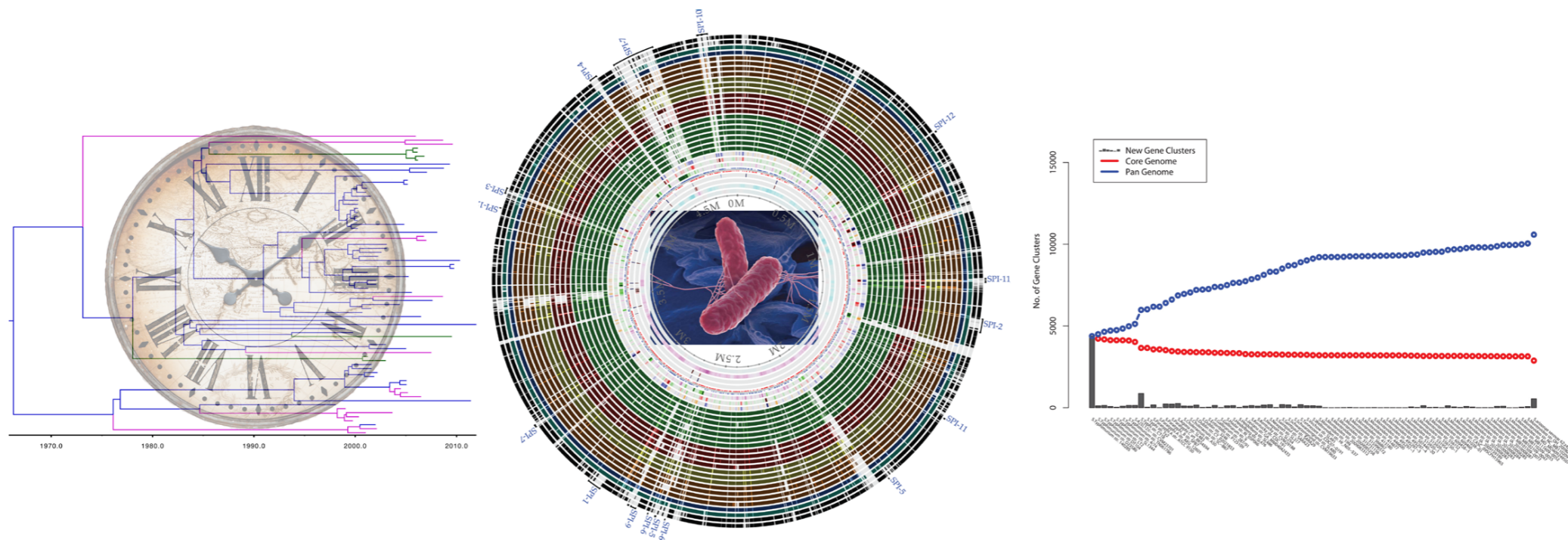
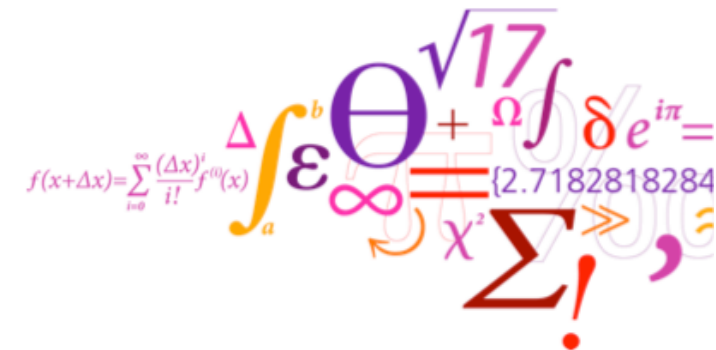


Genomic Epidemiology



Pimlapas Leekitcharoenphon (Shinny)
Senior Researcher
Research Group for Genomic Epidemiologies
National Food Institute (DTU Food)

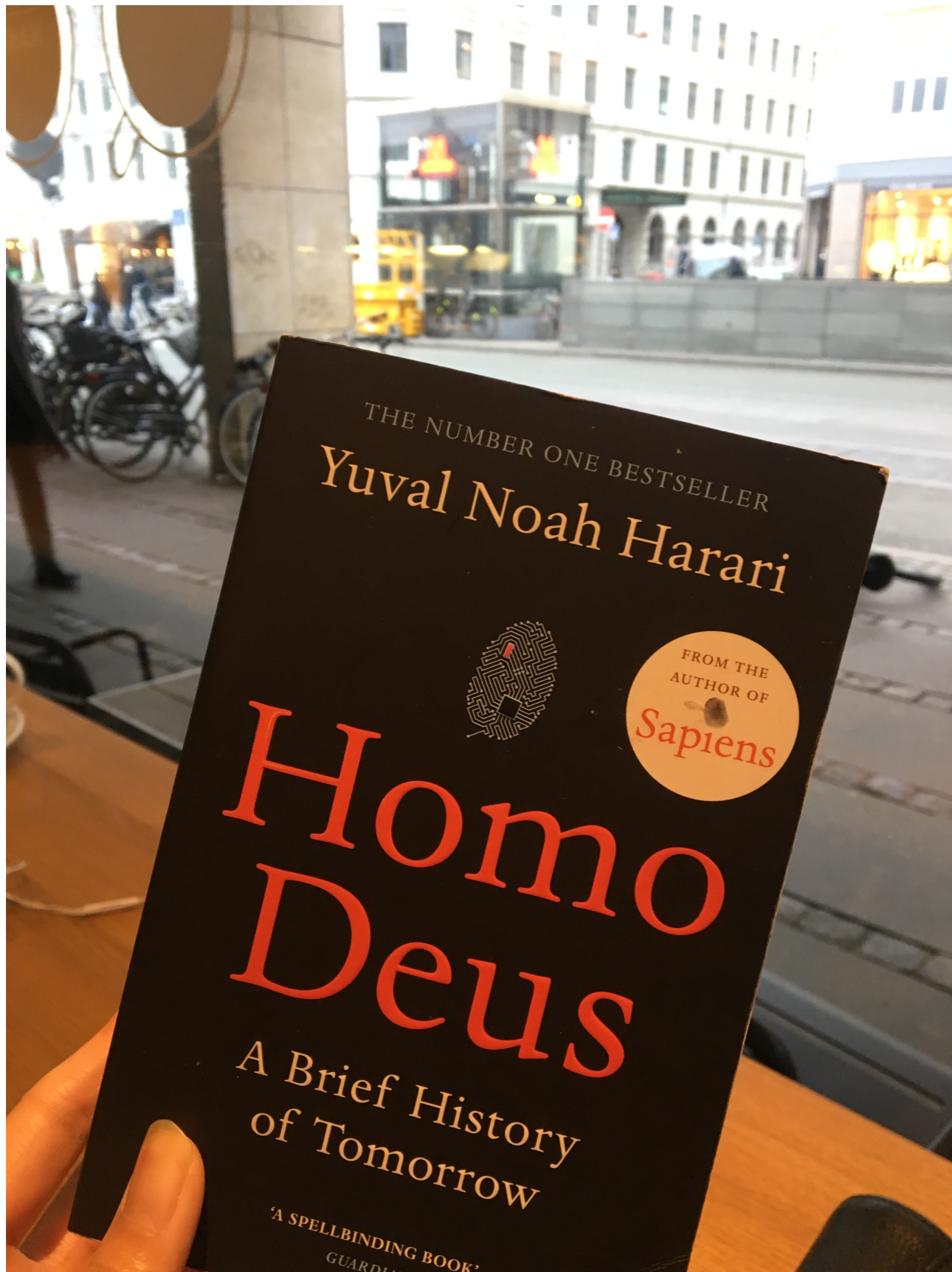
pile@food.dtu.dk



14th January 2025

Topics

- Genomic epidemiology
- Bioinformatics tools for surveillance of infectious diseases



- Famine
- War
- Infectious diseases

Epidemiology

- The science that studies the patterns, causes, and effects of health and disease conditions in defined populations
- Questions;
 - What is it ?
 - Has it been seen before ?
 - How can we fight it ?
 - Is it an outbreak ?

Identification and Typing

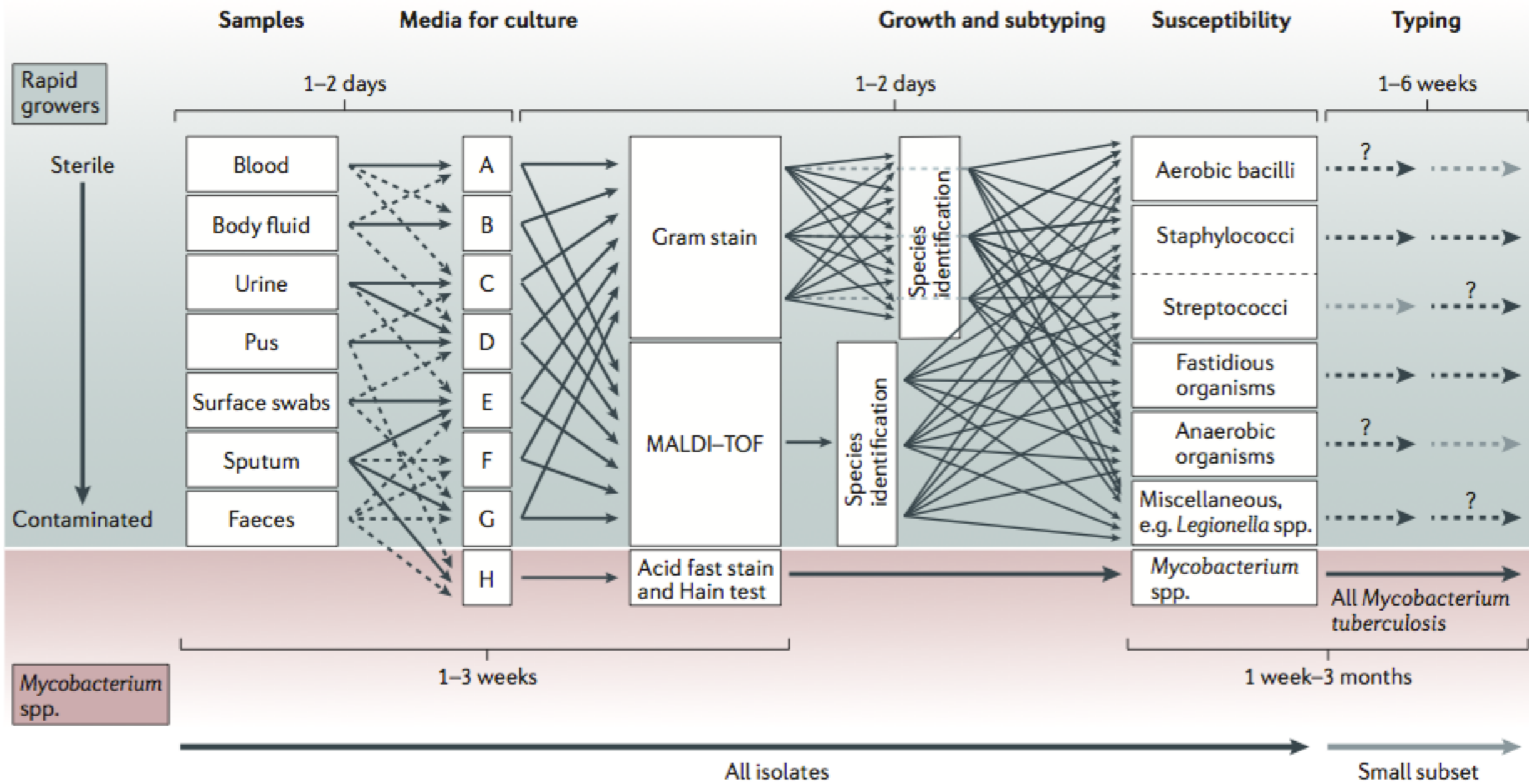
- Any characterization below the (sub-) species level is termed “typing”
- Methods used for this characterization are per definition “typing methods”

