

DTU



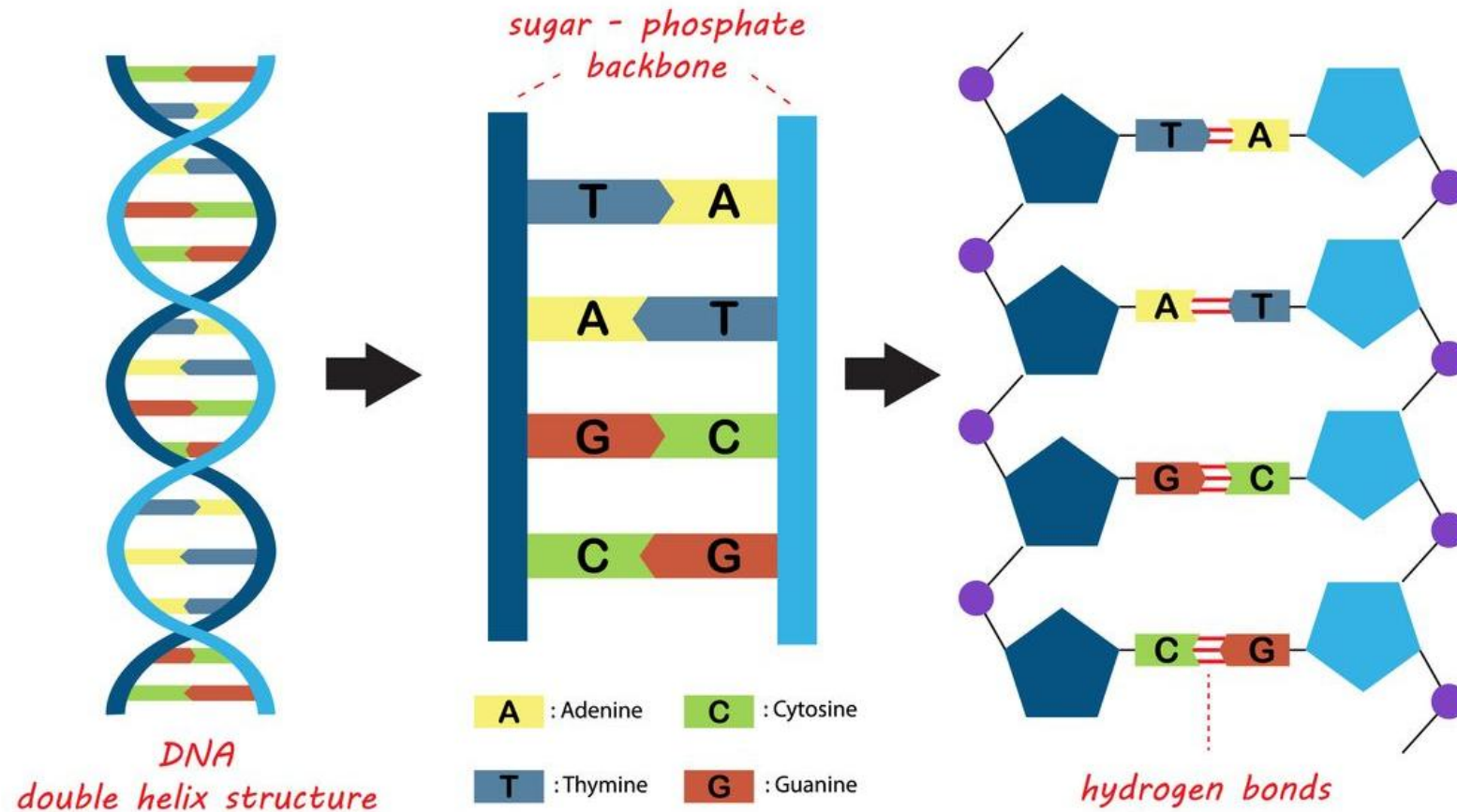
22126: Next Generation Sequencing Analysis

