

22126: Next Generation Sequencing Analysis DTU - January 2026 Mick Westbury

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Title

Date

DATA BASICS

Date

Technical University of Denmark



NGS Analysis workflow





Question

Raw

5 -proce 9

mapping or de Assembly **000** calling Variant

ost-processing

Comparison

Answer



Why Raw Data Matters

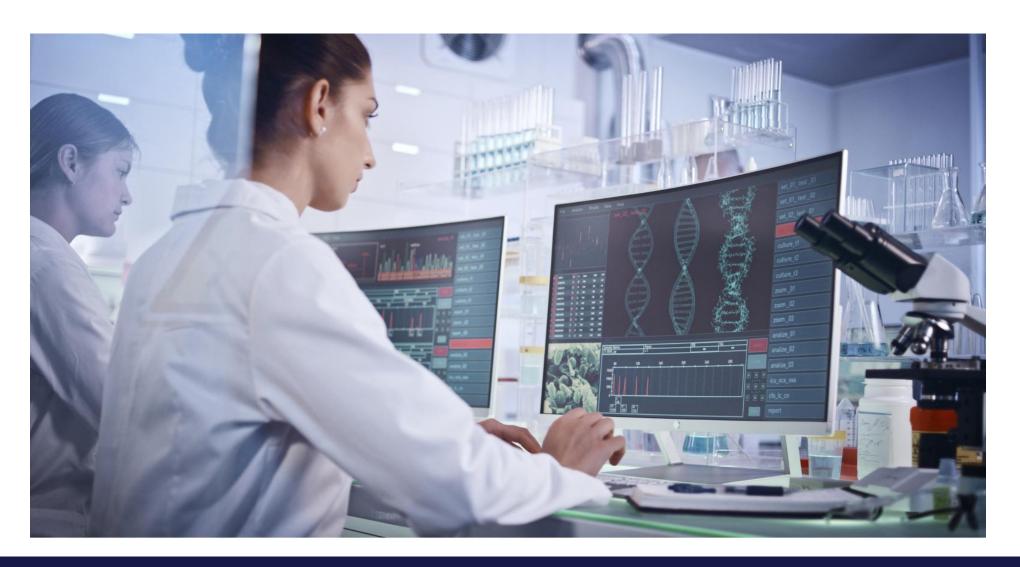
- NGS analysis begins with raw sequencing reads
- Understanding data quality prevents downstream errors
- Poor input leads
 - Poor variant calling
 - Poor assembly
 - Poor quantification