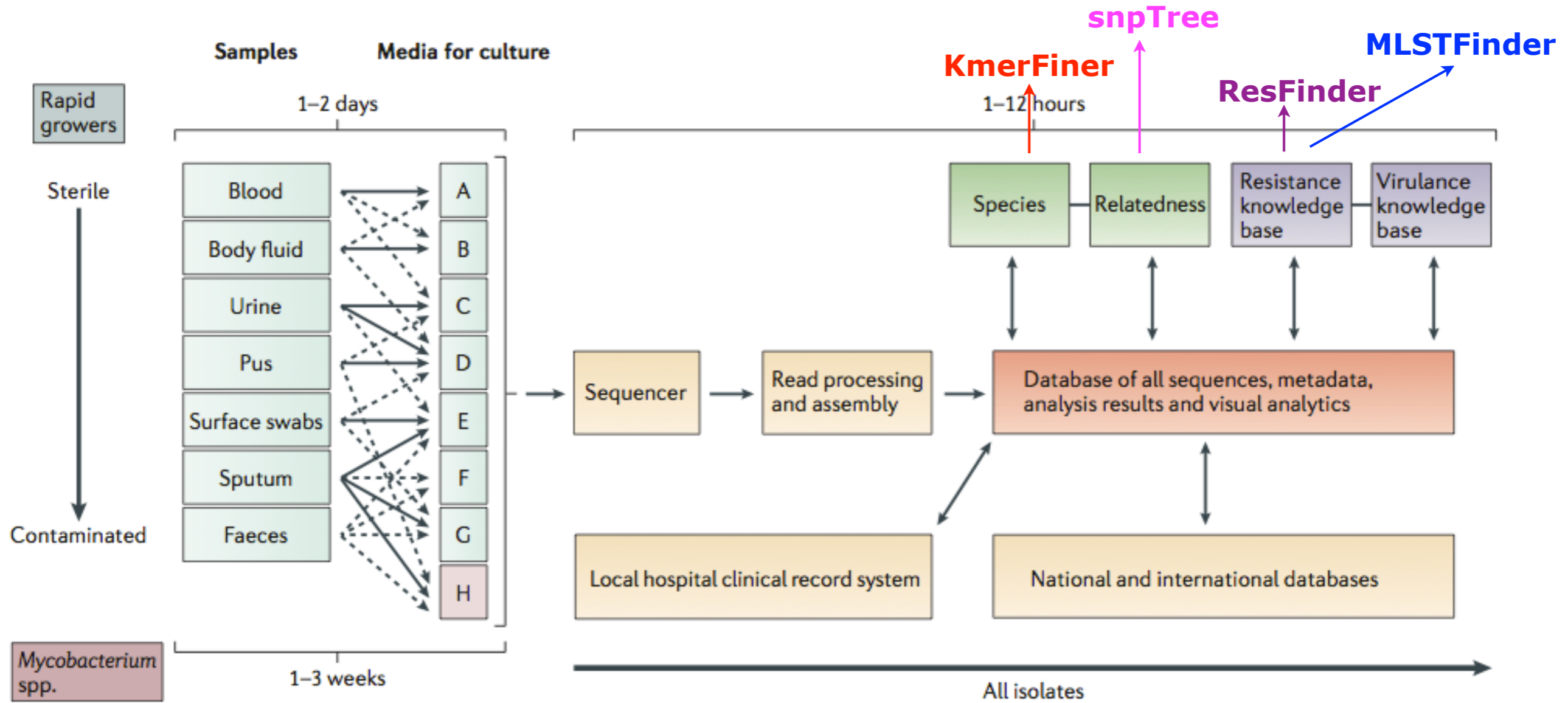
Family
Genus
Species
(Subspecies)

Identification

Serovar
Phagetype
Ribotype
PFGE type
MLVA type
MLST type
DNA Microarray analysis
Whole genomic sequence

Typing





Epidemiology

- The science that studies the patterns, causes, and effects of health and disease conditions in defined populations
- Questions;
 - **What is it ?**
 - Has it been seen before ?
 - How can we fight it ?
 - Is it an outbreak ?

Species Identification

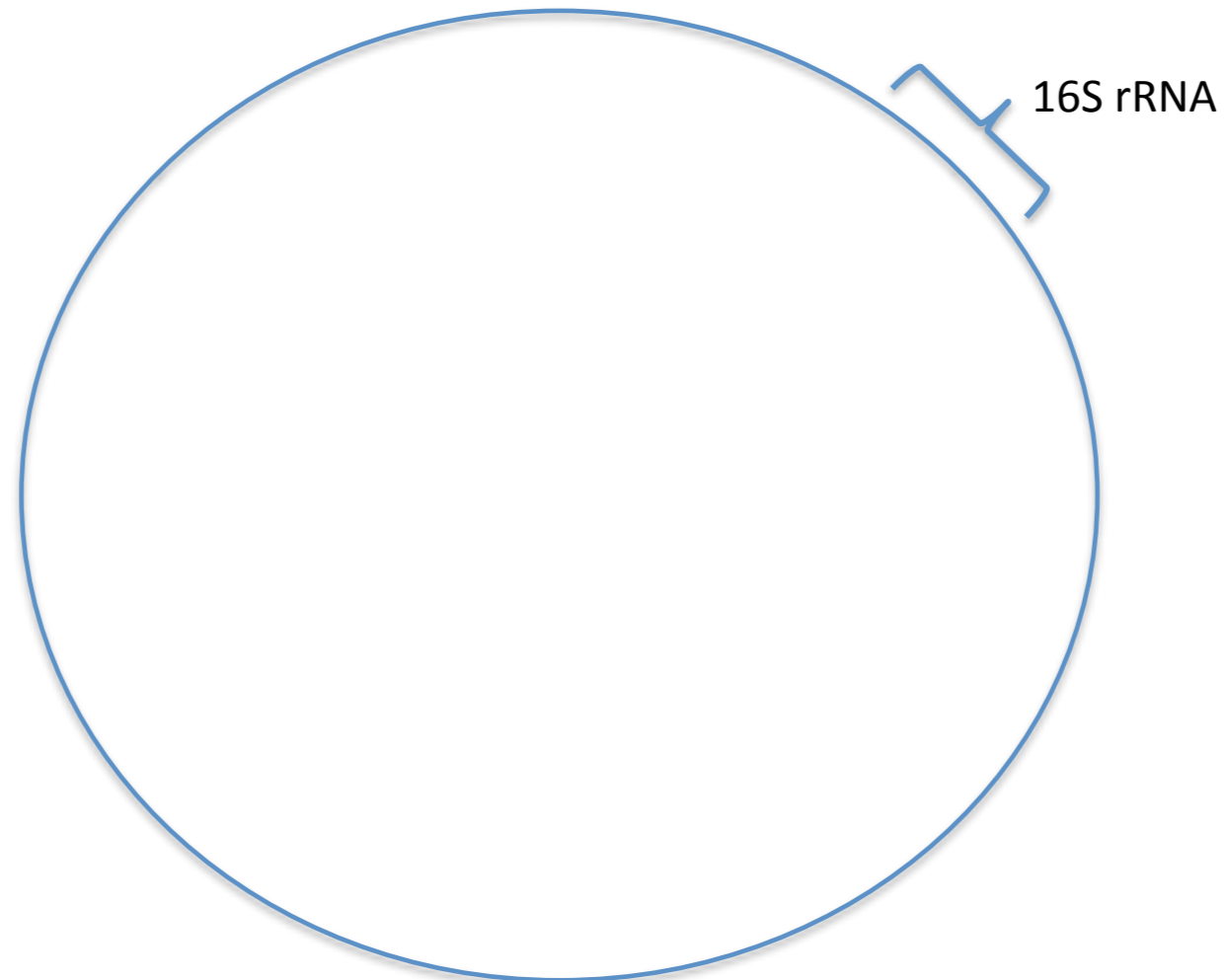
CGE implementation of 16S rRNA species identification - SpeciesFinder

Reference database

- 16S rRNA genes are isolated from genomes in NCBI

Sequence	Isolate in ref. db.	Species
ACGCCG.....CACG	CP32523	<i>K. pneumonia</i>
GATGAG....CGGG	CP64333	<i>E. coli</i>
TGAGGT...TGTTT	CP11212	<i>S. aureus</i>
TGAGGT...TTTTT	CP87878	<i>S. aureus</i>
AAATAG...TGTTT	CP11122	<i>S. enterica</i>
TATAAA....AAAA	CP12121	<i>L. lactis</i>
GATGAG....CGGG	CP86533	<i>E. coli</i>
GTTTAG....CGGG	CP12333	<i>E. coli</i>
GTATTA....AAAA	CP99888	<i>S. pyogenes</i>

The 16s rRNA gene represents only a small fraction of the entire genome



K-mer ?

- A k-mer is a contiguous sequence of k bases
- k is any positive integer
- Sequences with high similarity must share k-mers

sequence

ATGGAAGTCGCGGAATC

7 mers

ATGGAAG
TGGAAAGT
GGAAGTC
GAAGTCG
AAGTCGC
AGTCGCG
GTCGCGG
TCGCGGA
CGCGGAA
GCGGAAT
CGGAATC

Species identification by K-mer

Known species **ATGGAAGTCGCGGAATC**

ATGGAAGTCGCGGAATC Unknown species

k-mers

k-mers

ATGGAAG
TGGAAGT
GGAAGTC
GAAGTCG
AAGTCGC
AGTCGCG
GTCGCGG
TCGCGGA
CGCGGAA
GCGGAAT
CGGAATC

ATGGAAG
TGGAAGT
GGAAGTC
GAAGTCG
AAGTCGC
AGTCGCG
GTCGCGG
TCGCGGA
CGCGGAA
GCGGAAT
CGGAATC



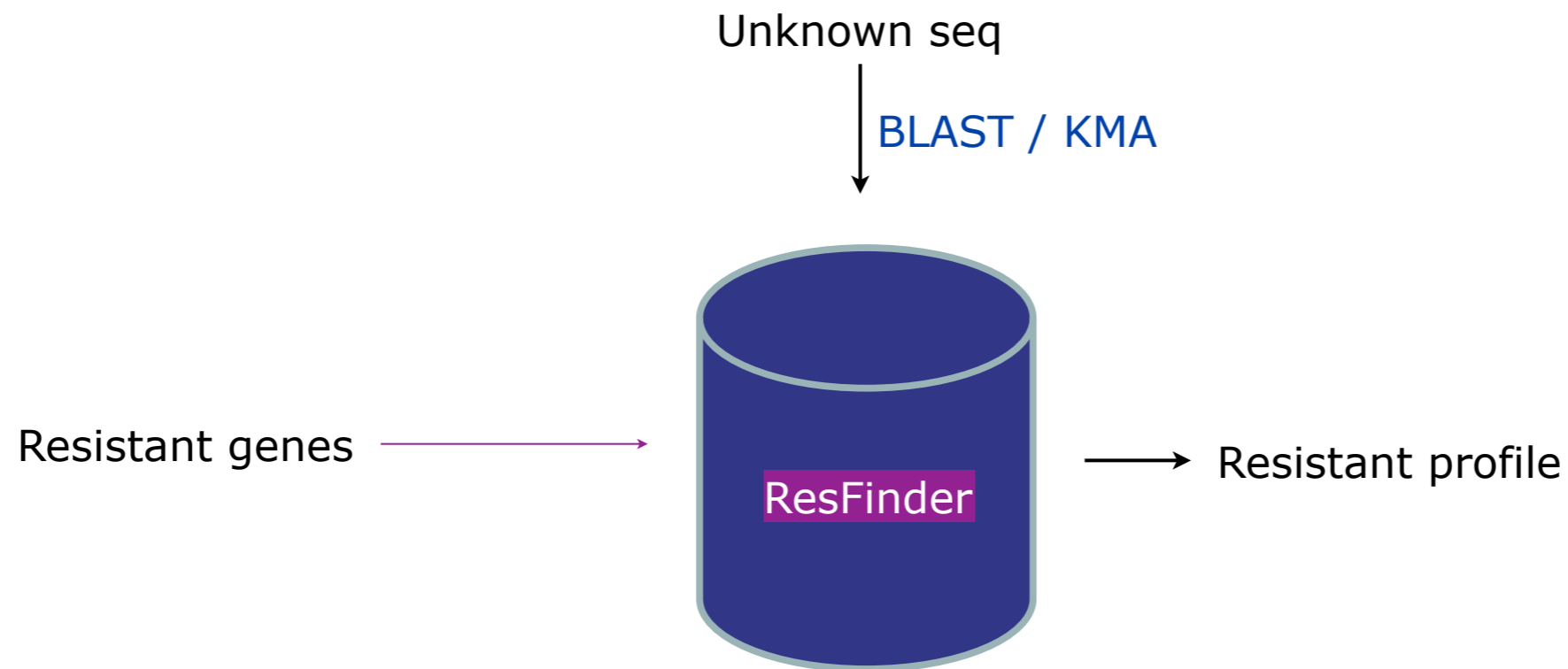
Species

Epidemiology

- The science that studies the patterns, causes, and effects of health and disease conditions in defined populations
- Questions;
 - What is it ?
 - Has it been seen before ?
 - **How can we fight it ?**
 - Is it an outbreak ?

ResFinder

Resistant finding



Epidemiology

- The science that studies the patterns, causes, and effects of health and disease conditions in defined populations
- Questions;
 - What is it ?
 - **Has it been seen before ?**
 - How can we fight it ?
 - **Is it an outbreak ?**

What is phylogeny used for

- Classify taxonomy – The classic use
- Outbreak detection – Increasing with WGS data

What is phylogeny used for

- Cholera outbreak in Haiti 2010
- Listeria outbreak 2014

Whole-genome Sequencing Used to Investigate a Nationwide Outbreak of Listeriosis Caused by Ready-to-eat Delicatessen Meat, Denmark, 2014.