DTU - January 2026

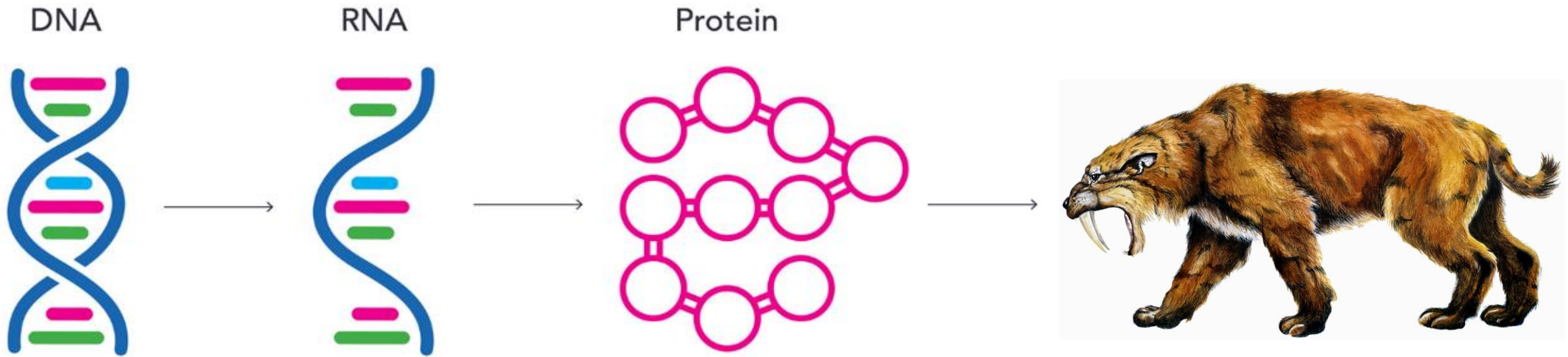
Mick Westbury

*Mick Westbury
Associate Professor
Section of Bioinformatics
Technical University of Denmark
micwe@dtu.dk*

