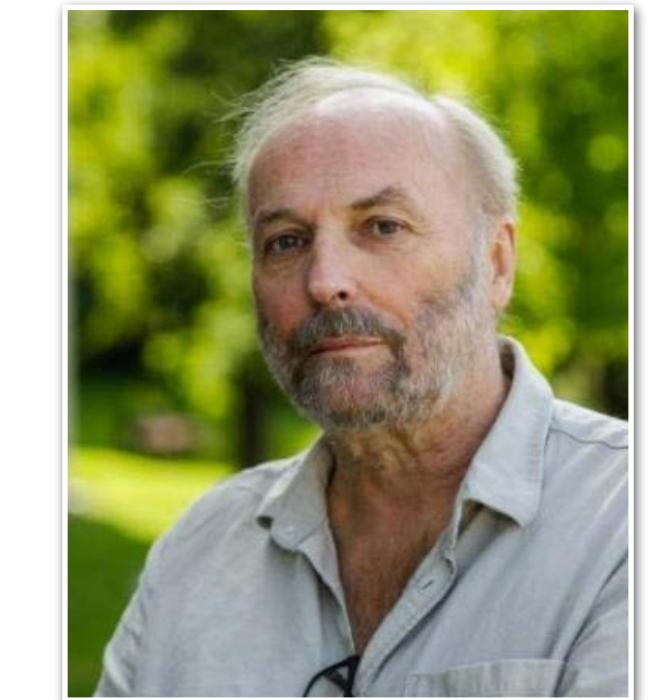
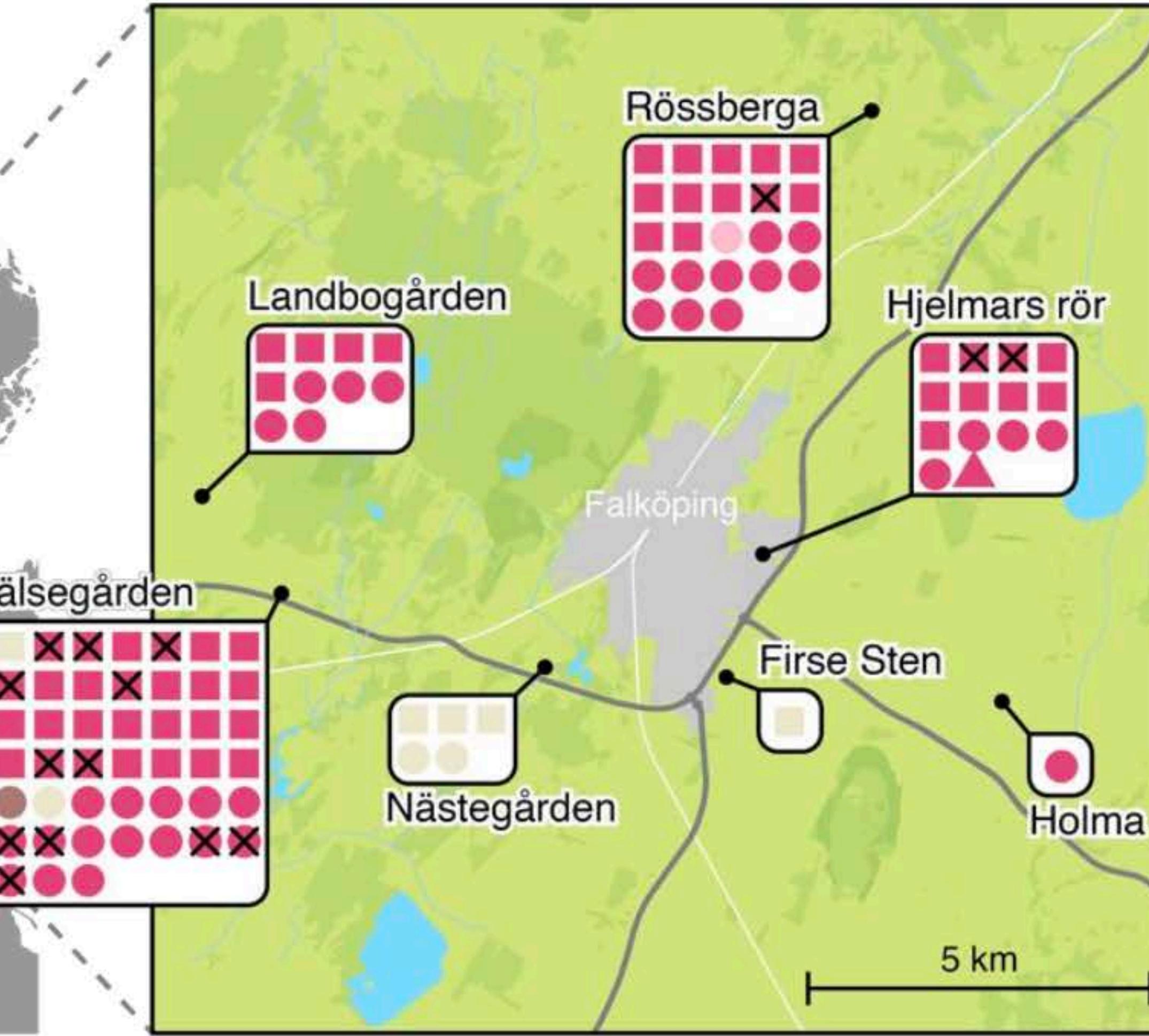
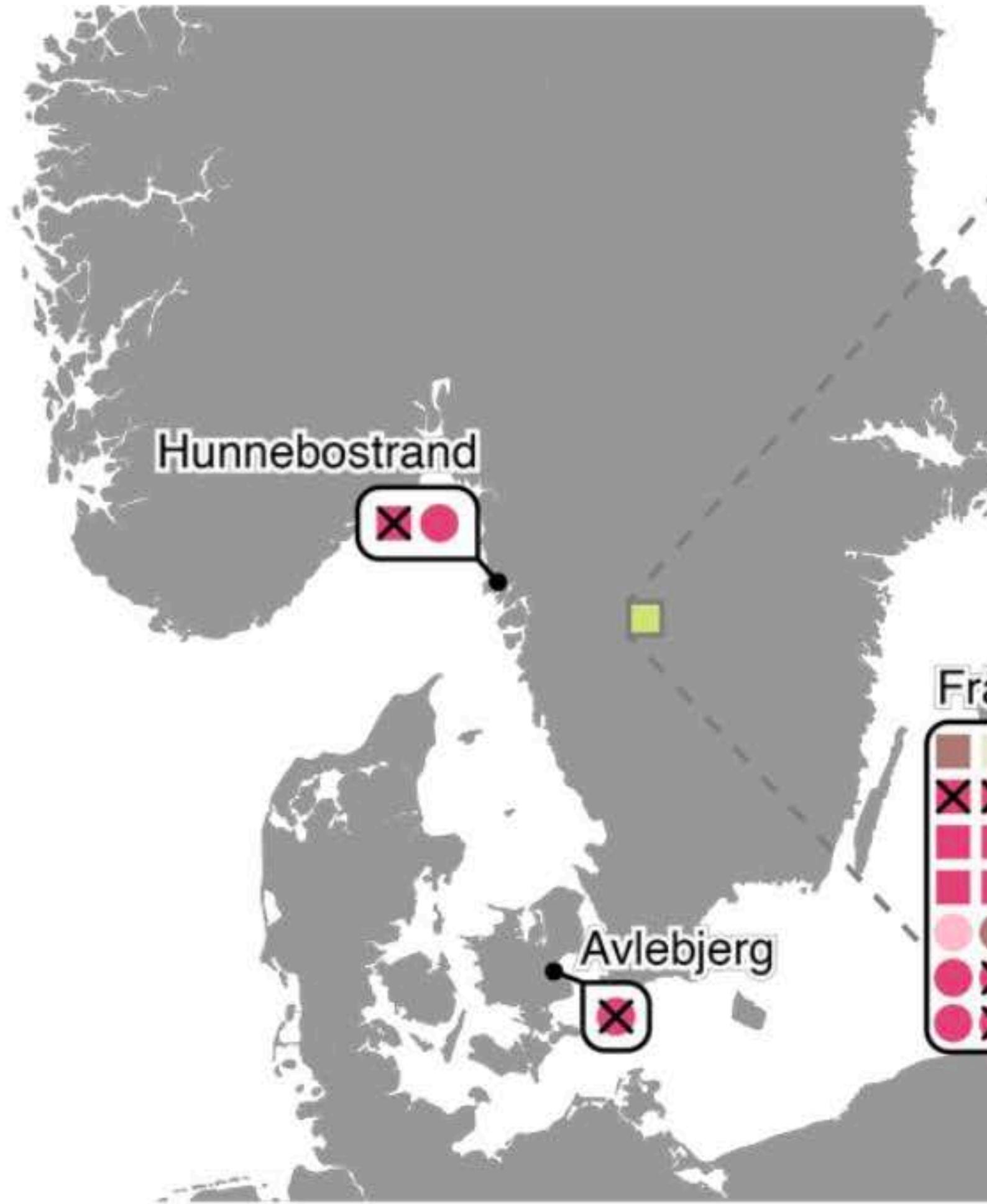
Infectious diseases



