

PSI-BLAST

Fishing in the (sequence) twilight zone

Introduction to Bioinformatics,
Faroe Islands 2024
Bent Petersen

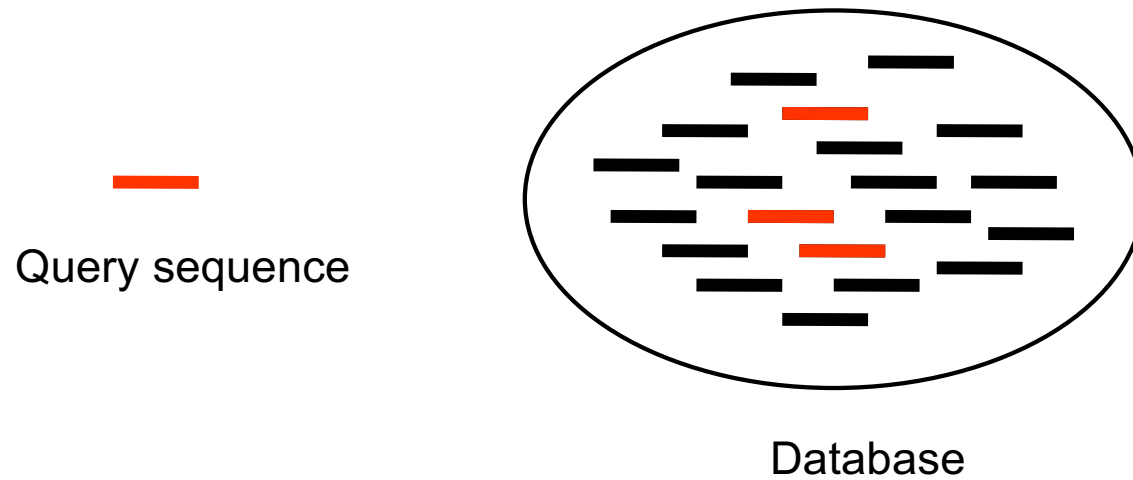
(With some borrowed concepts / slides from
Morten Nielsen, Rasmus Wernersson / Henrik Nielsen and Anders Gorm Pedersen)

Part 1

THE PROBLEM WITH PAIRWISE ALIGNMENTS

Reminder: how BLAST works

Use pairwise alignments to search
databases for similar sequences



BLASTP output

Sequences producing significant alignments:

Select: [All](#) [None](#) Selected:0

Alignments [Download](#) [GenPept](#) [Graphics](#) [Distance tree of results](#) [Multiple alignment](#)

	Description	Max score	Total score	Query cover	E value	Ident	Accession
<input type="checkbox"/>	putative uncharacterized protein [Odoribacter sp. CAG:788] >emb CCZ09189.1 putative uncharacterized pr	212	212	100%	6e-62	57%	WP_021987206.1
<input type="checkbox"/>	peptidase [Prevotella micans] >qb EHO65998.1 hypothetical protein HMPREF9140_02008 [Prevotella mic	207	207	99%	2e-58	55%	WP_006953704.1
<input type="checkbox"/>	hypothetical protein [Porphyromonas macacae]	204	204	99%	2e-57	53%	WP_018359894.1
<input type="checkbox"/>	putative uncharacterized protein [Odoribacter laneus CAG:561] >emb CCZ82493.1 putative uncharacterize	201	201	100%	1e-56	55%	WP_022048307.1
<input type="checkbox"/>	hypothetical protein [Odoribacter laneus] >qb EHP47141.1 hypothetical protein HMPREF9449_01748 [Odc	201	201	100%	2e-56	55%	WP_009136896.1
<input type="checkbox"/>	hypothetical protein [Porphyromonas somerae]	201	201	99%	2e-56	58%	WP_018029058.1
<input type="checkbox"/>	por secretion system C-terminal sorting domain protein [Porphyromonas sp. CAG:1061] >emb CCY10534.	201	201	99%	3e-56	58%	WP_021903554.1
<input type="checkbox"/>	hypothetical protein [Odoribacter laneus] >qb EHP45655.1 hypothetical protein HMPREF9449_02627 [Odc	199	199	100%	4e-56	54%	WP_009137771.1
<input type="checkbox"/>	hypothetical protein [Bacteroidales bacterium ph8]	198	198	99%	2e-55	53%	WP_019129881.1
<input type="checkbox"/>	putative uncharacterized protein [Odoribacter laneus CAG:561] >emb CCZ80898.1 putative uncharacterize	197	197	99%	2e-55	55%	WP_022047147.1
<input type="checkbox"/>	hypothetical protein [Porphyromonas levii]	195	195	100%	2e-54	53%	WP_018358555.1
<input type="checkbox"/>	putative uncharacterized protein [Odoribacter sp. CAG:788] >emb CCZ09222.1 putative uncharacterized pr	194	194	99%	2e-54	56%	WP_021987222.1
<input type="checkbox"/>	putative uncharacterized protein [Bacteroides sp. CAG:709] >emb CDA96737.1 putative uncharacterized pr	194	194	99%	3e-54	54%	WP_022147892.1
<input type="checkbox"/>	hypothetical protein [Porphyromonas macacae]	194	194	99%	6e-54	55%	WP_018360734.1