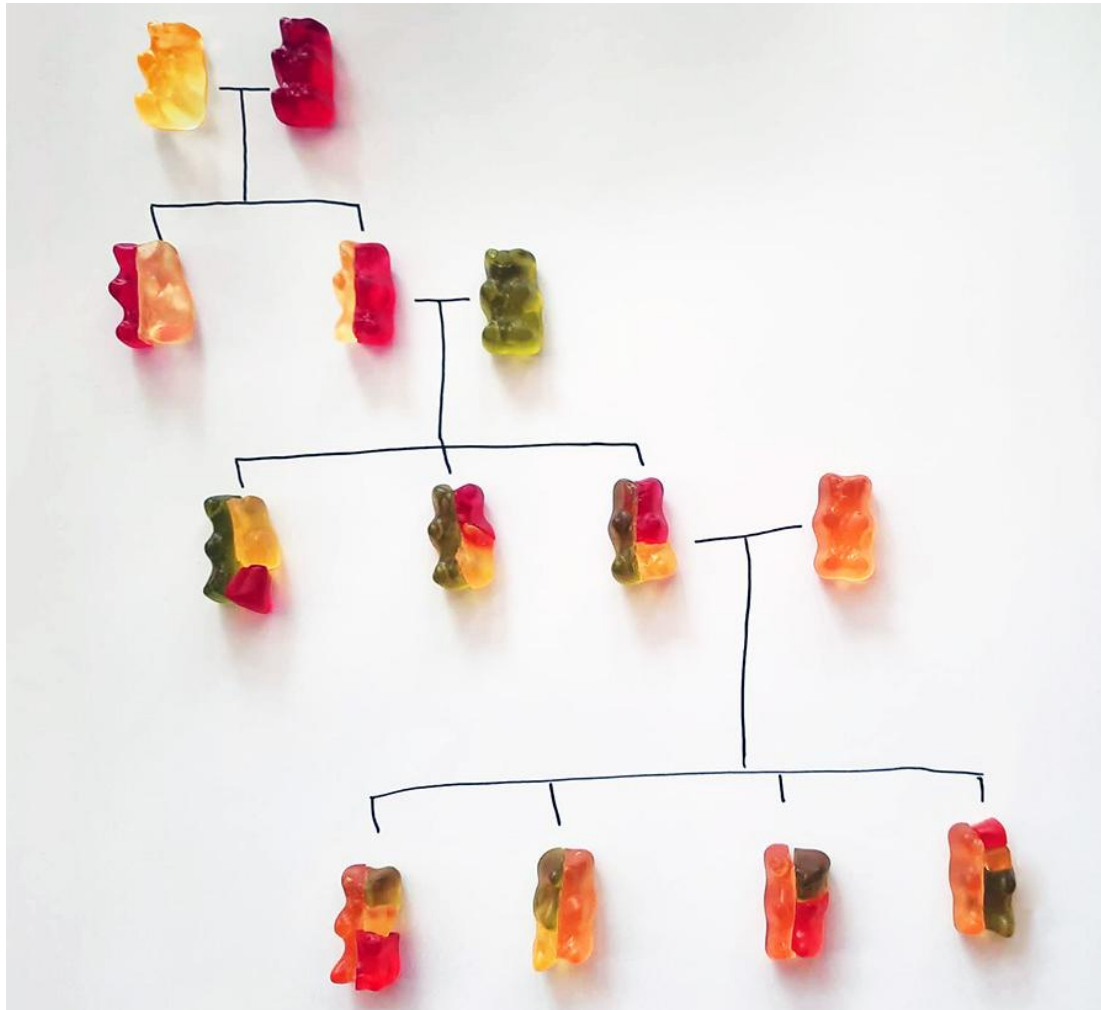
Basic genetics – composition



Basic genetics – function



Basic genetics – inheritance

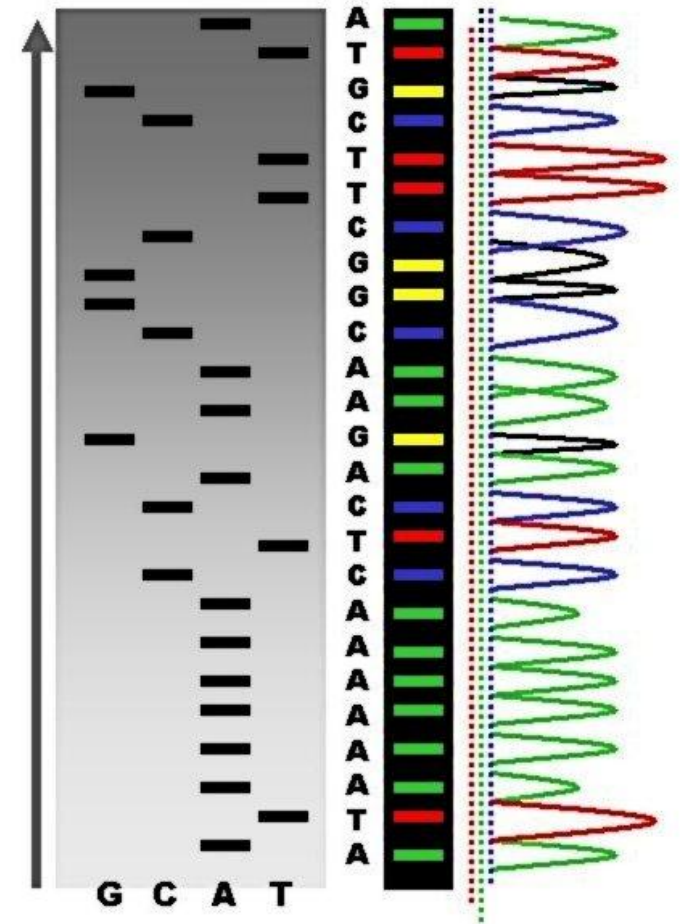


IN THE BEGINNING...



First-generation sequencing

- Sanger sequencing
- Very accurate, long reads (~800–1000 bp).
- Foundation for sequencing logic
- Relatively low throughput (96 reads/run)



First-generation sequencing

- Medical & Human Genetics
 - Diagnose disease-causing mutations
 - Targeted drug development
- Ancient DNA
- Developmental & Molecular Genetics
 - Sequencing of model organism genes
- Population & Evolutionary Genetics
 - Human migration studies
- Agriculture & Targeted Breeding
 - Sequencing of key genes controlling traits