Diet and microbiome

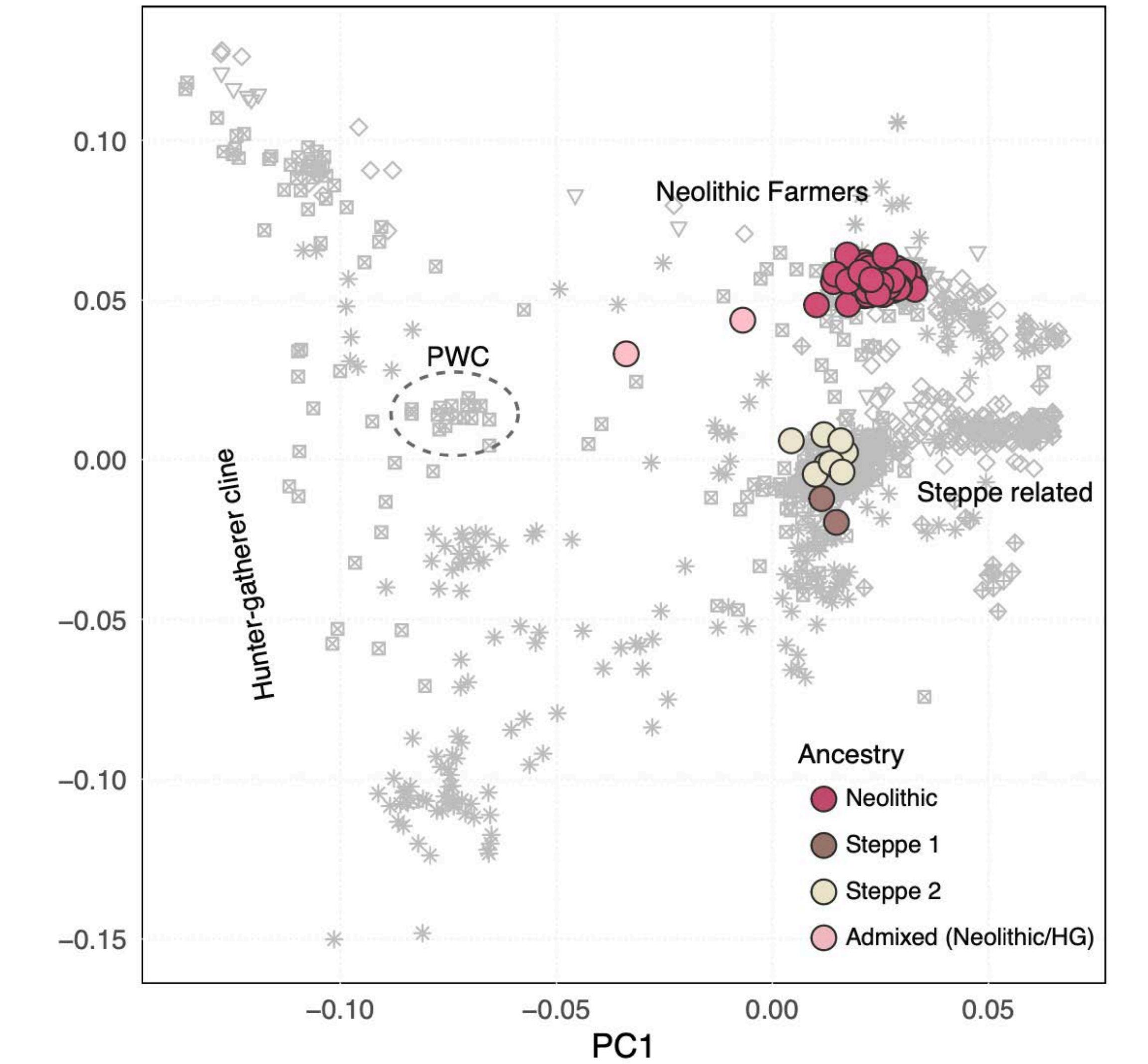
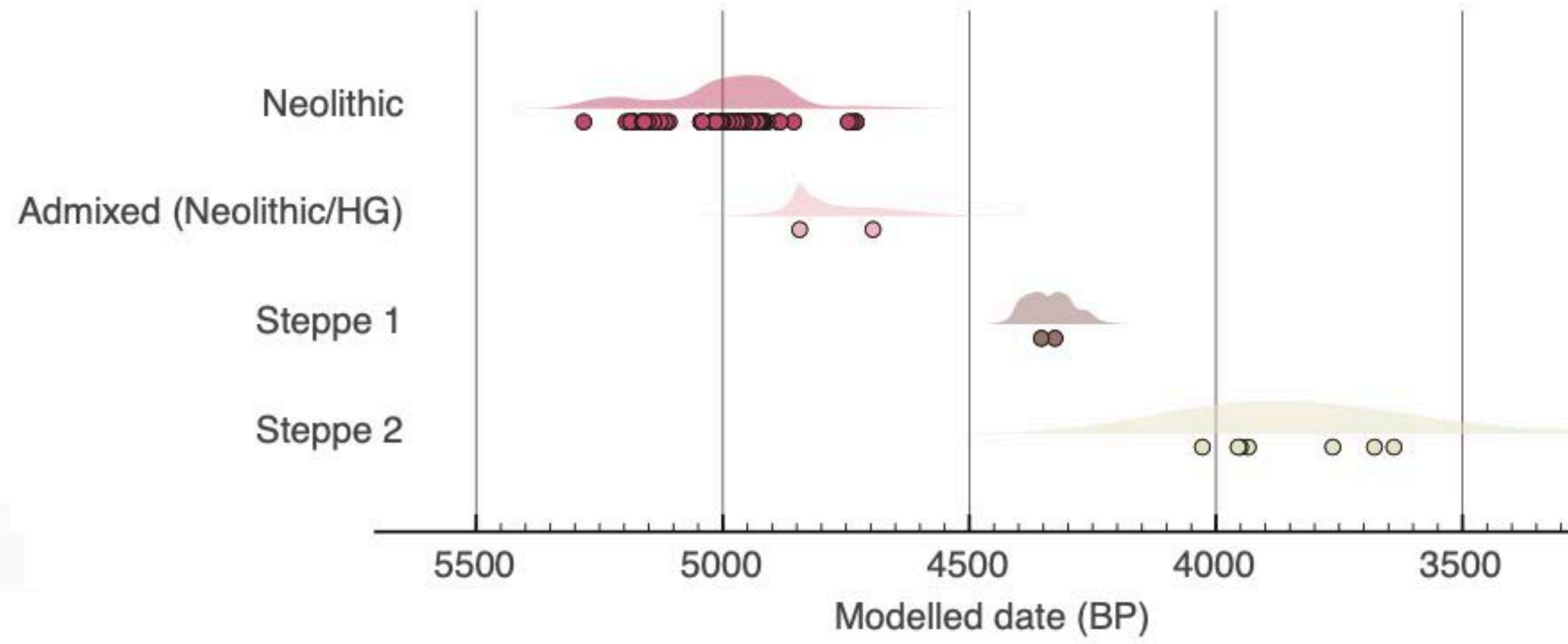