Kvistholm Jensen et al. Clin Infect Dis. (2016) 63 (1): 64-70. doi: 10.1093/cid/ciw192

Case story

- *Vibrio Cholerae* outbreak in Haiti followed the 2010 earthquake
- Rumors said that the outbreak may have come from Nepal, travelling along with UN soldiers from Nepal
- No proof had been given of this until the Hendriksen *et al.* paper in 2011

Population Genetics of *Vibrio cholerae* from Nepal in 2010: Evidence on the Origin of the Haitian Outbreak. Hendriksen et al. 23 August 2011 mBio vol. 2 no. 4 e00157-11. doi: 10.1128/mBio.00157-11

Case story

- Data
 - 24 recent *V. cholerae* strains from Nepal
 - 10 previously sequenced *V. cholerae* isolates, including 3 from the Haitian outbreak
- Analysis
 - Antimicrobial susceptibility testing
 - PFGE (pulsed-field gel electrophoresis) to analyze for genetic relatedness
 - Whole genome sequencing, SNP identification and phylogenetic analysis

Case story - Results

Resistance profile	Susceptible	Decreased susceptibility	Resistant
Nepalese strains <i>Hendriksen et al. 2011</i>	Tetracycline	Ciprofloxacin	Trimethoprim, Sulfamethoxazole Nalidixic
Haitian outbreak strains Centers for Disease Control and Prevention, 2010	Tetracycline	Ciprofloxacin	Trimethoprim, Sulfamethoxazole Nalidixic

SNPs detection

...ATCGAATTCCGGGTTTTTAACCGGATCGTACGATCGGGAAAAA..

TTCCAGG

TTCCAGG

TTCCAGG

TTCCAGG

TTCCAGG

TTCCAGG

SNPs detection



Variant calling format (VCF)

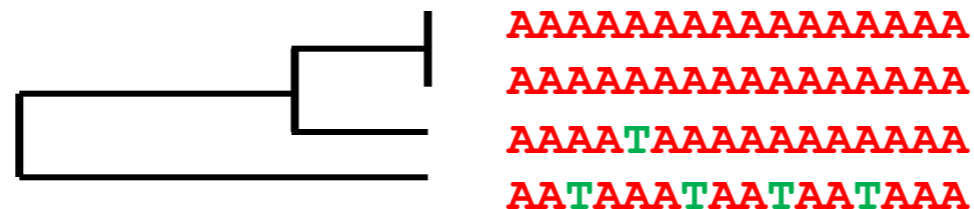
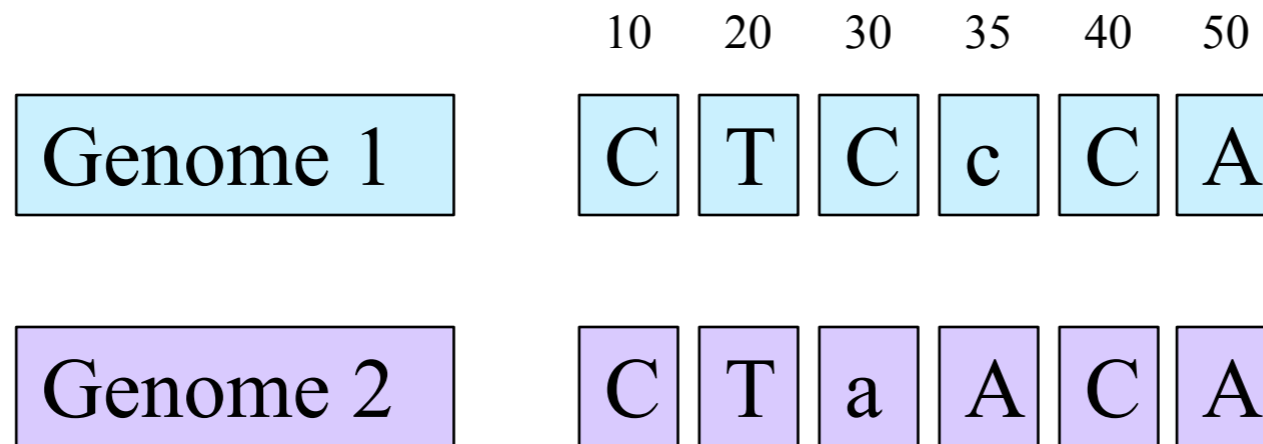
Genome 1	position	ref	change
Ref_genome	10	T	C
Ref_genome	20	C	T
Ref_genome	30	A	C
Ref_genome	40	A	C
Ref_genome	50	G	A

Genome 2	position	ref	change
Ref_genome	10	T	C
Ref_genome	20	C	T
Ref_genome	35	C	A
Ref_genome	40	A	C
Ref_genome	50	G	A

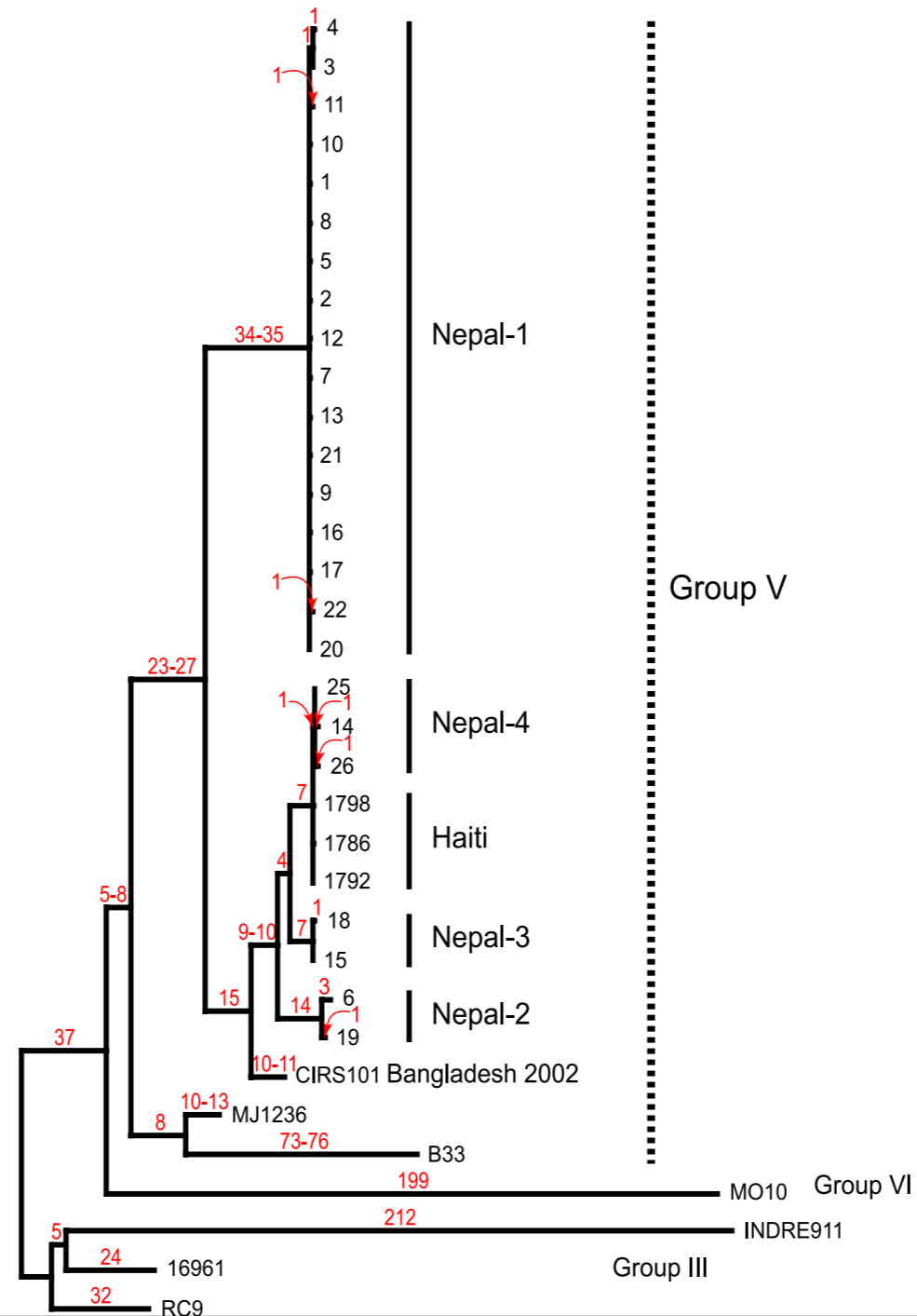
Concatenated SNPs

Genome 1	position	ref	change
Ref_genome	10	T	C
Ref_genome	20	C	T
Ref_genome	30	A	C
Ref_genome	40	A	C
Ref_genome	50	G	A

Genome 2	position	ref	change
Ref_genome	10	T	C
Ref_genome	20	C	T
Ref_genome	35	C	A
Ref_genome	40	A	C
Ref_genome	50	G	A



Case story - Results



Nextstrain

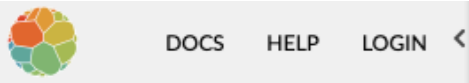
Real-time tracking of pathogen evolution

Nextstrain is an open-source project to harness the scientific and public health potential of pathogen genome data. We provide a continually-updated view of publicly available data alongside powerful analytic and visualization tools for use by the community. Our goal is to aid epidemiological understanding and improve outbreak response. If you have any questions, or simply want to say hi, please give us a shout at hello@nextstrain.org.

READ MORE

SARS-CoV-2 (COVID-19)

We are incorporating SARS-CoV-2 genomes as soon as they are shared and providing analyses and situation reports. In addition we have developed a number of resources and tools, and are facilitating independent groups to run their own analyses. Please see the [SARS-CoV-2 resources page](#) for more information.



DOCS HELP LOGIN

Genomic epidemiology of novel coronavirus - Global subsampling

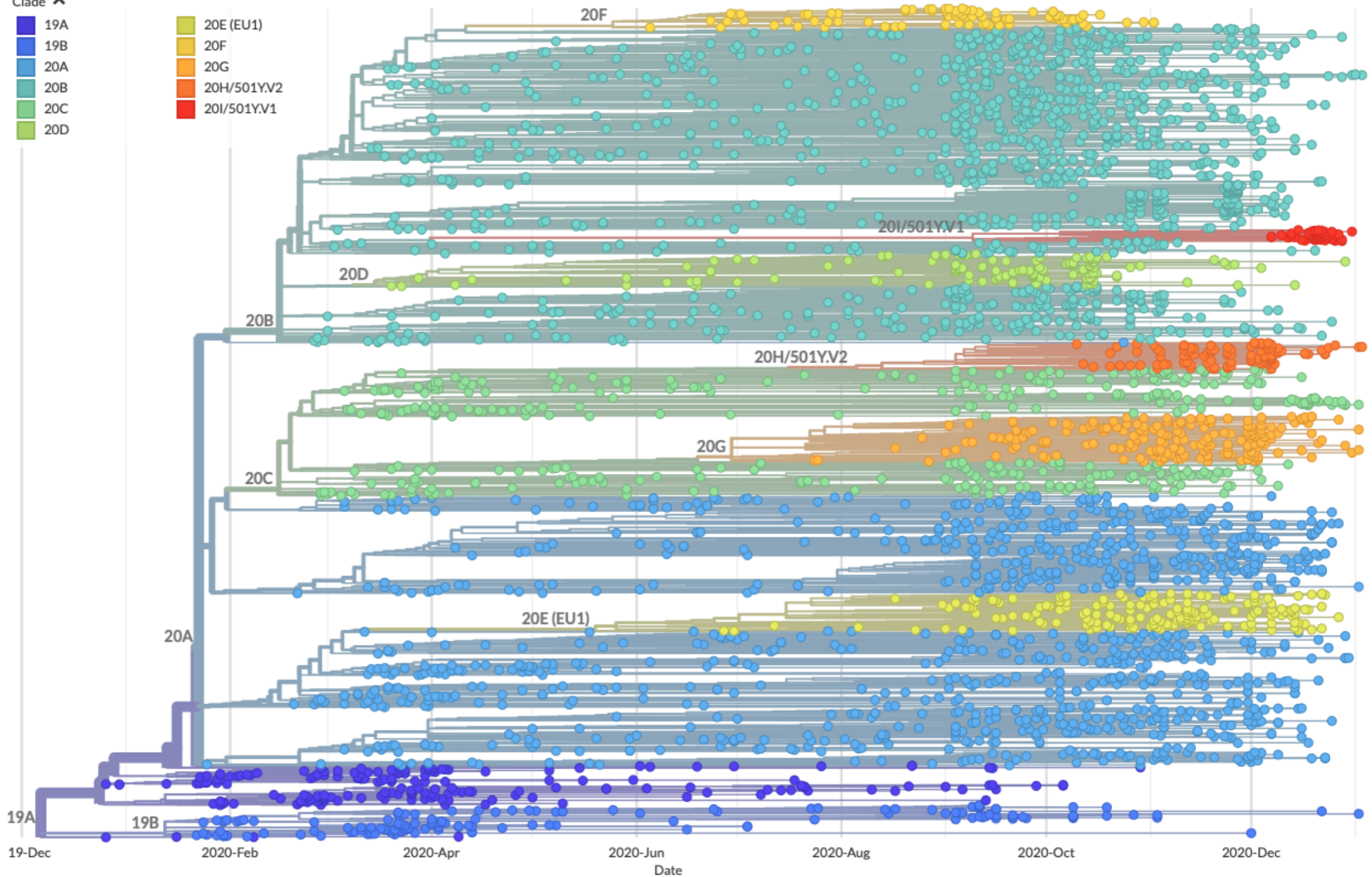
Maintained by the [Nextstrain](#) team. Enabled by data from [GISAID](#)

Showing 3917 of 3917 genomes sampled between Dec 2019 and Jan 2021.