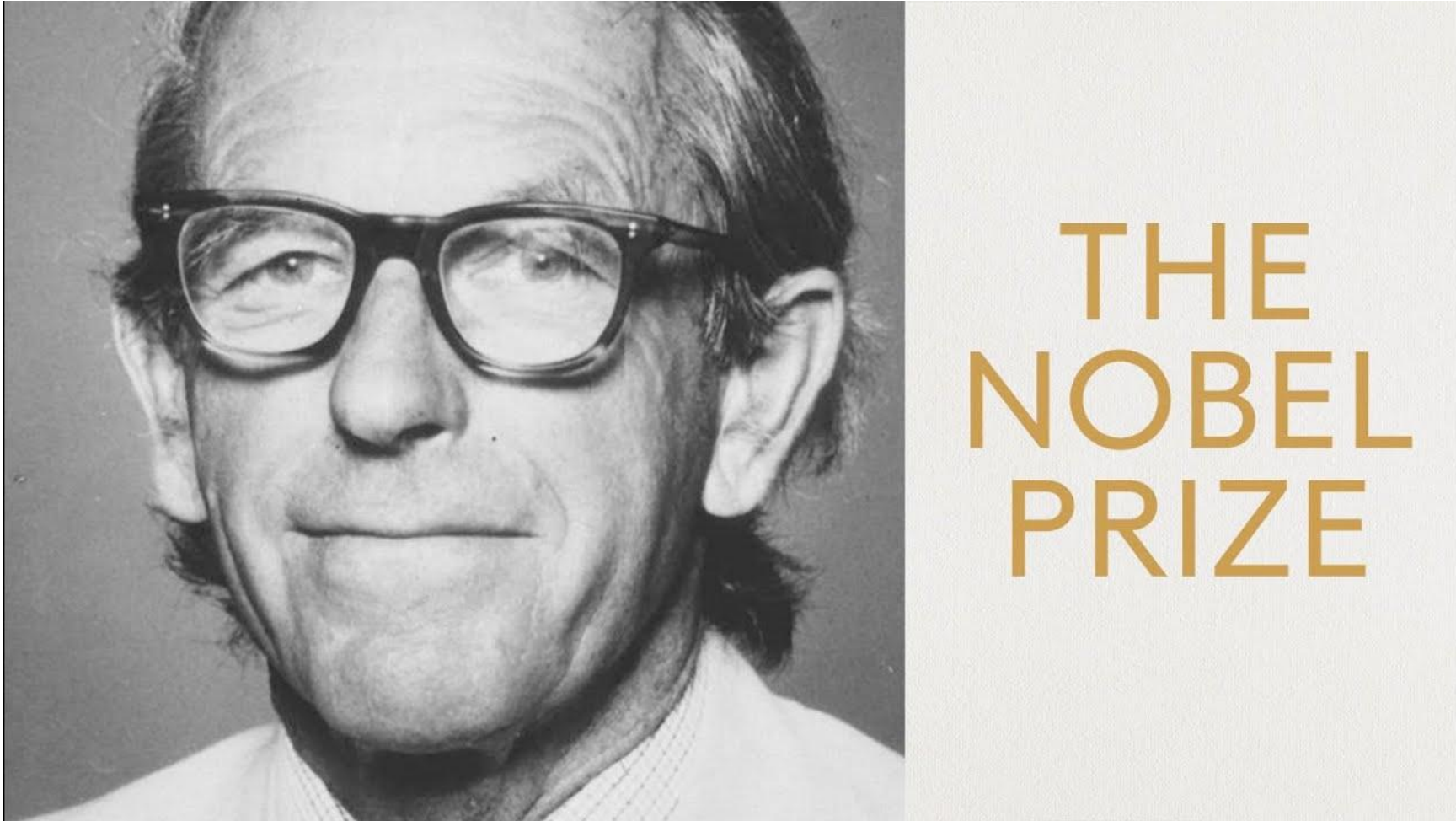


Sanger + HIV biology

- 1985–1987: **First full HIV genomes (~9.7kb) were sequenced**
 - Reveal mutational hotspots and drug targets
 - Allowed the first antiretroviral drugs
- By the mid-1990s, genotypic drug-resistance test became part of HIV care.
 - Detect known mutations linked to treatment evasion
 - Sequencing = find right drug combination.



First-generation sequencing



NGS basics

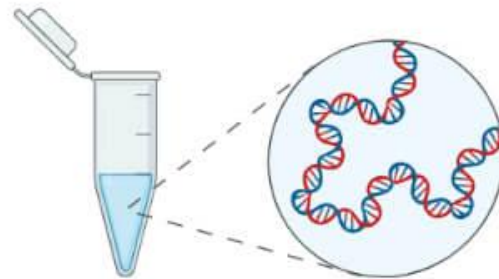
Next-generation sequencing

- Millions to billions of fragments sequenced in parallel
- Fluorescent imaging of each base incorporation
- PCR amplification on a flow cell (clusters)
- Advantages
 - high accuracy
 - high throughput
 - low cost
- Disadvantages
 - short reads (100–300 bp)