Population genomics of late Neolithic Scandinavia



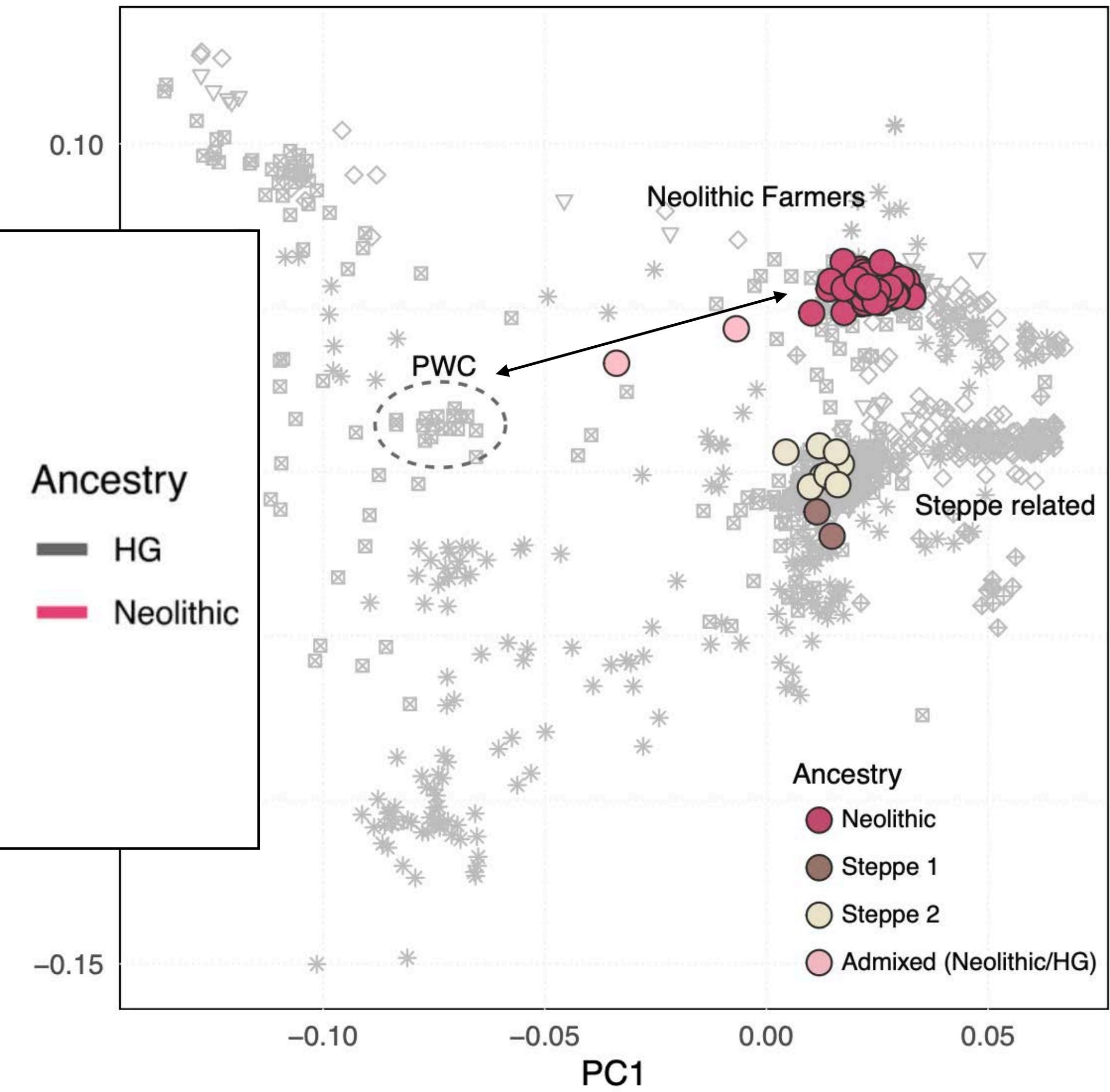
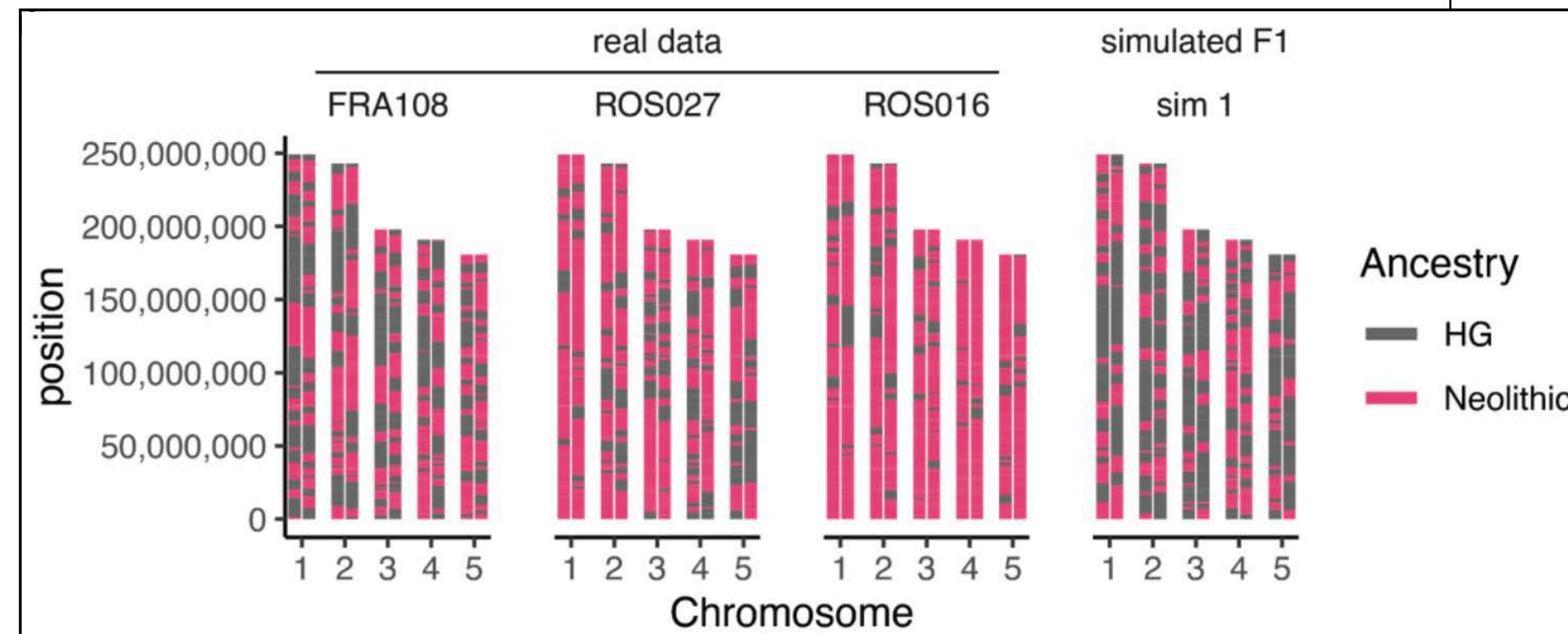
109 ancient human genomes from megalithic graves in southern Scandinavia