Alignment score (in bits)



(Example from the BLAST exercise: At the protein level it was quite evident, that the unknown sequence was a serine peptidase)

BLASTP alignment

Alignment score

>ref|WP_006953704.1| **peptidase** [Prevotella micans]
Length=922

Score = 207 bits (526) Expect = 2e-58, Method: Compositional matrix adjust.
Identities = 117/211 (55%), Positives = 145/211 (69%), Gaps = 14/211 (7%)

```
Query 2 GHGTHVAGTVA AVNNNGIGVAGVAGGNGSTNSGARLMSTQIFNSDGDYTNSETLVYRAIV 61
      GHGTHVAGTVAA NNGG+GVAG+AGG+GSTNSG RL+S QIF + ++E AI
Sbjct 279 GHGTHVAGTVA ARNNNGLGVAGIAGGDGSTNSGVRLLSQCIFRKSKEEGSAEA----AIK 334

Query 62 YGADNGAVISQNSWGSQSL-TIKELQKA---AIDYFIDYAGMDETGEIQT-GPMRGGIFI 116
      Y ADNGAVI+Q SWG S +KEL K+ AIDYFI +AG D G ++ PM+GG+ I
Sbjct 335 YAADNGAVIAQC SWGYASKENVKELPKSLKEAIDYFITFAGCDAHGAQRSDSPMKGGVMI 394

Query 117 AAAGNDNVSTPNMPSAYERVLAVASMGPDFTKASYSTFGTWT DITAPGGDIDKFDLSEYG 176
      AAGN+N++ P+AYE+V++VAS +F KASYS + W I+APGGD D F L + G
Sbjct 395 FAAGNENMNFKEFPAAAYEKVISVASTAWNFQKASYSNYADWVSI SAPGGDQDAFGL-KAG 453

Query 177 VLSTYADNY----YAYGEGTSMACPHVAGAA 203
      VLST Y Y +GTSMACPHV+G A
Sbjct 454 VLSTMPKKIASSGYGYMQGTSMACPHVSGIA 484
```

(Example from the BLAST exercise: At the protein level it was quite evident, that the unknown sequence was a serine peptidase)

Not all positions are biological equal

Conserved region:

Is likely important for the function of the enzyme

```
Query 2 GHGTHVAGTVA AVNNNGIGVAGVAGGNGSTNSGARLMSTQIEFNSDGDYTNSETLVYRAIV 61
Sbjct 279 GHGTHVAGTVAA NNG+GVAG+AGG+GSTNSG RL+S QIEF + ++E AI
GHGTHVAGTVA ARNNGGLGVAGIAGGDGSTNSGVRLLSQIEFRKSKEEGSAEA----AIK 334

Query 62 YGADNGAVISQNSWGSQSL-TIKELQKA---AIDYFIDYAGMDETGEIQT-GPMRGGIFI 116
Sbjct 335 Y ADNGAVI+Q SWG S +KEL K+ AIDYFI +AG D G ++ PM+GG+ I
YAADNGAVIAQC SWGYASKENVKELPKSLKEAIDYFITFAGCDAHGAQRSDSPMKGGVMI 394

Query 117 AAAGNDNVSTPNMPSAYERVLAVASMGPDFTKASYSTFGTWTDITAPGGDIDKFDLSEYG 176
Sbjct 395 AAGN+N++ P+AYE+V++VAS +F KASY S + W I+APGGD D F L + G
FAAGNENMNFKEFPAAAYEKVISVASTAWNFKASYSNYADWVVISAPGGDQDAFGL-KAG 453

Query 177 VLSTYADNY----YAYGEGTSMACPHVAGAA 203
Sbjct 454 VLST Y Y +GTSMACPHV+G A
VLSTMPKKIASSGYGYMQGTSMACPHVSGIA 484
```