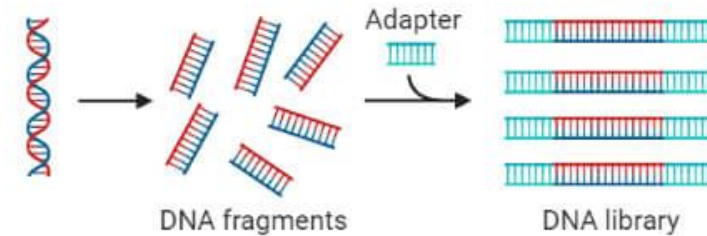


Short-read NGS workflow

Step 1:
DNA extraction

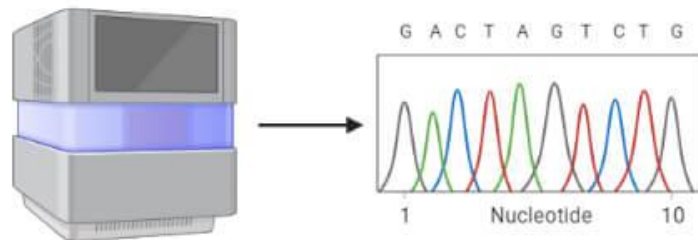


Step 2:
Library preparation

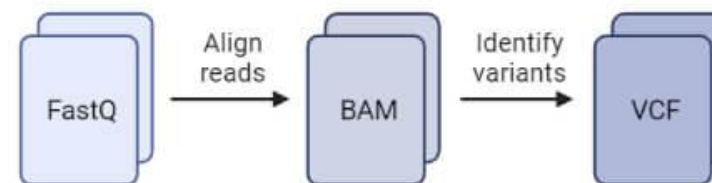


Next Generation Sequencing Workflow

Step 3:
Sequencing



Step 4:
Analysis



NGS Analysis workflow



Question

Raw data

Pre-processing

**Assembly –
mapping or de
novo**

Variant calling

Post-processing

Comparison

Answer

History of NGS sequencers

- 2005 — 454
 - First massively parallel sequencing platform (pyrosequencing).
- 2007 — Illumina (Solexa)
 - Sequencing by synthesis
 - Dominant global platform
- 2010 — Ion Torrent
 - Semiconductor-based sequencing
 - Inexpensive and fast but suffers from homopolymer errors
- 2015 — BGI/DNBseq
 - DNA nanoball–based sequencing-by-synthesis
 - Cheaper, major international competitor to Illumina



History of NGS sequencers

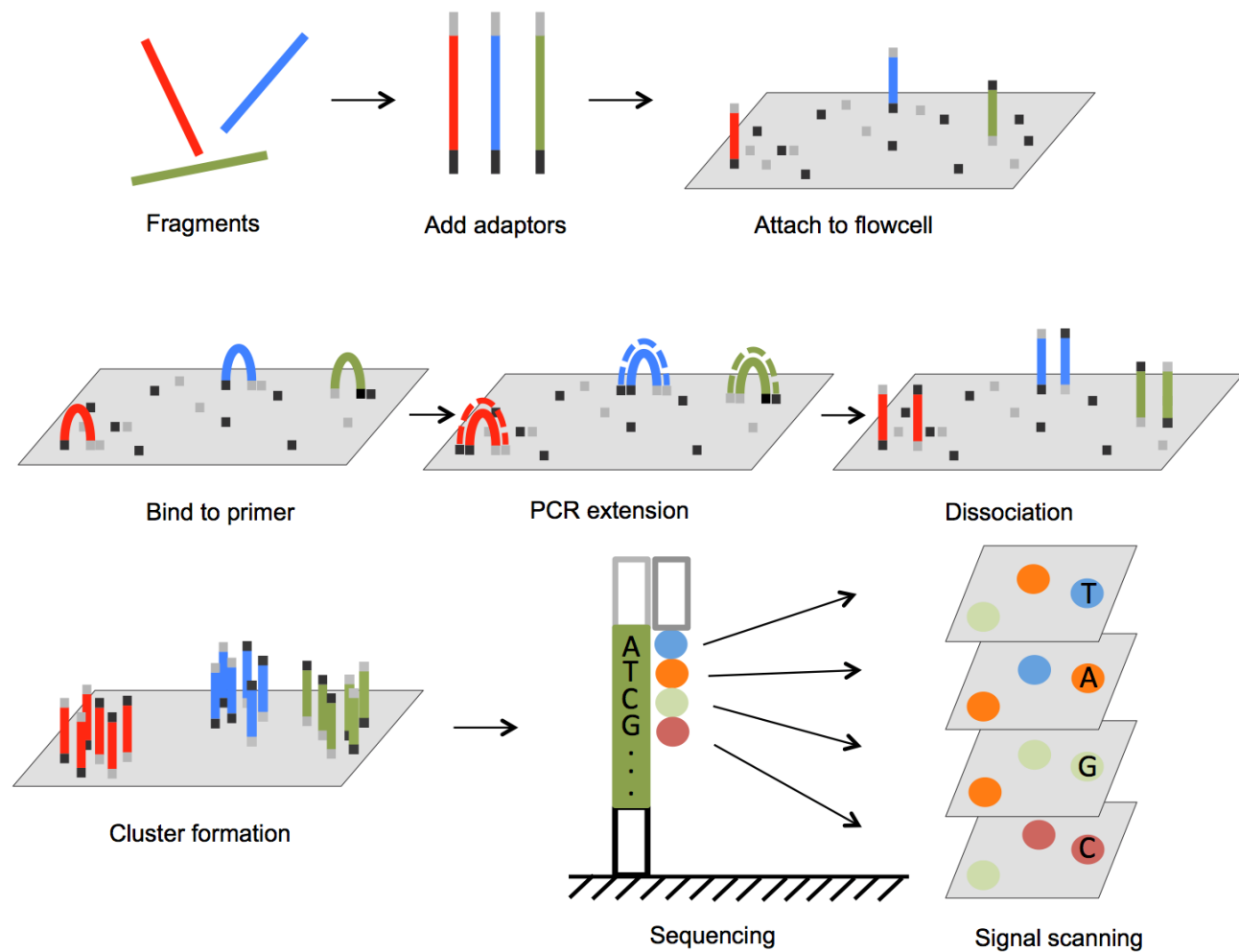
- 2005 — 454
 - First massively parallel sequencing platform (pyrosequencing).
- **2007 — Illumina (Solexa)**
 - **Sequencing by synthesis**
 - **Dominant global platform**
- 2010 — Ion Torrent
 - Semiconductor-based sequencing
 - Inexpensive and fast but suffers from homopolymer errors
- 2015 — BGI/DNBseq
 - DNA nanoball-based sequencing-by-synthesis
 - Cheaper, major international competitor to Illumina