Genetic diversity of human populations



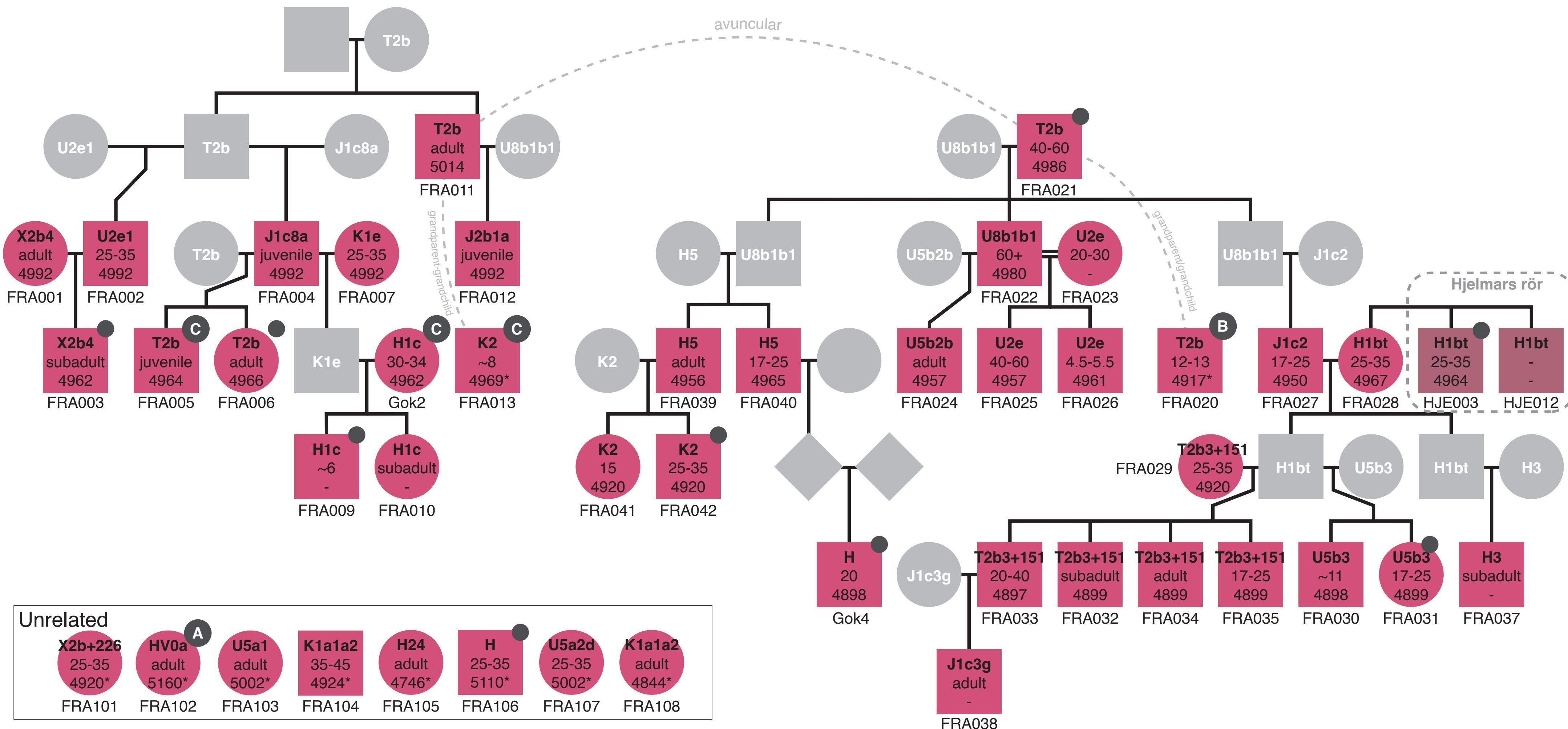
Repeated use of megalithic graves by peoples with different ancestries

Recent admixture with Pitted-ware culture hunter-gatherers



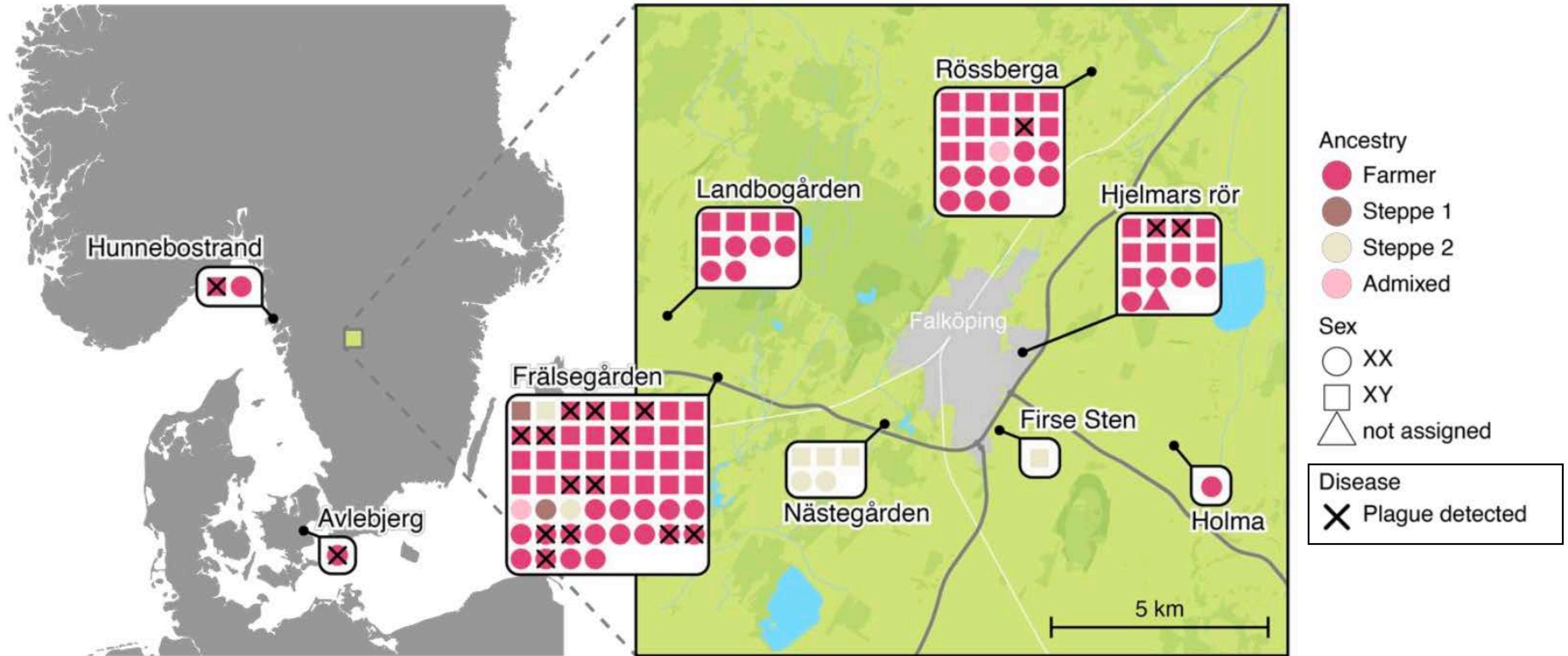
Likely first- and second generation offspring at Fralsegarden and Rossberga