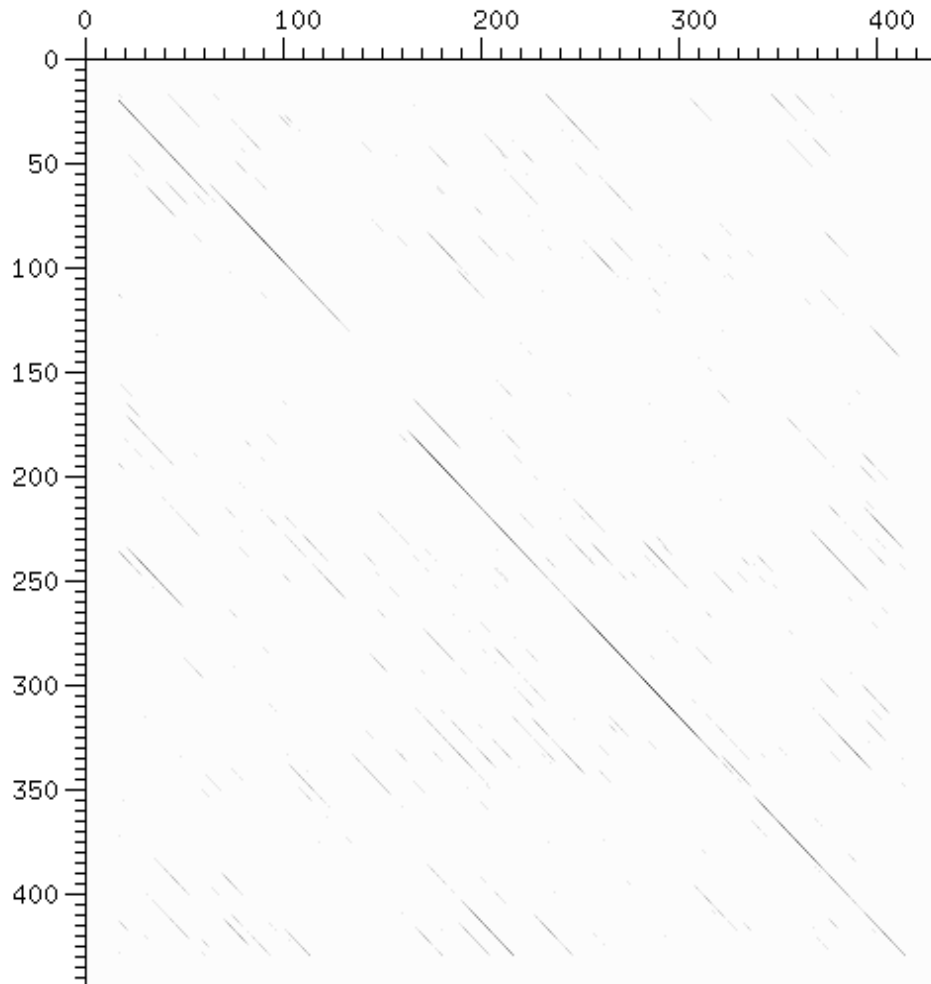
Variable region:

Is likely not that important for the function of the enzyme

Scoring of pairwise alignments

- In a normal pairwise alignment the same scores (the same matrix) is used for all positions
- As we saw before the selection pressure on the different parts of the sequence is not equal, and ideally we should take this into account
- **IMPORTANT:** if the sequences is of high enough similarity, this is usually not a big issue