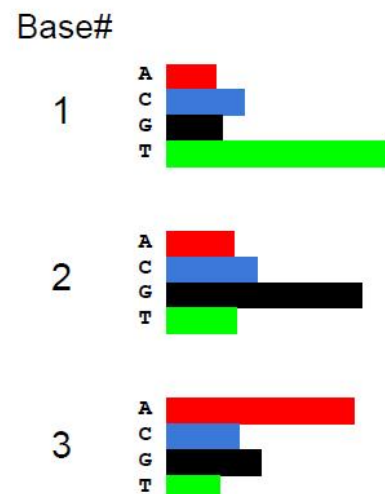
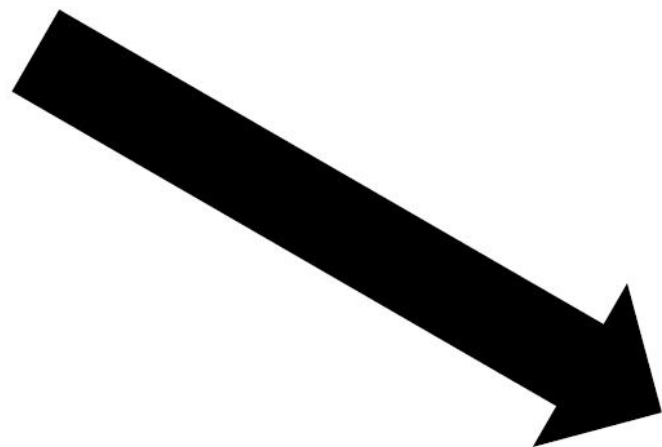
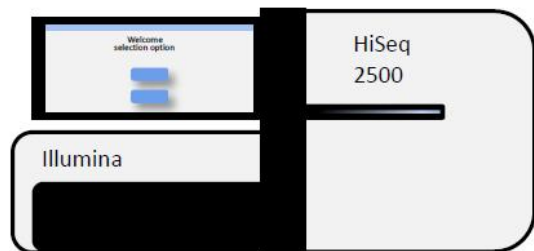
Illumina NGS