A five generation pedigree of Neolithic farmers



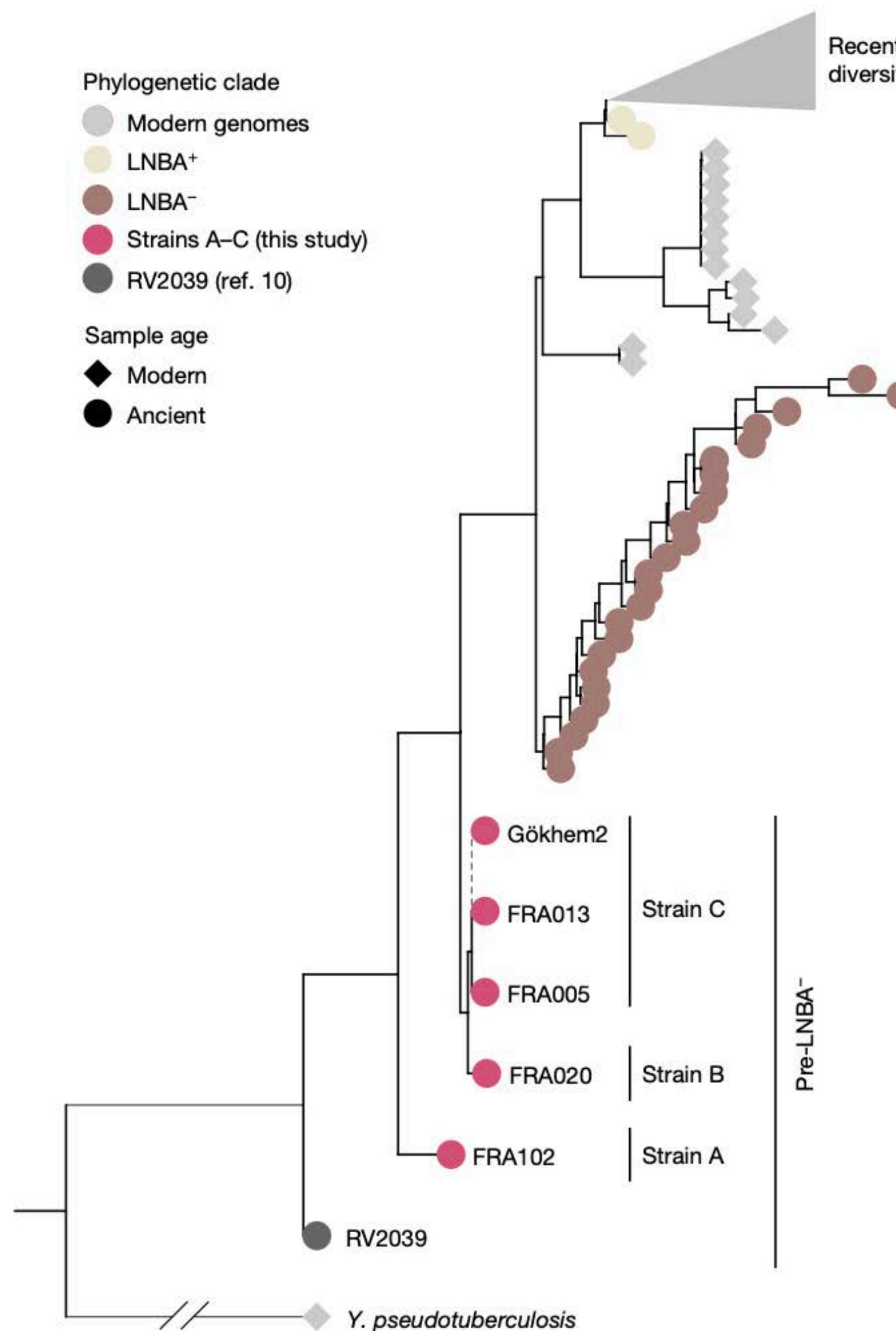
Pedigree of 38 sequenced individuals suggests patrilineal and patrilocal social organisation