What NGS Data Looks Like





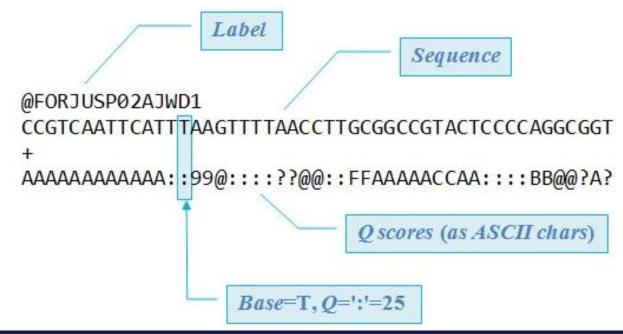
What NGS Data (Actually) Looks Like



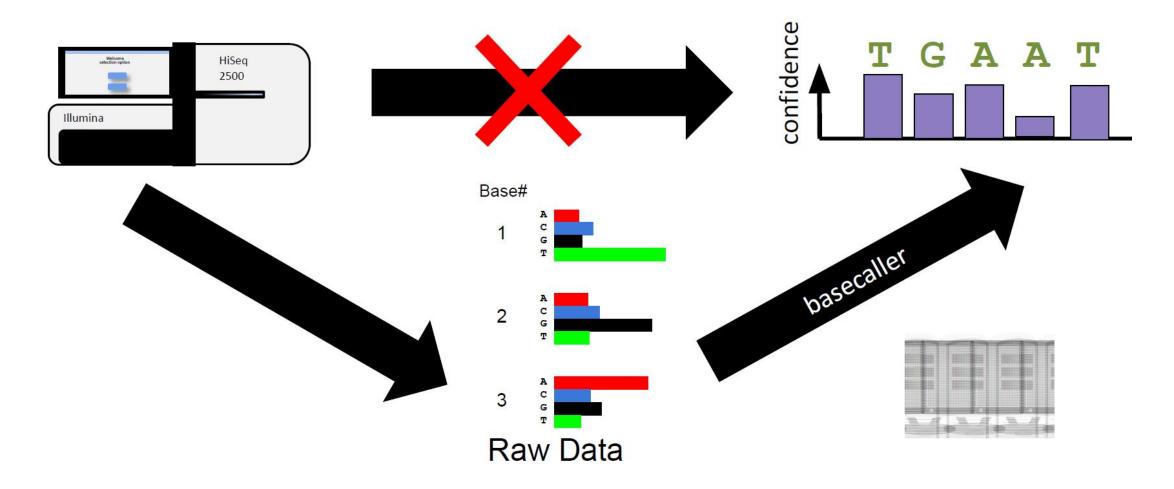


FASTQ Format

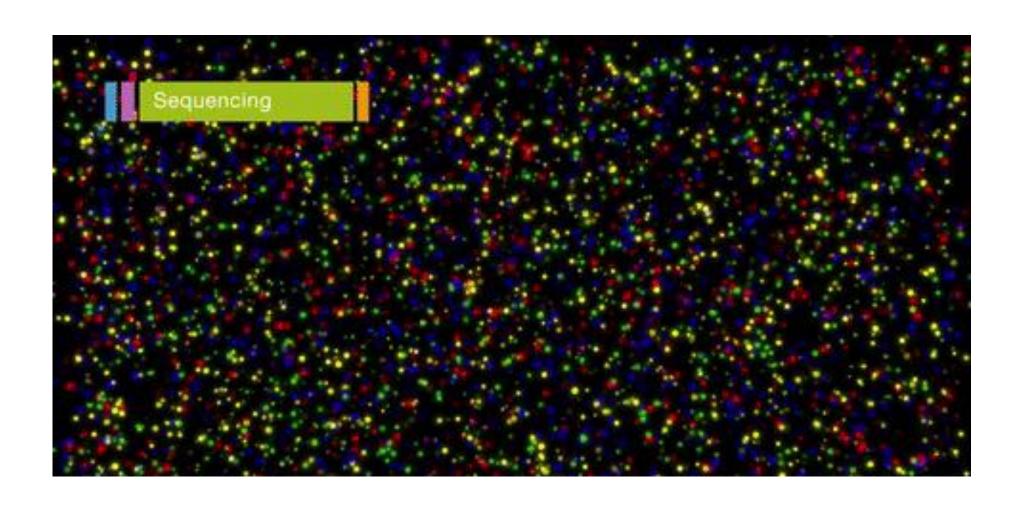
- Header line starts with "@"
- Sequence line (A, C, G, T, N)
- "+" line
- Quality line with ASCII-encoded PHRED values













```
S - Sanger
                                                  Phred+33, raw reads typically (0, 40)
                                                 Solexa+64, raw reads typically (-5, 40)
                                          X - Solexa
I - Illumina 1.3+ Phred+64, raw reads typically (0, 40)
J - Illumina 1.5+ Phred+64, raw reads typically (3, 41)
!"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHIJKLMNOPQRSTUVWXYZ[\]^ `abcdefghijklmnopqrstuvwxyz{|}~
                                            with 0=unused, 1=unused, 2=Read Segment Quality Control Indicator
                                            (Note: See discussion above).
33
           59
                                        126
                                          L - Illumina 1.8+ Phred+33, raw reads typically (0, 41)
0......26...31......40
                                                  Phred+33, Duplex reads typically (0, 50)
           E - ElemBio AVITI Phred+33, raw reads typically (0, 55)
              Phred+33, HiFi reads typically (0, 93)
                                           P - PacBio
```



- Measure probability of a sequencing error
- PHRED = $-10 \times \log 10$ (error probability)
- Higher score means more confidence
- Quality typically decreases toward end of read



Sequencing Errors

- Substitutions (most common in Illumina)
- Insertions or deletions
- Systematic errors:
 - -GC-rich motifs, repeats, homopolymers
- Random errors increase with cycle number



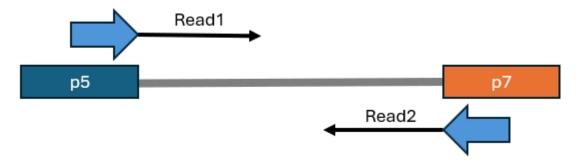
Read Layouts

- Single-end reads (SE)
- Paired-end reads (PE)
- Insert size
- Fragment size

SE sequencing



SE sequencing





Library Preparation Biases

- PCR amplification bias
- GC bias
- Adapter sequences
- Primer dimers
- Overrepresented sequences





Basic QC Concepts

- Per-base sequence quality
- Sequence composition
- GC content distribution
- Read length distribution
- N (missing data) content
- Duplicate sequences



Coverage Concepts

- Depth: average reads per base
- Breadth: proportion of genome covered
- Depends on
 - -read count
 - -read length
 - -genome size
- Uneven coverage affects variant calling