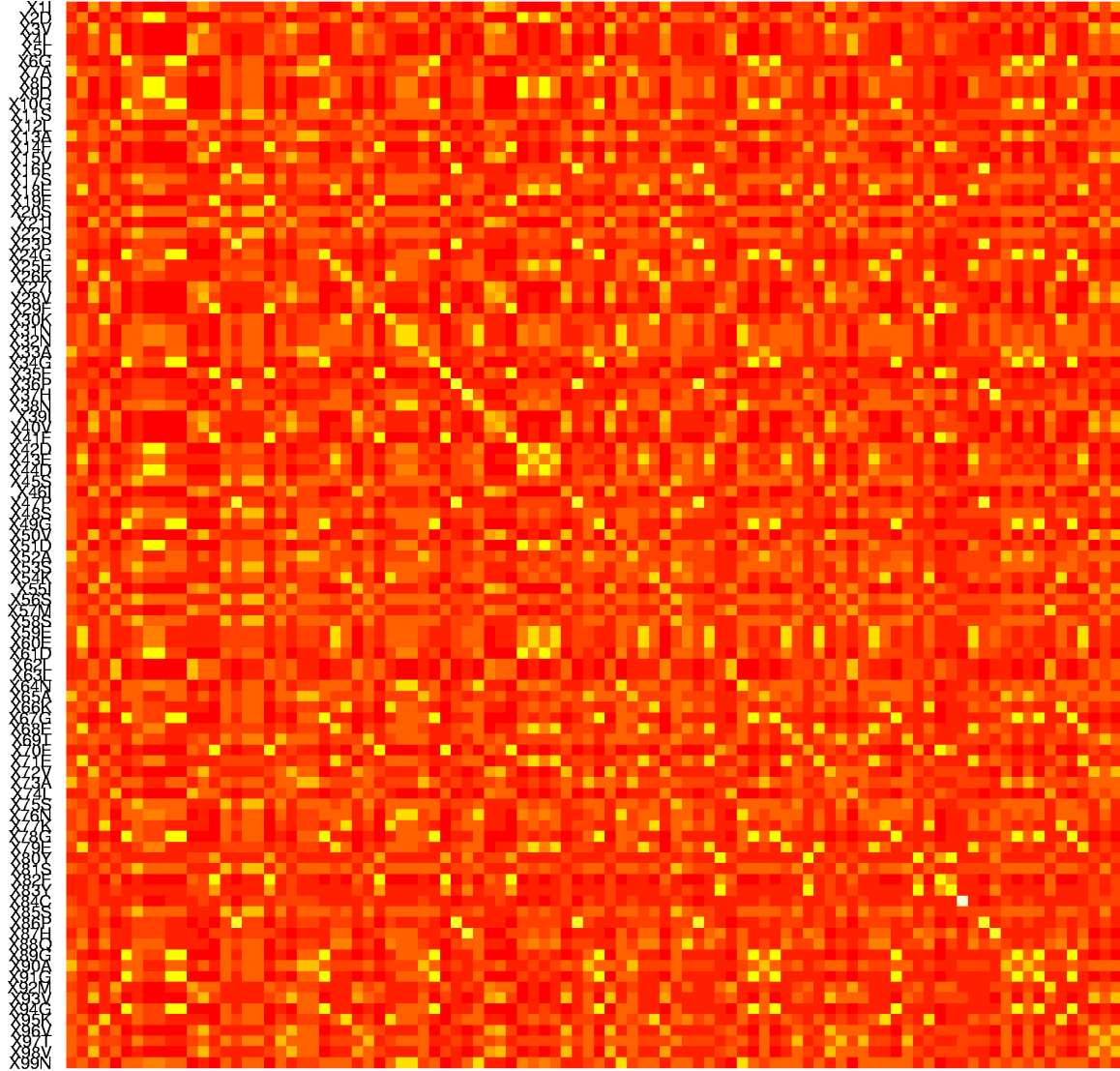
Reminder: Dot-plot



1. Place two sequences along axes of plot
2. Place dot at grid points where two sequences have identical residues
3. Diagonals correspond to conserved regions