- <https://www.youtube.com/watch?v=fCd6B5HRaZ8>

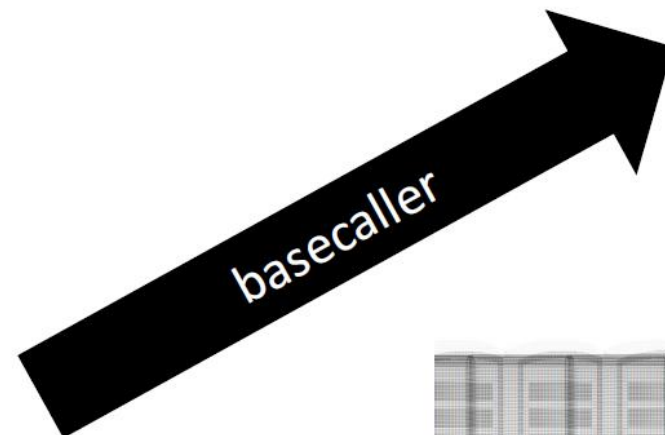
Illumina NGS



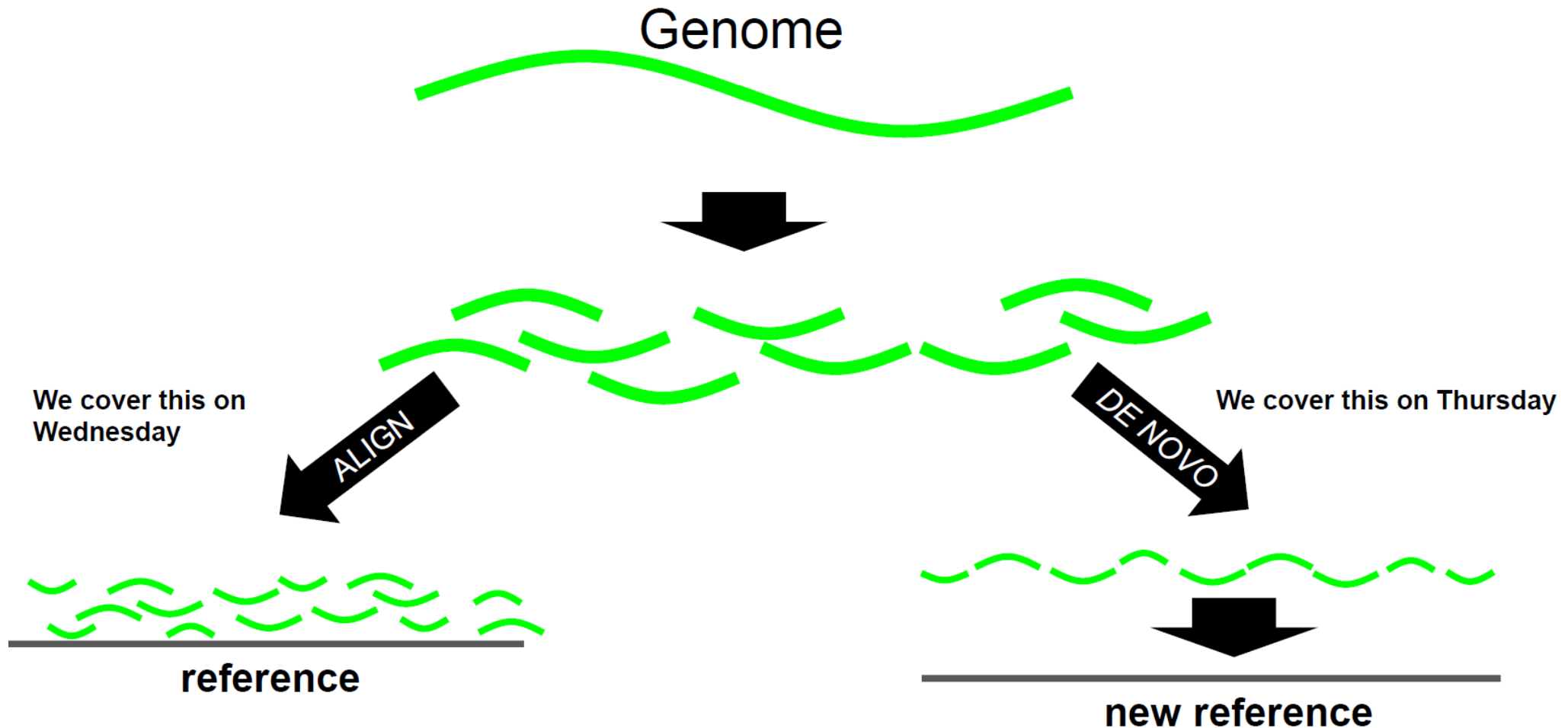
Short-read NGS



Raw Data



Short-read NGS workflow



Key terms

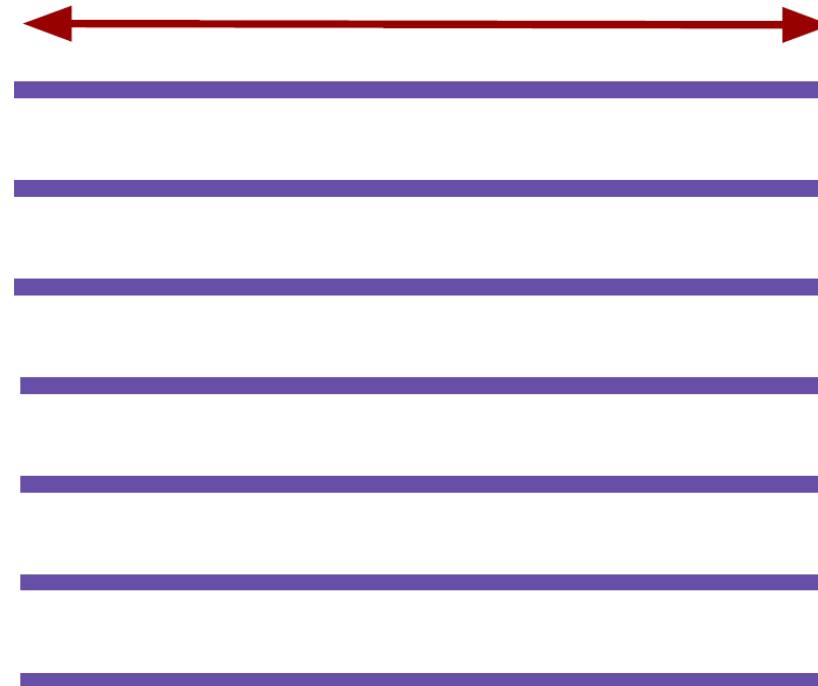
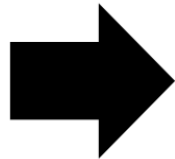
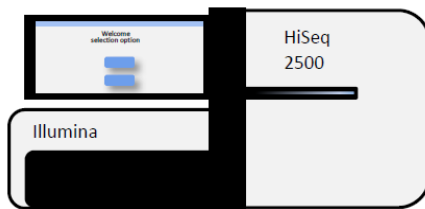
- DNA fragment
- Sequencing read
- Base call
- Errors
- Mapping/alignment
- Assembly