High prevalence of plague in late Neolithic Scandinavia

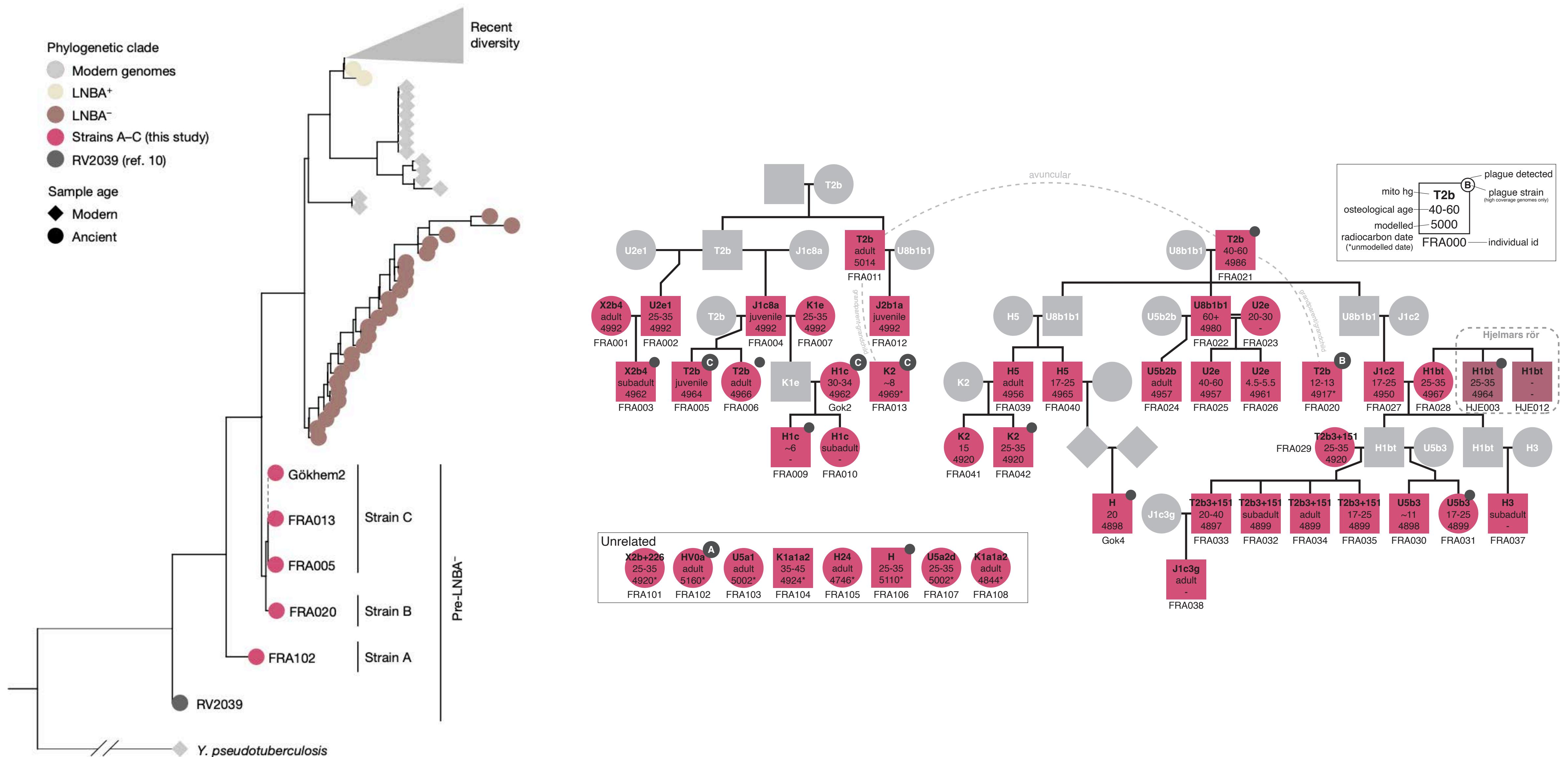


Plague detected in ~17% of individuals

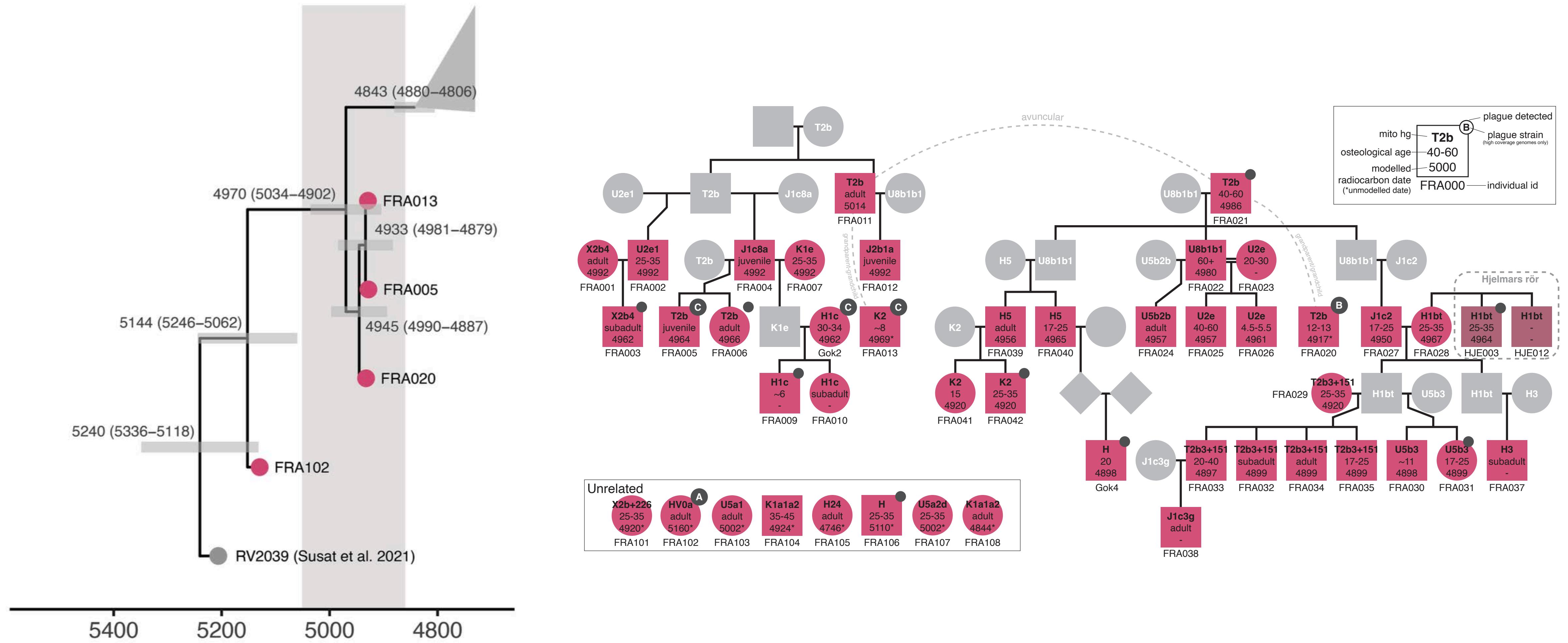
Multiple plague strains in Neolithic farmer families