Phylogeny

- Clade ^
- 19A
 - 19B
 - 20A
 - 20B
 - 20C
 - 20D

- 20E (EU1)
- 20F
- 20G
- 20H/501Y.V2
- 20I/501Y.V1



ZOOM TO SELECTED RESET LAYOUT

Dataset

ncov
global

Date Range

2019-12-03 2021-01-07

Color By

Clade

Filter Data

Type filter query here...

Tree Options

- Layout
- RECTANGULAR
 - RADIAL
 - UNROOTED
 - CLOCK

Branch Length

TIME DIVERGENCE

Show confidence intervals

Branch Labels

clade

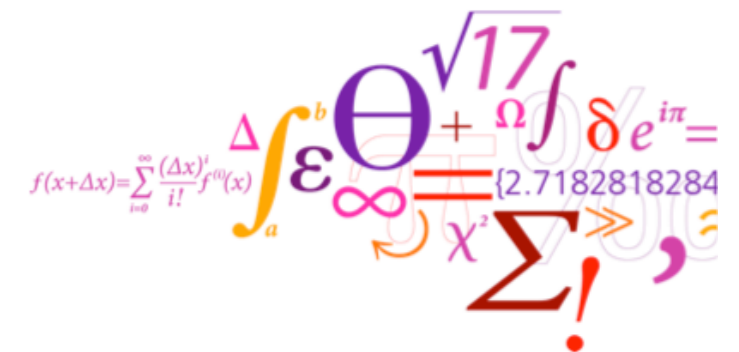
Tip Labels

Sample Name

Second Tree

Select...

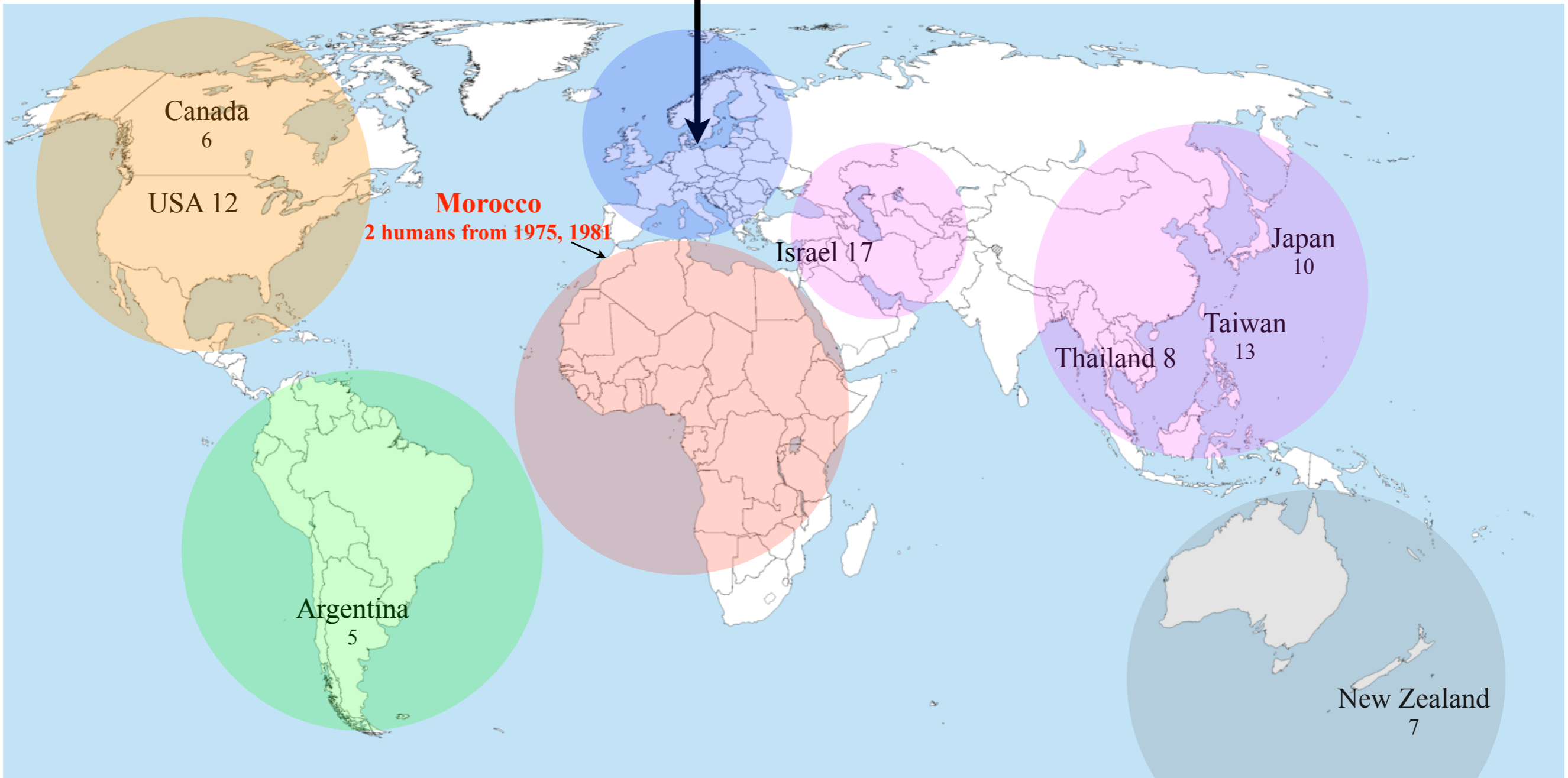
Genomic epidemiology of the global occurrence *S. Typhimurium* DT104



S. Typhimurium DT104

- During the last three decades, *S. Typhimurium* phage type DT104 emerged as the most important phage type and one of the best-studied because of its rapid global dissemination [Lan R, *et al.* Infect Genet Evol. 2009] [Helms M, *et al.* Emerg Infect Dis. 2005]
- DT104 has a multiple antimicrobial resistance pattern to ampicillin, chloramphenicol, streptomycin, sulphonamide and tetracycline (ACSSuT) [Mulvey MR, *et al.* Microbes Infect. 2006]
- Previous epidemics with MDR phage types of *S. Typhimurium*, such as DTs 29, 204, 193 and 204c, were mostly restricted to cattle [Threlfall EJ. J Antimicrob Chemother. 2000]
- DT104 spread among all domestic animals including cattle, poultry, pigs and sheep [Threlfall EJ. J Antimicrob Chemother. 2000]

Ireland 10	Scotland 14	Denmark 79
Spain	Austria 30	Netherlands 22
1 human from 1976	Germany 27	
France 9	Luxembourg 13	Switzerland 8
1 human from 1969	Poland 13	Czech Republic 9