Key concepts

- **Read length**
 - How much of a fragment is read at once
- **Coverage**
 - How many times each base is read
- **Error types**
 - Mismatches, insertions, deletions

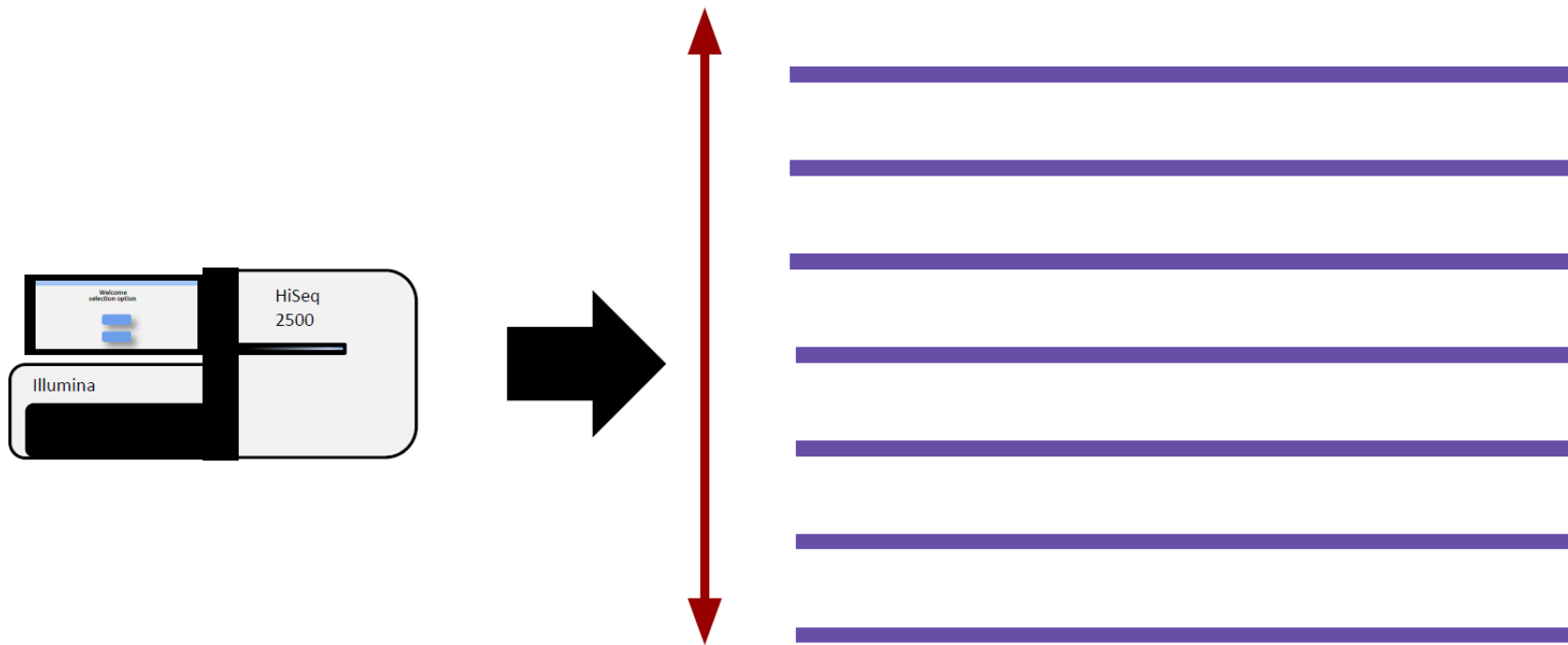
Key concepts

- **Read length**
 - How much of a fragment is read at once



Key concepts

- **Coverage**
 - How many times each base is read



Key concepts

- **Error types**
 - Mismatches (wrong base)

DNA fragment

ACGTACGTACGT**A**CGTACGCTAGCT

ACGTACGTACGT**C**CGTACGCTAGCT

Sequencing read

Key concepts

- **Error types**
 - Insertions (extra base)

DNA fragment

ACGTACGTACGT-CGTACGCTAGCT

ACGTACGTACGT**C**CGTACGCTAGCT

Sequencing read

Key concepts

- **Error types**
 - Deletions (missing base)

DNA fragment

ACGTACGTACGT**A**CGTACGCTAGCT

ACGTACGTACGT-**CGT**ACGCTAGCT

Sequencing read