Multiple plague strains in Neolithic farmer families

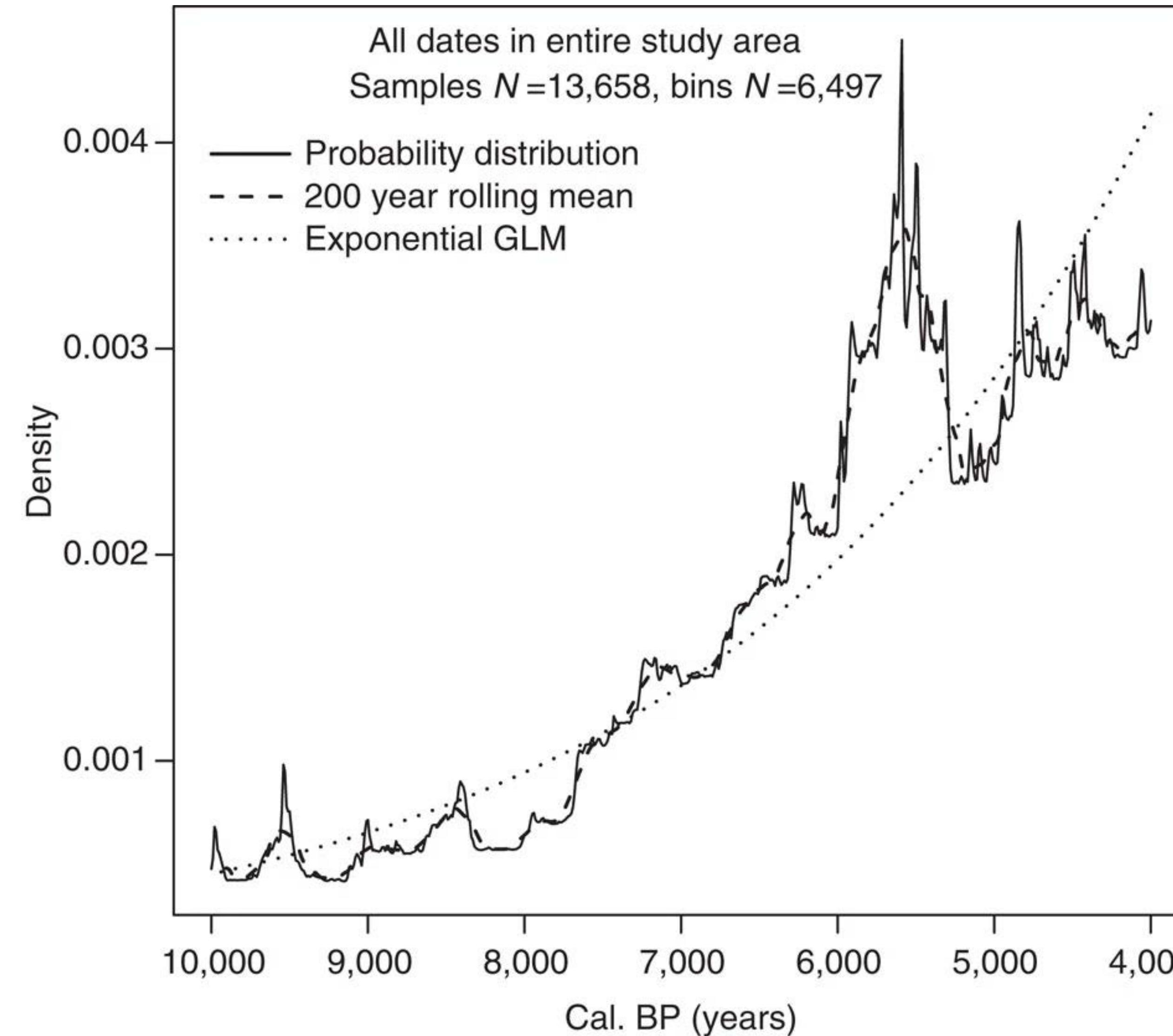


Multiple plague strains in Neolithic farmer families



Divergence of strains A and B during Fralsegarden pedigree chronology

Did plague play a role in the Neolithic decline?



Novel sources for ancient DNA

ARTICLE

<https://doi.org/10.1038/s41467-019-13549-9>

OPEN

A 5700 year-old human genome and oral microbiome from chewed birch pitch

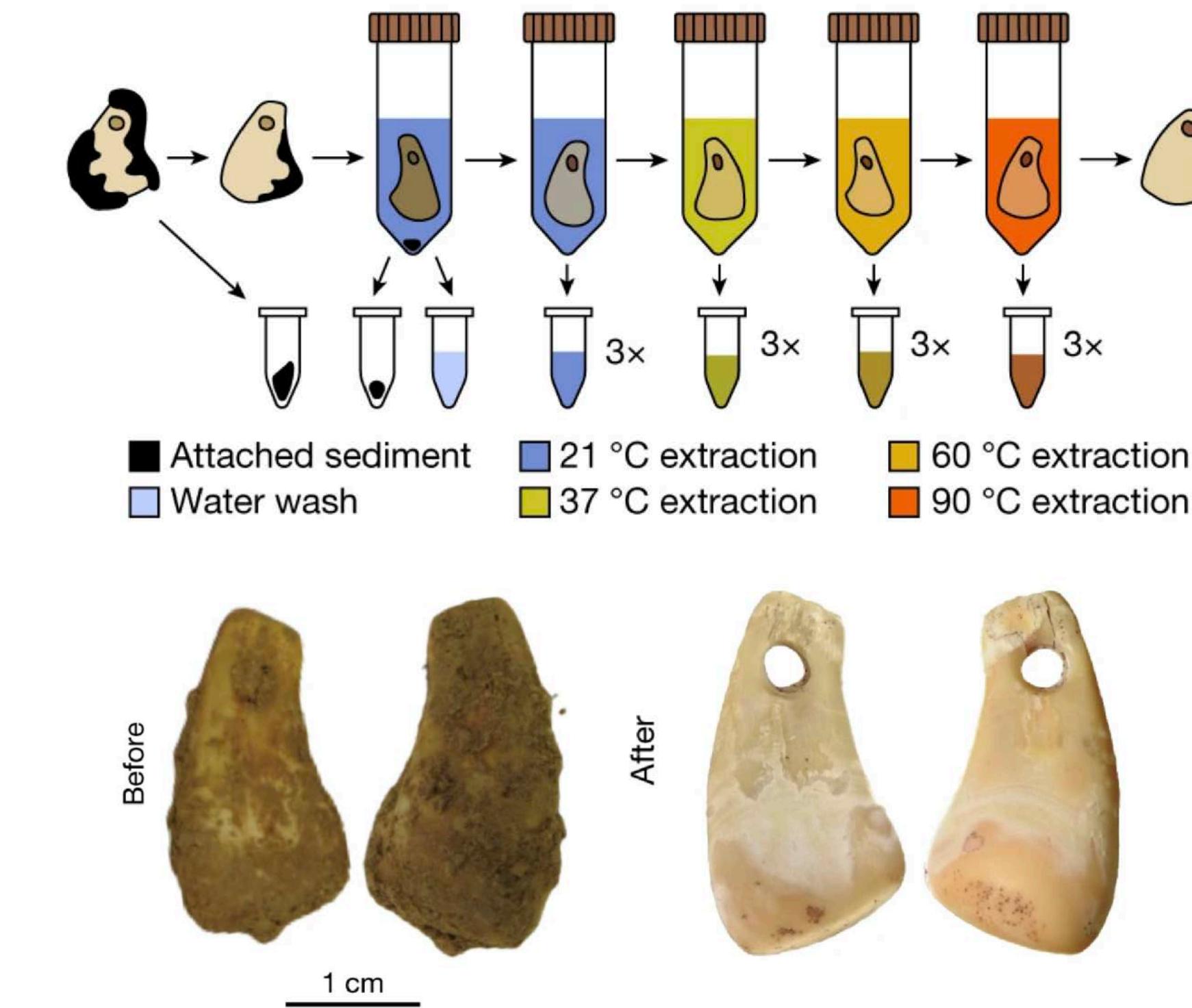
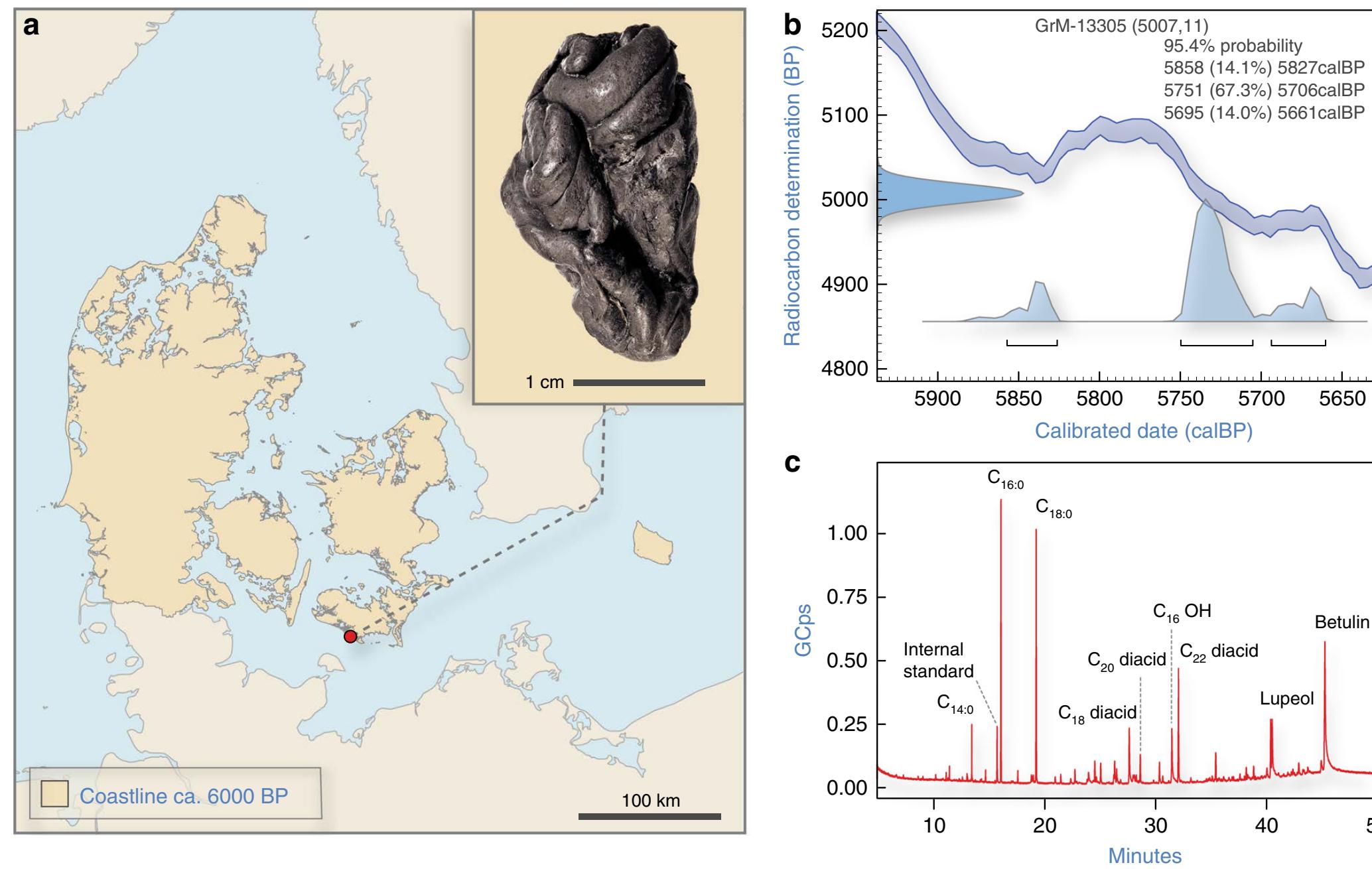
Theis Z.T. Jensen^{1,2,10}, Jonas Niemann^{1,2,10}, Katrine Højholt Iversen^{3,4,10}, Anna K. Fotakis¹, Shyam Gopalakrishnan¹, Åshild J. Vågene¹, Mikkel Winther Pedersen¹, Mikkel-Holger S. Sinding¹, Martin R. Ellegaard¹, Morten E. Allentoft¹, Liam T. Lanigan¹, Alberto J. Taurozzi¹, Sofie Holtsmark Nielsen¹, Michael W. Dee⁵, Martin N. Mortensen^{1,6}, Mads C. Christensen⁶, Søren A. Sørensen⁷, Matthew J. Collins^{1,8}, M. Thomas P. Gilbert^{1,9}, Martin Sikora¹, Simon Rasmussen^{1,4} & Hannes Schroeder^{1*}