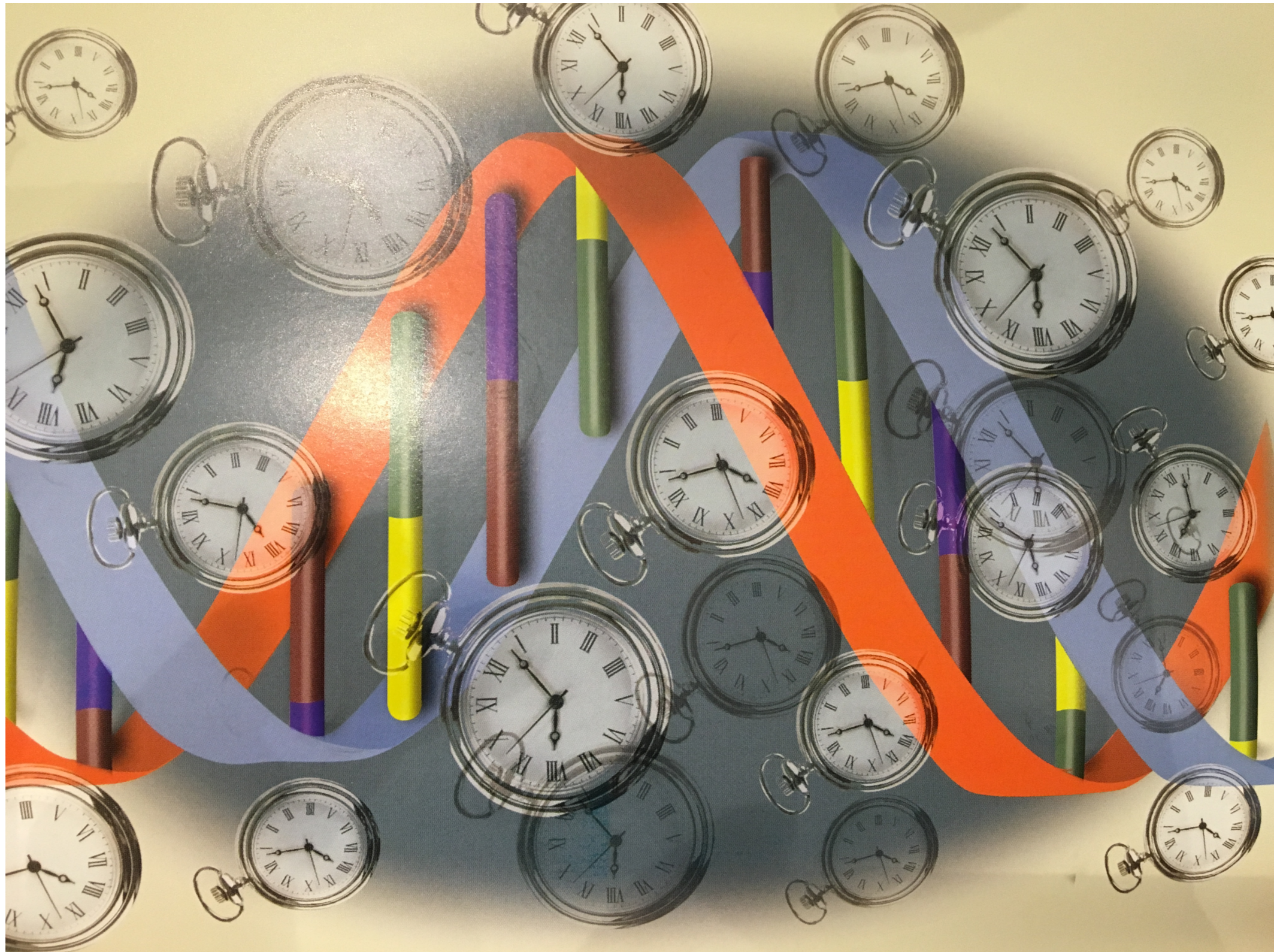


315 genomes

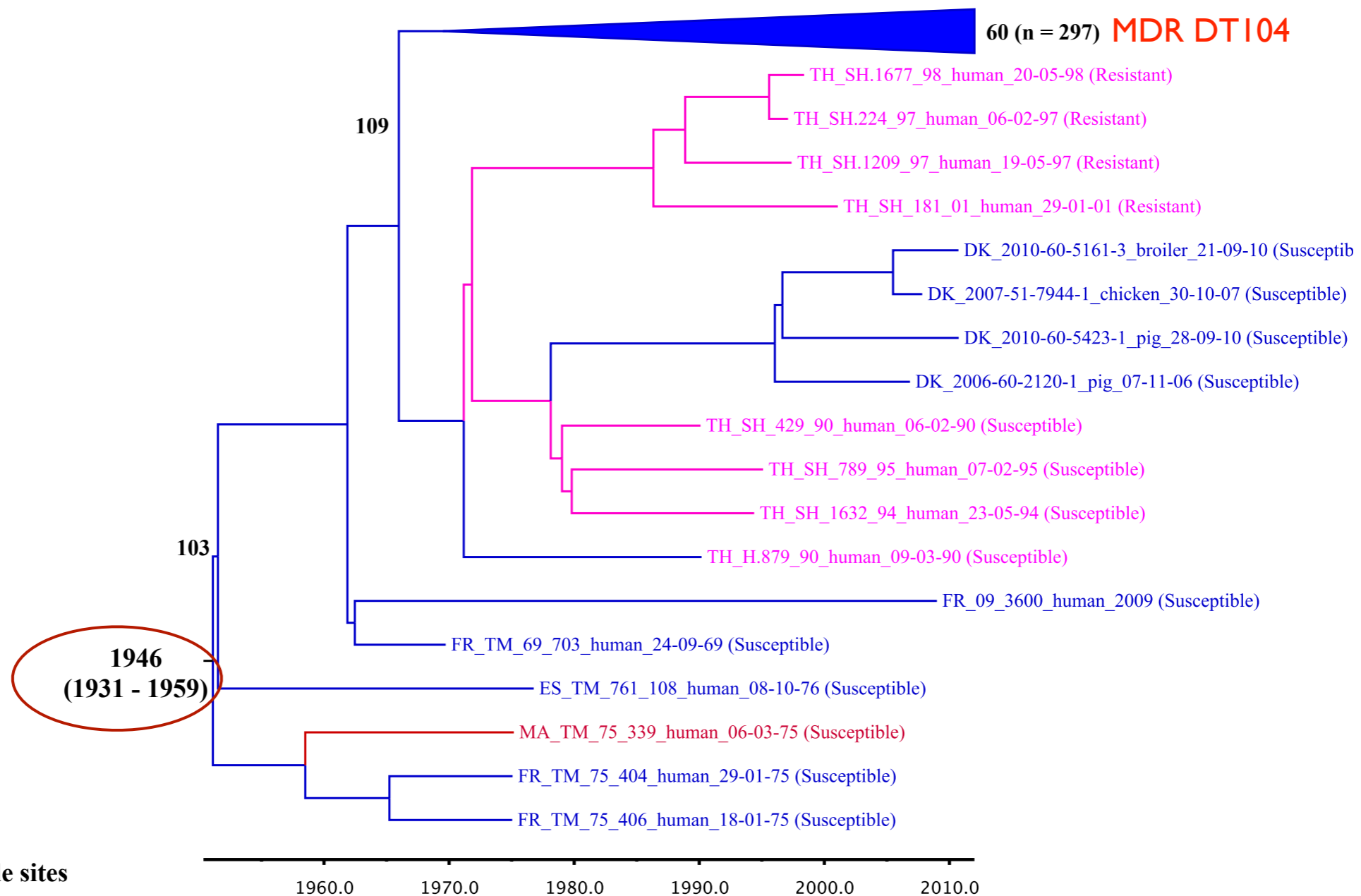
197 animal isolates

118 human isolates

BEAST

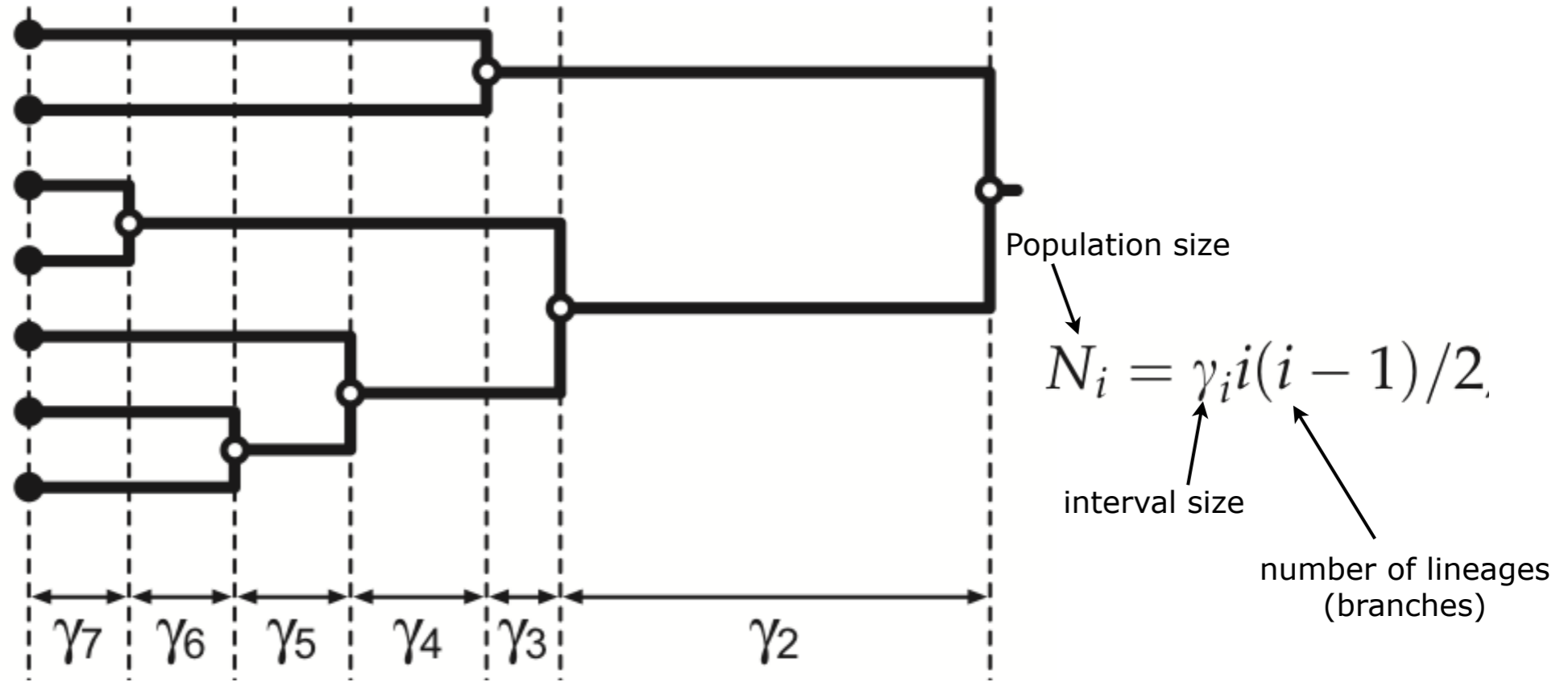


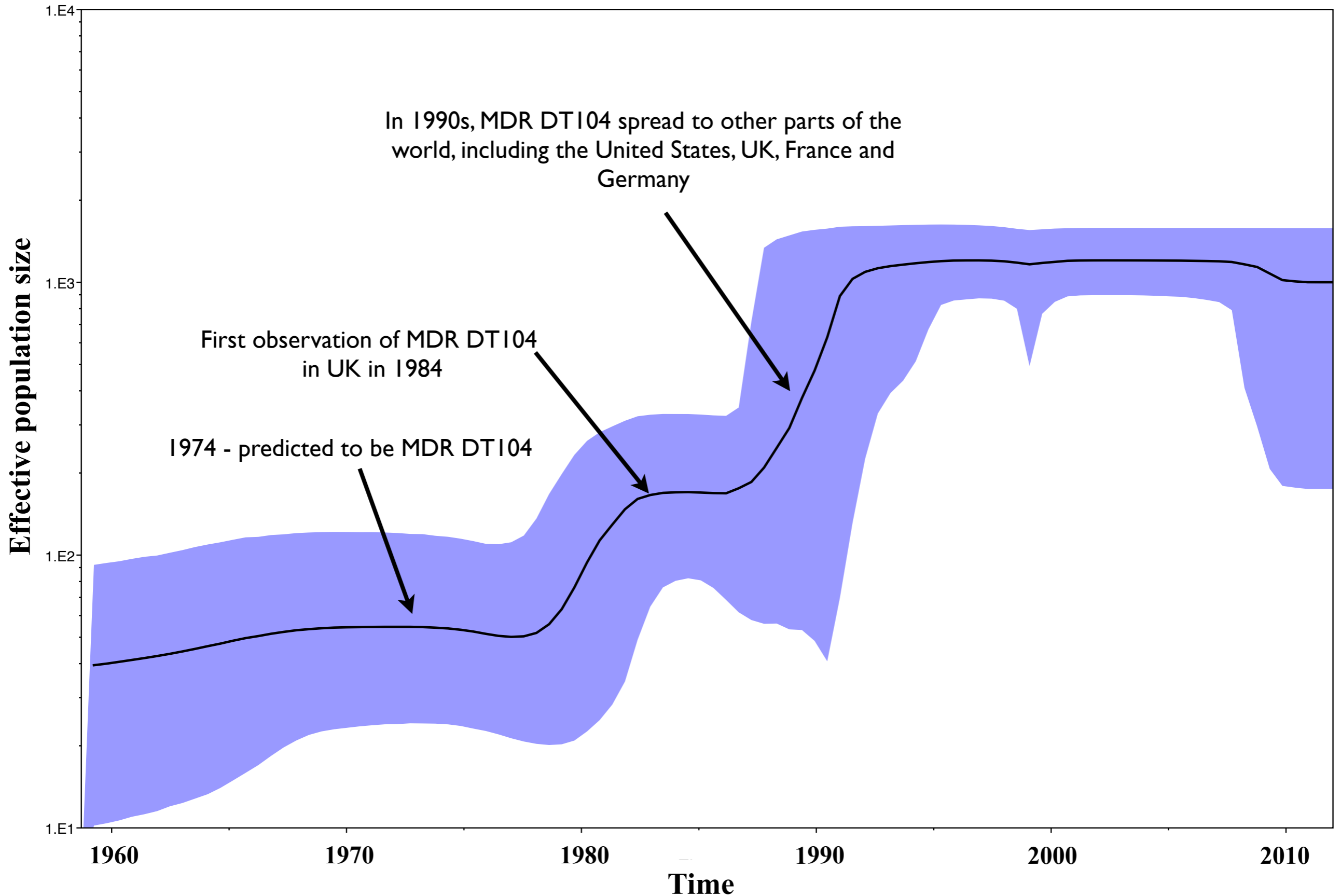
- Europe
- North America
- South America
- Asia
- Australia



4619 variable sites

2.97×10^{-7} SNP/site/year ~ 1.47 SNP/year





Search

Get Directions History

our Buildings ed es able_fasta_danish...

3/1960

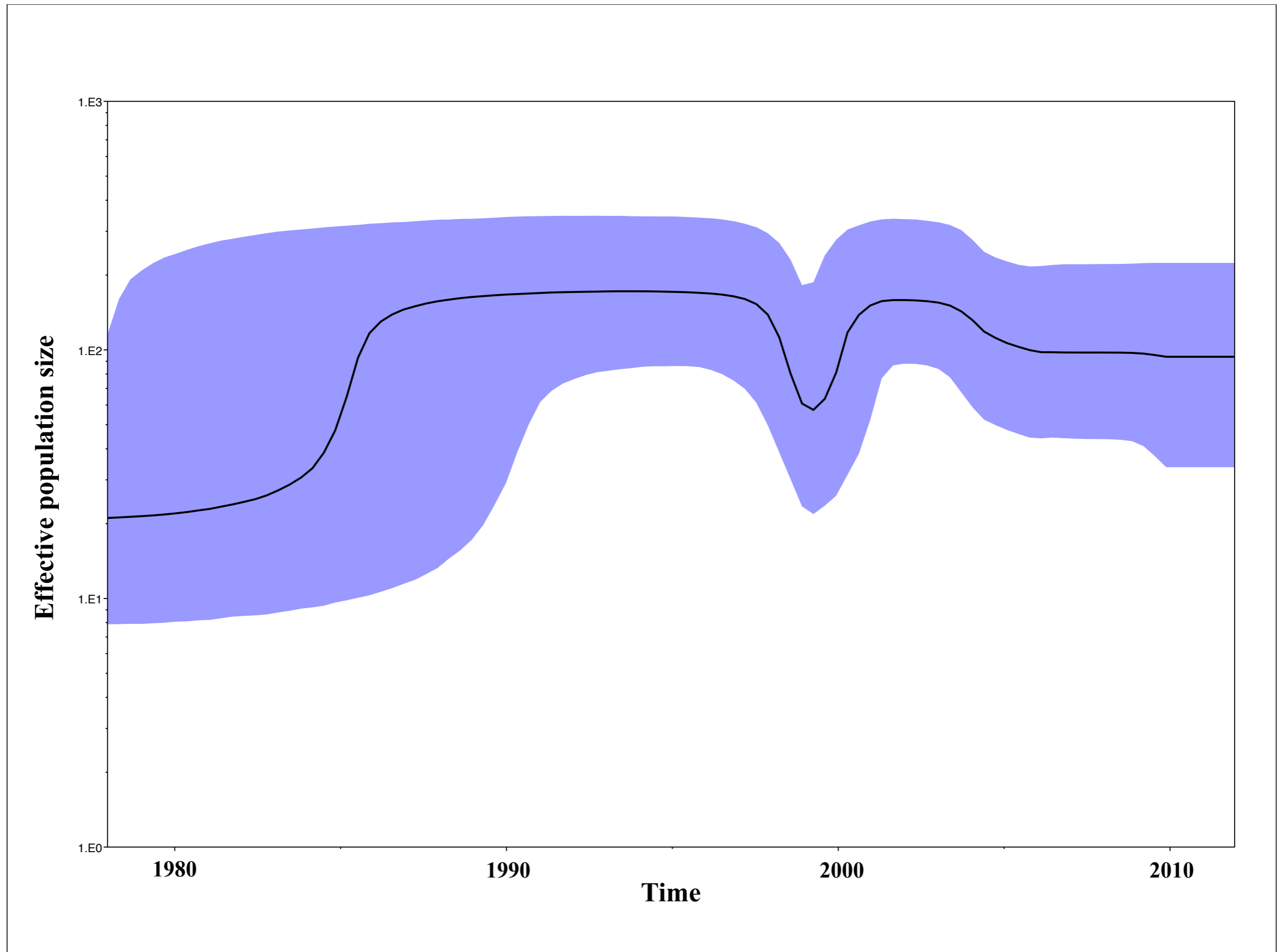
1960 2011



© 2009 GeoBasis-DE/BKG
 Data SIO, NOAA, U.S. Navy, NGA, GEBCO
 © 2013 Google
 Image Landsat

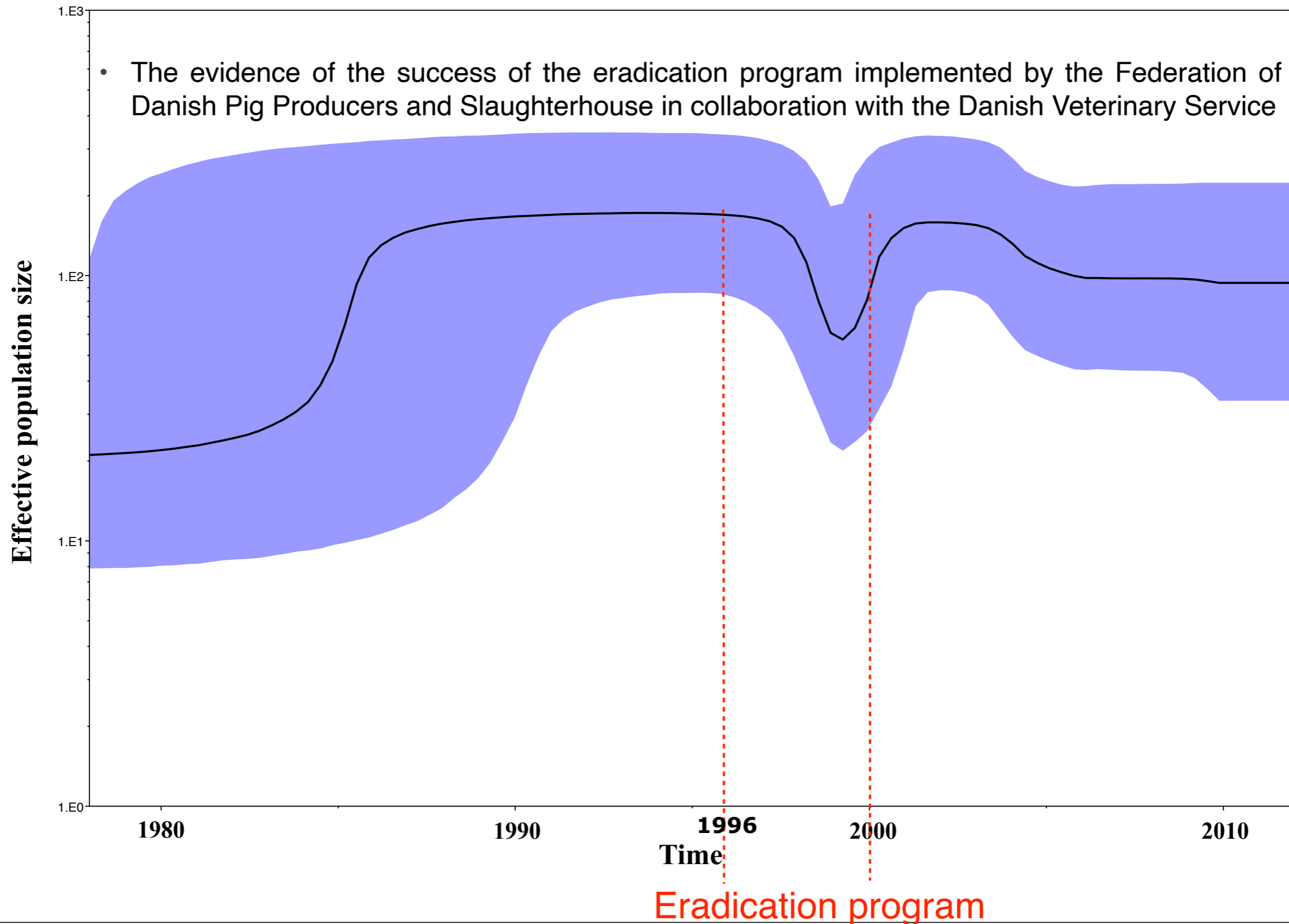
Google earth

Demographic history of Danish MDR DT104

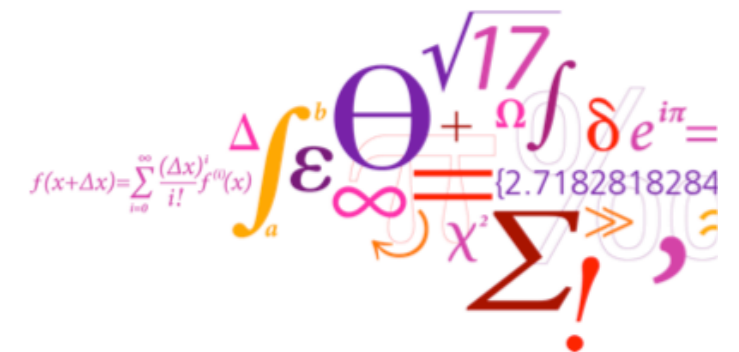


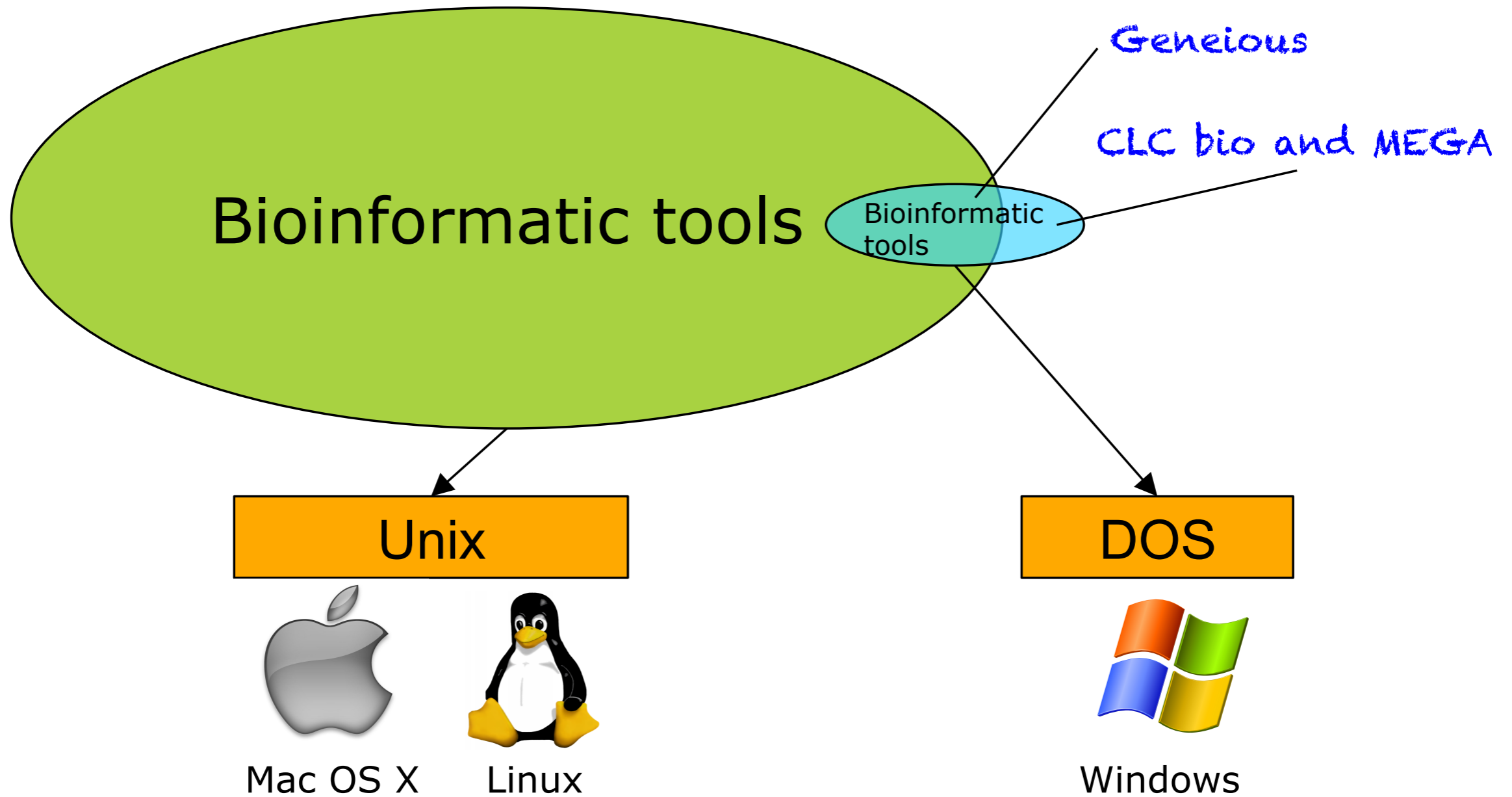
- The program aimed to eradicate MDR DT104 from infected pig herds by depopulation of pig herds, cleaning and disinfection of building before repopulation with pigs free from DT104

- The evidence of the success of the eradication program implemented by the Federation of Danish Pig Producers and Slaughterhouse in collaboration with the Danish Veterinary Service



Bioinformatics tools for surveillance of infectious diseases





Center for Genomic Epidemiology

Home

Organization

Project

Services

Contact

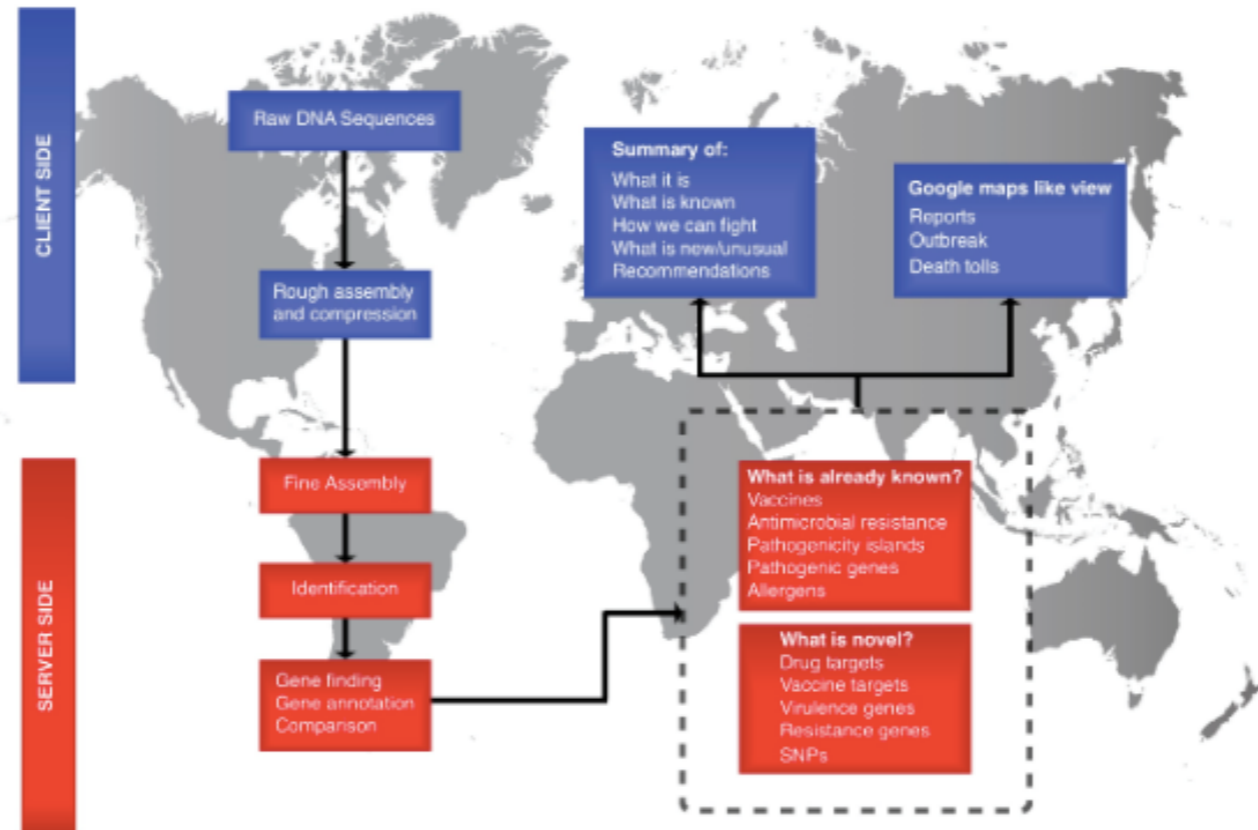
Services

Phenotyping:

- Identification of acquired antibiotic resistance genes. [ResFinder](#)
- Prediction of a bacteria's pathogenicity towards human hosts. [PathogenFinder](#)
- Identification of acquired virulence genes. [VirulenceFinder](#)

Typing:

- Multi Locus Sequence Typing (MLST) from an assembled genome or from a set of reads [MLST](#)
- PlasmidFinder identifies plasmids in total or partial sequenced isolates of bacteria. [PlasmidFinder](#)
- Multi Locus Sequence Typing (MLST) from an assembled plasmid or from a set of reads [pMLST](#)
- Prediction of bacterial species using a fast K-mer algorithm. [KmerFinder](#)
- Prediction of bacterial species using the S16 ribosomal DNA sequence. [SpeciesFinder](#)



Welcome to the Center for Genomic Epidemiology

The cost of sequencing a bacterial genome is \$50 and is expected to decrease further in the near future and the equipment needed cost less than \$150 000. Thus, within a few years all clinical microbiological laboratories will have a sequencer in use on a daily basis. The price of genome sequencing is already so low that whole genome sequencing will also find worldwide application in human and veterinary practices as well as many other places where bacteria are handled. In Denmark alone this equals more than 1 million isolates annually in 15-20 laboratories and globally up to 1-2 billion isolates per year. The limiting factor will therefore in the future not be the cost of the sequencing, but how to assemble, process and handle the large amount of data in a standardized way that will make the information useful, especially for diagnostic and surveillance.

News

Course on the use of the CGE tools in November 2014

September 2014
The course is for clinical microbiologists to learn how to use the CGE tools. The course will be taught in English and take place at the Technical University of Denmark [Course flyer \(pdf\)](#)

Benchmarking of Methods for Genomic Taxonomy

April 2014
How to optimally determine taxonomy from whole genome sequences. [Link to article...](#)

CGE tools applied for bacteriophage characterization

March 2014
Applying the ResFinder and VirulenceFinder web-services for easy identification of acquired antibiotic resistance and E. coli virulence genes in bacteriophage and prophage nucleotide sequences. [Link to article...](#)

Evaluation of Whole Genome Sequencing for Outbreak Detection of Salmonella enterica

March 2014
We evaluated WGS for outbreak detection of Salmonella enterica including different approaches for analyzing and comparing with a traditional typing, PFGE. [Link to article...](#)

Low-bandwidth and non-compute intensive remote identification of microbes from raw sequencing reads

January 2014
Cheap dna sequencing may soon become routine not only for human

Center for Genomic Epidemiology

Username
Password

Home

Services

Datasets

User Home

Overview of Services

Workflows

[Bacterial Analysis Batch Upload Pipeline](#) (Works)

Phenotyping

[ResFinder](#) (Works)
[PathogenFinder](#) (Works)
[VirulenceFinder](#) (Works)
[Restriction-ModificationFinder](#) (Works)

Typing

[SeqSero](#) (Works)
[SerotypeFinder](#) (Works)
[PAst](#) (in development)
[VirusFinder](#) (in development)
[spaTyper](#) (Works)
[MLST](#) (Works)
[pMLST](#) (Works)
[PlasmidFinder](#) (Works)
[KmerFinder](#) (Works)
[SpeciesFinder](#) (Works)
[Read2Type](#) (This service is not implemented on the new server)
[TaxonomyFinder](#) (This program is in development)
[Tapir](#) (This service is not implemented on the new server)

Phylogeny

[snpTree](#) (Works)
[NDtree](#) (Works)
[CSIPhylogeny](#) (Works)
[TreeViewer](#) (Works)

Other

[Assembler](#) (Works)
[ENAUUploader](#) (in development)
[PanFunPro](#) (Works)
[MGmapper](#) (Works)
[MyDbFinder](#) (Works)
[SPIFinder](#) (Works)
[HostPhinder](#) (in development)
[GeneticDiseaseProject](#) (Not associated with CGE)
[NetFCM](#) (Not associated with CGE)



ResFinder

<https://cge.food.dtu.dk/services/ResFinder/>

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Overview of genes

Article abstract

ResFinder 3.0

ResFinder identifies acquired antimicrobial resistance genes and/or find chromosomal mutations in total or partial sequenced isolates of bacteria.

View the [version history](#) of this server.

The database is curated by:
Valeria Bortolaia
(click to contact)

Chromosomal point mutations

Acquired antimicrobial resistance genes

Select type of your reads

Assembled Genome/Contigs*

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

Name	Size	Progress	Status

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Overview of genes

Article abstract

ResFinder 3.0

ResFinder identifies acquired antimicrobial resistance genes and/or find chromosomal mutations in total or partial sequenced isolates of bacteria.

View the [version history](#) of this server.

The database is curated by:
Valeria Bortolaia
(click to contact)

Chromosomal point mutations

Resistance caused by mutations

Select species

- Campylobacter
- E. coli
- Salmonella
- N. gonorrhoeae
- M. tuberculosis

Acquired antimicrobial resistance genes

Select type of your reads

Assembled Genome/Contigs*

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Overview of genes

Article abstract

ResFinder 3.0

ResFinder identifies acquired antimicrobial resistance genes and/or find chromosomal mutations in total or partial sequenced isolates of bacteria.

View the [version history](#) of this server.

The database is curated by:
Valeria Bortolaia
(click to contact)

Chromosomal point mutations

Resistance caused by mutations

Select species

E. coli

Show unknown mutations

Show only known mutations
Show all mutations, known and unknown

Acquired antimicrobial resistance genes

Select type of your reads

Assembled Genome/Contigs*

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Overview of genes

Article abstract

ResFinder 3.0

ResFinder identifies acquired antimicrobial resistance genes and/or find chromosomal mutations in total or partial sequenced isolates of bacteria.

View the [version history](#) of this server.

The database is curated by:
Valeria Bortolaia
(click to contact)

Chromosomal point mutations

Resistance caused by mutations

Select species

Show unknown mutations

Show only known mutations
Show all mutations, known and unknown

Acquired antimicrobial resistance genes

Select Antimicrobial configuration

Select multiple items, with Ctrl-Click (or Cmd-Click on Mac) - by default all databases are selected

Aminoglycoside
Beta-lactam
Colistin
Fluoroquinolone
Fosfomycin
Fusidic Acid

Select threshold for %ID

Select minimum length

Chromosomal point mutations

Resistance caused by mutations

Select species

E. coli

Show unknown mutations

- Show only known mutations
- Show all mutations, known and unknown

Acquired antimicrobial resistance genes

Select type of your reads

Assembled Genome/Contigs*

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

Isolate File

Name	Size	Progress	Status
strain01.fasta	4.63 MB	<div style="width: 100%;"></div>	

Upload

Remove



Chromosomal point mutations - Results

Species: *e.coli*

Known Mutations

parE
No mutations found in parE

parC
No known mutations found in parC

folP
No mutations found in folP

gyrA				
Mutation	Nucleotide change	Amino acid change	Resistance	PMID
gyrA p.S83L	TCG → TTG	S → L	Quinolones, Fluoroquinolones	15848289

pmrB
No known mutations found in pmrB

pmrA
No mutations found in pmrA

16S_rrsB
No mutations found in 16S_rrsB

16S_rrsH
No known mutations found in 16S_rrsH

gyrB
No mutations found in gyrB

ampC				
Mutation	Nucleotide change	Amino acid change	Resistance	PMID
ampC promoter n.-42C>T	C → T	Promoter mutations	B-lactam resistance	21653764

Center for Genomic Epidemiology

Home Services Instructions Output Overview of genes Article abstract

ResFinder-2.1 Server - Results

Aminoglycoside						
Resistance gene	%Identity	Query/HSP length	Contig	Position in contig	Predicted phenotype	Accession number
<i>strA</i>	100.00	804 / 804	strain_1_contig_11	3559..4362	Aminoglycoside resistance Alternate name; aph(3'')-Ib	AF321551
<i>strB</i>	100.00	837 / 837	strain_1_contig_11	4362..5198	Aminoglycoside resistance Alternate name; aph(6)-IId	M96392

Beta-lactam						
Resistance gene	%Identity	Query/HSP length	Contig	Position in contig	Predicted phenotype	Accession number
<i>blaCTX-M-15</i>	100.00	876 / 876	strain_1_contig_14	81110..81985	Beta-lactam resistance Alternate name; UOE-1	DQ302097
<i>blaTEM-1B</i>	100.00	861 / 861	strain_1_contig_14	84807..85667	Beta-lactam resistance Alternate name; RblaTEM-1	JF910132

Colistin

No resistance genes found.

RAPID COMMUNICATIONS

Detection of *mcr-1* encoding plasmid-mediated colistin-resistant *Escherichia coli* isolates from human bloodstream infection and imported chicken meat, Denmark 2015

H Hasman¹, AM Hammerum¹, F Hansen¹, RS Hendriksen², B Olesen³, Y Agersø², E Zankari², P Leekitcharoenphon², M Stegger^{1,4}, RS Kaas², LM Cavaco², DS Hansen³, FM Aarestrup², RL Skov¹

1. Department of Microbiology and Infection Control, Statens Serum Institut, Copenhagen, Denmark

2. National Food Institute, Technical University of Denmark, Lyngby, Denmark

3. Department of Clinical Microbiology, Herlev and Gentofte Hospital, Copenhagen University Hospital, Herlev, Denmark

4. Pathogen Genomics Division, Translational Genomics Research Institute (TGen), Flagstaff, Arizona, USA

Correspondence: Henrik Hasman (henh@ssi.dk)

Citation style for this article:

Hasman H, Hammerum A, Hansen F, Hendriksen R, Olesen B, Agersø Y, Zankari E, Leekitcharoenphon P, Stegger M, Kaas R, Cavaco L, Hansen D, Aarestrup F, Skov R. Detection of *mcr-1* encoding plasmid-mediated colistin-resistant *Escherichia coli* isolates from human bloodstream infection and imported chicken meat, Denmark 2015. *Euro Surveill.* 2015;20(49):pii=30085. DOI: <http://dx.doi.org/10.2807/1560-7917.ES.2015.20.49.30085>

Article submitted on 04 December 2015 / accepted on 10 December 2015 / published on 10 December 2015

"The approximately 3,000 Gram-negative (*E. coli* or *Salmonella*) bacteria, which have previously been mapped using whole genome sequencing, have been reexamined to see whether MCR-1 is present. Results show that MCR-1 was found in one patient, who suffered from a blood infection in 2015 and in five food samples that have been imported from 2012-2014. All the bacteria are multi-resistant ESBL bacteria containing the MCR-1 gene, which can further complicate treatment."



CSIPhylogeny

<https://cge.food.dtu.dk/services/CSIPhylogeny/>

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Article abstract

CSI Phylogeny 1.1 (Call SNPs & Infer Phylogeny)

CSI Phylogeny calls SNPs, filters the SNPs, does site validation and infers a phylogeny based on the concatenated alignment of the high quality* SNPs.

Note: The old version of this service is still available at: [CSI Phylogeny 1.0a](#). But it is now deprecated and no longer supported.

Service updated (14:30 10-Mar-2016 GMT+1). Service was down for several days due to errors in the queing system. The downtime was exploited to implement a new queing method for this service. It has been tested and should work but please don't hesitate to write Scientific support if your jobs are failing. The update does not affect output results, only where the pipeline is executed on the CGE server.

Input data

Upload reference genome (fasta format)

Note: Reference genome must not be compressed.

no file selected

Include reference in final phylogeny.

Select min. depth at SNP positions

10x

Select min. relative depth at SNP positions

10 %

Select minimum distance between SNPs (prune)

10 bp

Select min. SNP quality

30

Select min. read mapping quality

25

Select min. Z-score

1.96

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Article abstract

CSI Phylogeny 1.1 (Call SNPs & Infer Phylogeny)

CSI Phylogeny calls SNPs, filters the SNPs, does site validation and infers a phylogeny based on the concatenated alignment of the high quality* SNPs.

Note: The old version of this service is still available at: [CSI Phylogeny 1.0a](#). But it is now deprecated and no longer supported.