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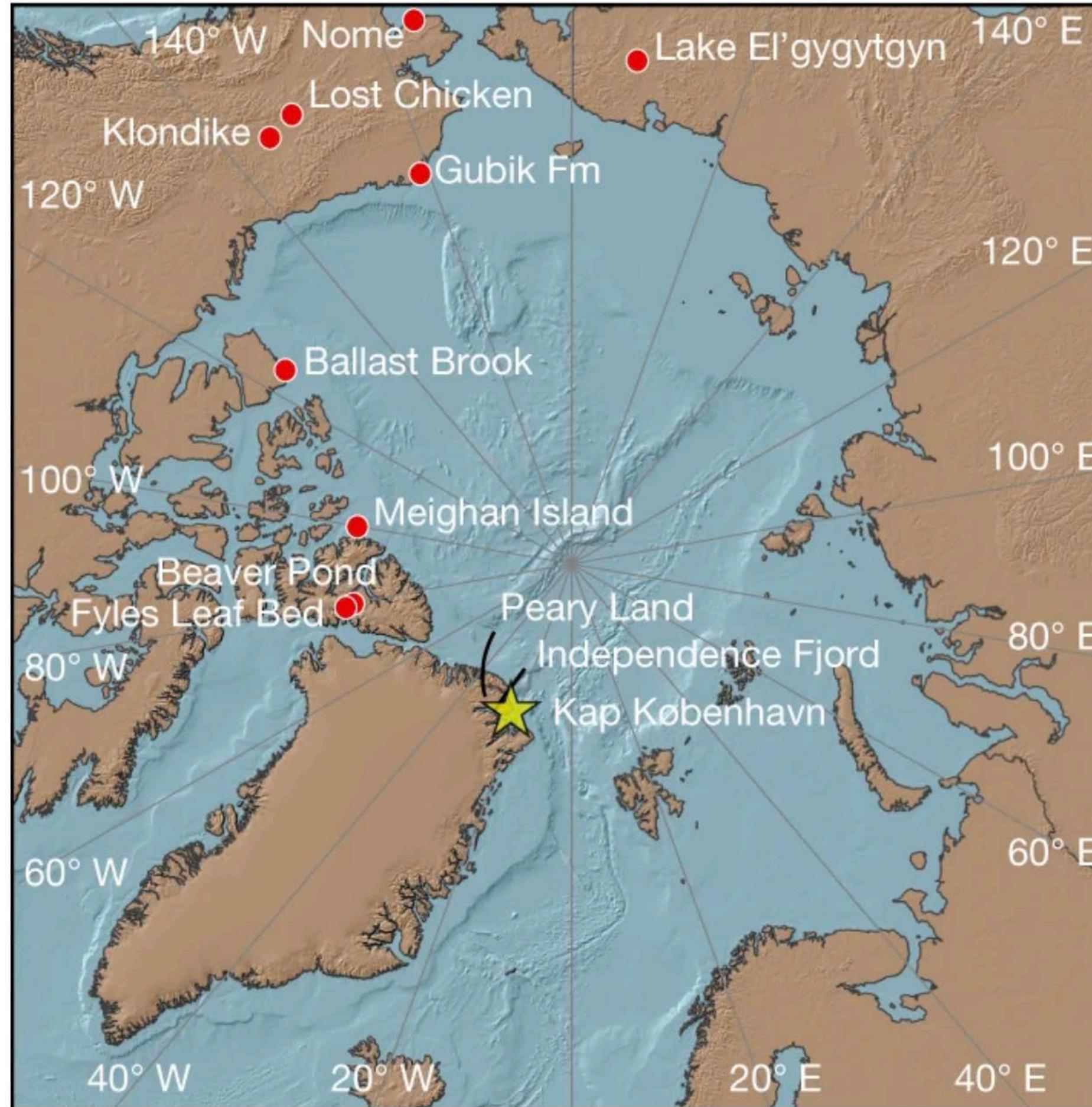
Ancient human DNA recovered from a Palaeolithic pendant

Elena Essel , Elena I. Zavala, Ellen Schulz-Kornas, Maxim B. Kozlikin, Helen Fewlass, Benjamin Vernot, Michael V. Shunkov, Anatoly P. Derevianko, Katerina Douka, Ian Barnes, Marie-Cécile Soulier, Anna Schmidt, Merlin Szymanski, Tsenka Tsanova, Nikolay Sirakov, Elena Endarova, Shannon P. McPherron, Jean-Jacques Hublin, Janet Kelso, Svante Pääbo, Mateja Hajdinjak, Marie Soressi & Matthias Meyer

Nature **618**, 328–332 (2023) | [Cite this article](#)



Ancient environmental DNA - the next revolution



Reconstruction of a 2 million year-old ecosystem from ancient environmental DNA

Ancient DNA

Next Generation Sequencing Analysis

DTU, 9/1/2026

Interested in a project?

martin.sikora@sund.ku.dk