Service updated (14:30 10-Mar-2016 GMT+1). Service was down for several days due to errors in the queing system. The downtime was exploited to implement a new queing method for this service. It has been tested and should work but please don't hesitate to write Scientific support if your jobs are failing. The update does not affect output results, only where the pipeline is executed on the CGE server.

Input data

Upload reference genome (fasta format)

Note: Reference genome must not be compressed.

no file selected

Include reference in final phylogeny.

Select min. depth at SNP positions

10x

Select min. relative depth at SNP positions

10 %

Select minimum distance between SNPs (prune)

10 bp

Select min. SNP quality

30

Select min. read mapping quality

25

Select min. Z-score

1.96

Center for Genomic Epidemiology

Username
Password

Home

Services

Instructions

Output

Article abstract

CSI Phylogeny 1.1 (Call SNPs & Infer Phylogeny)

CSI Phylogeny calls SNPs, filters the SNPs, does site validation and infers a phylogeny based on the concatenated alignment of the high quality* SNPs.

Note: The old version of this service is still available at: [CSI Phylogeny 1.0a](#). But it is now deprecated and no longer supported.

Service updated (14:30 10-Mar-2016 GMT+1). Service was down for several days due to errors in the queing system. The downtime was exploited to implement a new queing method for this service. It has been tested and should work but please don't hesitate to write Scientific support if your jobs are failing. The update does not affect output results, only where the pipeline is executed on the CGE server.

Input data

Upload reference genome (fasta format)

Note: Reference genome must not be compressed.

 D23580.fasta

Include reference in final phylogeny.

Select min. depth at SNP positions

10x

Select min. relative depth at SNP positions

10 %

Select minimum distance between SNPs (prune)

10 bp

Select min. SNP quality

30

Select min. read mapping quality

25

Select min. Z-score

1.96


Use altered FastTree (more accurate)

Note: Read more [here](#)



Upload read files and/or assembled genomes (fasta or fastq format)

Note: Read files must be compressed with gzip (compressed files often ends with .gz).

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

 Isolate File

Name	Size	Progress	Status
<hr/>			
<hr/>			

 Upload  Remove

***High quality SNPs**

A high quality SNP are defined as a SNP that obeys the following rules:

Confidentiality:

The sequences are kept confidential and will be deleted after 48 hours.

Use altered FastTree (more accurate)

Note: Read more [here](#)

Upload read files and/or assembled genomes (fasta or fastq format)

Note: Read files must be compressed with gzip (compressed files often ends with .gz).

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

Isolate File

Name	Size	Progress	Status
Salmonella-spp-02-03-002.fna	4.80 MB	<div style="width: 100%;"></div>	
Salmonella-spp-02-03-008.fna	4.81 MB	<div style="width: 100%;"></div>	
Salmonella-spp-05-102.fna	4.81 MB	<div style="width: 100%;"></div>	
Salmonella-spp-07-022.fna	4.80 MB	<div style="width: 100%;"></div>	

Upload

Remove



***High quality SNPs**

A high quality SNP are defined as a SNP that obeys the following rules:

Confidentiality:

The sequences are kept confidential and will be deleted after 48 hours.

CITATIONS

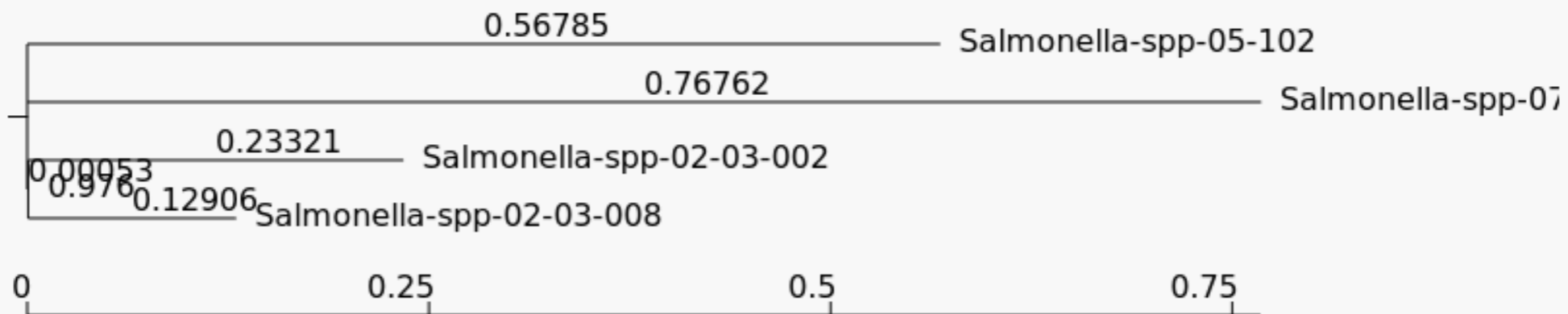
Center for Genomic Epidemiology

- Home
- Services
- Instructions
- Output
- Article abstract

Mapper: BWA # Submitting 7 jobs. Waiting for vcfwiz.sh to finish... 0

CSIPhylogeny Results

The tree presented in the picture below is only meant as a preview. If the tree is meant to be shared or published, we strongly recommend that the 'Newick' file is downloaded and processed using software created for this purpose. We suggest [FigTree](#).



Download phylogeny as:

Download the filtered SNP calls in Variant Calling Format (VCF):

Note: VCF files are compressed with gzip.

Download matrix of SNP pair counts:

Download matrix as:

Download SNP alignment:

https://bitbucket.org/account/user/genomicepidemiology/projects/CGE

The screenshot shows the Bitbucket interface for the user 'Genomic Epidemiology'. The left sidebar contains navigation options: 'Repositories' (selected), 'Projects', and 'Members'. The main content area is titled 'Repositories' and includes a 'Language' dropdown menu, a 'Create repository' button, and a search bar labeled 'Find repositories'. Below this is a table listing various repositories.

Repository	Project	Last updated	Builds
assimpler2	CGE	an hour ago	
adv_bioinf	Teaching	3 hours ago	
resfinder_db	Databases	4 hours ago	
ccphylo	CGE	7 hours ago	
mlst_db	Databases	15 hours ago	
kma	CGE	3 days ago	
CGE Tools Docker	CGE	2019-12-27	
pyCoDa	CGE	2019-12-20	
cge_core_module	CGE	2019-12-20	
mydbfinder	CGE	2019-12-18	
CholeraeFinder	CGE	2019-12-18	
intfinder	CGE	2019-12-18	
PlasmidFinder	CGE	2019-12-18	
VirulenceFinder	CGE	2019-12-18	
ResFinder	CGE	2019-12-18	
cgMLSTFinder	CGE	2019-12-05	

- ResFinder
- Source
- Commits
- Branches
- Pull requests
- Pipelines
- Deployments
- Issues
- Downloads

Genomic Epidemiology / CGE

ResFinder

master



Name	Size	Last commit	Message
.gitignore	34 B	2015-07-29	Update
README.md	5.13 KB	2019-08-09	Warning Biopython
resfinder.pl	60.49 KB	2018-10-04	Script updated
resfinder.py	25.01 KB	2019-12-18	fix bug multiple and no hit
test.fsa	4.25 MB	2015-07-16	Updated

Installation

Setting up ResFinder script and database

```
# Go to wanted location for resfinder  
cd /path/to/some/dir  
  
# Clone and enter the resfinder directory  
git clone https://git@bitbucket.org/genomicepidemiology/resfinder.git  
cd resfinder  
  
# Installing up the ResFinder database  
# Go to wanted location for resfinder database  
cd /path/to/some/dir  
  
# Clone and enter the resfinder directory  
git clone https://git@bitbucket.org/genomicepidemiology/resfinder_db.git  
cd resfinder_db
```

Usage

You can run resfinder command line using python3

Example of running resfinder

```
python3 resfinder.py -i test.fsa -o . -p /path/to/resfinder_db \
-mp /path/to/blastn -d aminoglycoside -t 0.90 -l 0.60
```

The program can be invoked with the -h option

```
Usage: resfinder.py [-h] [-i INPUTFILE] [-o OUT_PATH]
                  [-tmp TMP_DIR] [-mp METHOD_PATH] [-ao ACQ_OVERLAP]
                  [-matrix MATRIX] [-p DB_PATH] [-d DATABASES] [-l MIN_COV]
                  [-t THRESHOLD] [-x] [-q]
```

optional arguments:

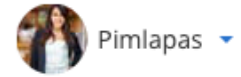
```
-h, --help          show this help message and exit
-i INPUTFILE, --inputfile INPUTFILE
                    Input file (fasta or fastq(s) files)
-o OUT_PATH, --outputPath OUT_PATH
                    Path to blast output
-p DB_PATH, --databasePath DB_PATH
                    Path to the databases
-mp METHOD_PATH --methodPath METHOD_PATH
                    Path to the method to use (kma or blastn)
-d DATABASES, --databases DATABASES
                    Databases chosen to search in - if none are specified
                    all are used
-l MIN_COV, --min_cov MIN_COV
                    Minimum coverage default 0.6
-t THRESHOLD, --threshold THRESHOLD
                    Blast threshold for identity
                    default minimum 0.9
-ao ACQ_OVERLAP --acq_overlap ACQ_OVERLAP
                    Genes are allowed to overlap this number of nucleotides (30)
-matrix, --matrix
                    If used, gives the counts all all called bases at each position
                    in each mapped template. Columns are: reference base,
                    A count, C count, G count, T count, N count, - count.
-x --extended_output
                    If used, give extended output with alignment files,
                    "template and query hits in fasta and a tab
                    "seperated file with gene profile results
-q --quiet
```

<https://www.coursera.org/learn/wgs-bacteria/>



Catalog Search catalog

For Enterprise



Home > Life Sciences > Clinical Science

Whole Genome Sequencing of bacterial genomes – tools and applications

- Overview
- Syllabus
- FAQs
- Creators
- Ratings and Reviews

Whole genome sequencing of bacterial genomes – tools and applications

[Go to Course](#)

Already enrolled

Financial Aid is available for learners who cannot afford the fee. [Learn more and apply.](#)

About this course: This course will cover the topic of Whole genome sequencing (WGS) of bacterial genomes which is becoming more and more relevant for the medical sector. WGS technology and applications are high on international political agenda, as the classical methods are being replaced by WGS technology and therefore bioinformatic tools are extremely important for allowing the people working in

[More](#)

Who is this class for: This course is for you if you are interested in getting to know more about Whole genome sequencing applied to bacterial characterization and surveillance. We aim at having a broad scope and international reach in different sectors. So this course is for you whether you are an undergraduate or graduate student, a researcher, medical or veterinary related professional, technical staff or simply interested in the subject!

Created by: Technical University of Denmark (DTU)

23262 Infectious disease bioinformatics

Course information

Danish title	Bioinformatik for smitsomme sygdomme
Language of instruction	English
Point(ECTS)	5
Course type	MSc Offered as a single course Technological Specialization Course, MSc. Eng., Bioinformatics and System Biologi
Schedule	Spring F3A (Tues 8-12)
Location	Campus Lyngby
Scope and form	Lectures and computer-based exercises.
Duration of Course	13 weeks
Type of assessment	Oral examination Project presentation in groups followed by individual questions based on the presentation. One final grade for group presentation and individual questions.
Aid	All Aid
Evaluation	7 step scale , internal examiner
Previous Course	27683 and 36683
Not applicable together with	27683/ 36683
Recommended prerequisites	23205 , Unix/Linux background. Basic understanding of bioinformatics in sequence analysis and infectious disease.

23205 Fighting infectious diseases

Course information

Danish title	Bekæmpelse af infektionssygdomme
Language of instruction	English
Point(ECTS)	5
Course type	MSc Offered as a single course Technological specialization course, MSc. Eng., Quantitative Biology and Disease Modelling Technological specialization course, MSc. Eng., Biotechnology Technological specialization course, MSc. Eng., Pharmaceutical Design and Engineering Technological Specialization Course, MSc. Eng., Bioinformatics and System Biologi
Schedule	Autumn E4B (Fri 8-12)
Location	Campus Lyngby
Scope and form	Lectures, exercises, team work
Duration of Course	13 weeks
Date of examination	E4B
Type of assessment	Written examination and exercises Written examination (75%), Team presentation of a scientific article (25%)
Exam duration	3 hours
Aid	All Aid
Evaluation	7 step scale , internal examiner
Recommended prerequisites	23258.23256 , Knowledge in microbial diversity and the basic biological processes related to molecular microbiology, microbial physiology, and host immune defenses. Basic microbiological experimental skills/knowledge, such as microbial cultivation and PCR. Basic statistical skills/knowledge, such as understanding probability and interpreting results of statistical tests as applied in e.g. experimental and epidemiological studies.



Pimlapas Leekitcharoenphon (Shinny)
Research Group for Genomic Epidemiology, DTU-Food

pile@food.dtu.dk