Dot-plot with BLOSUM colors

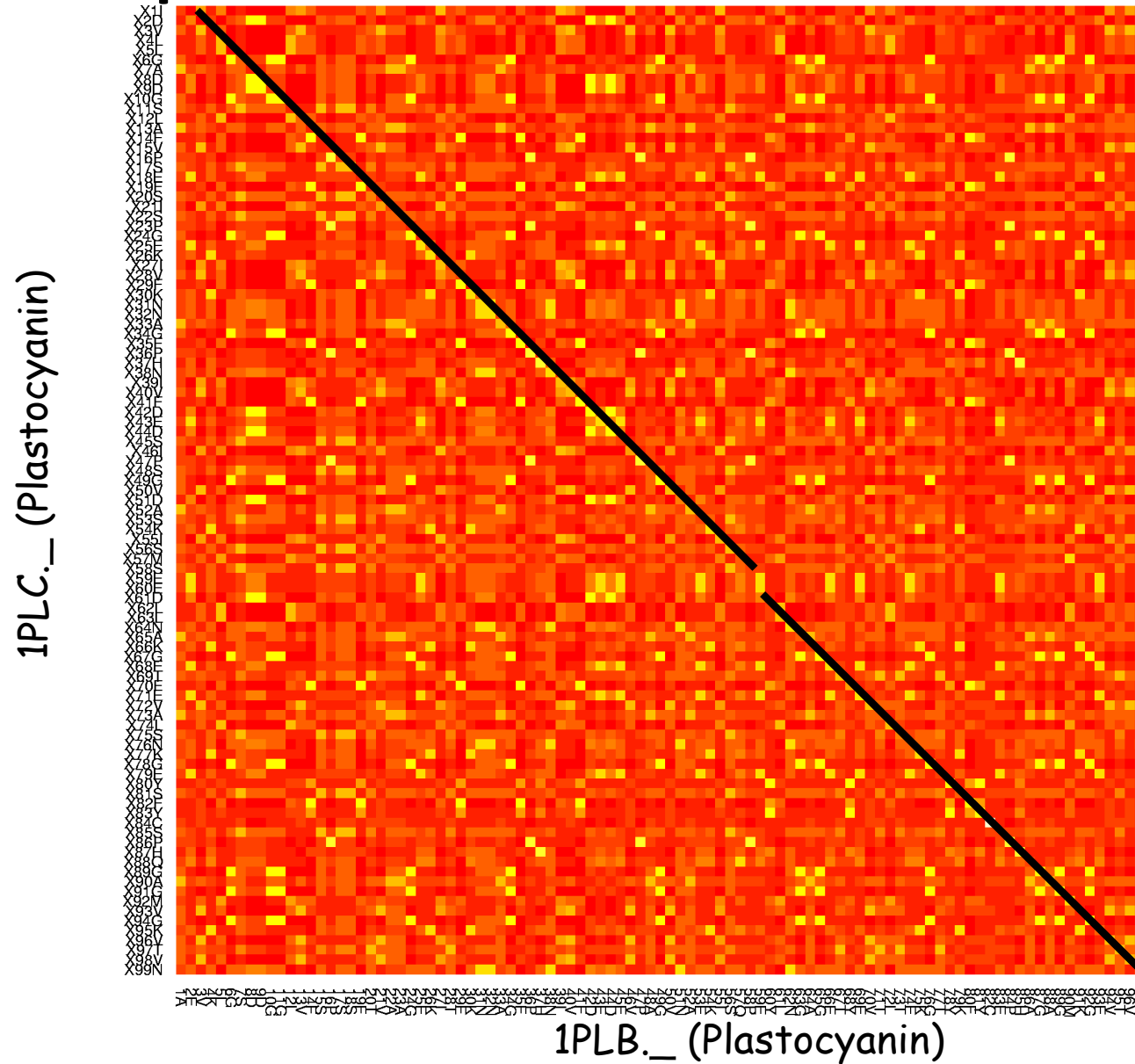
1PLC._ (Plastocyanin)



Relationship can be detected using BLASTP

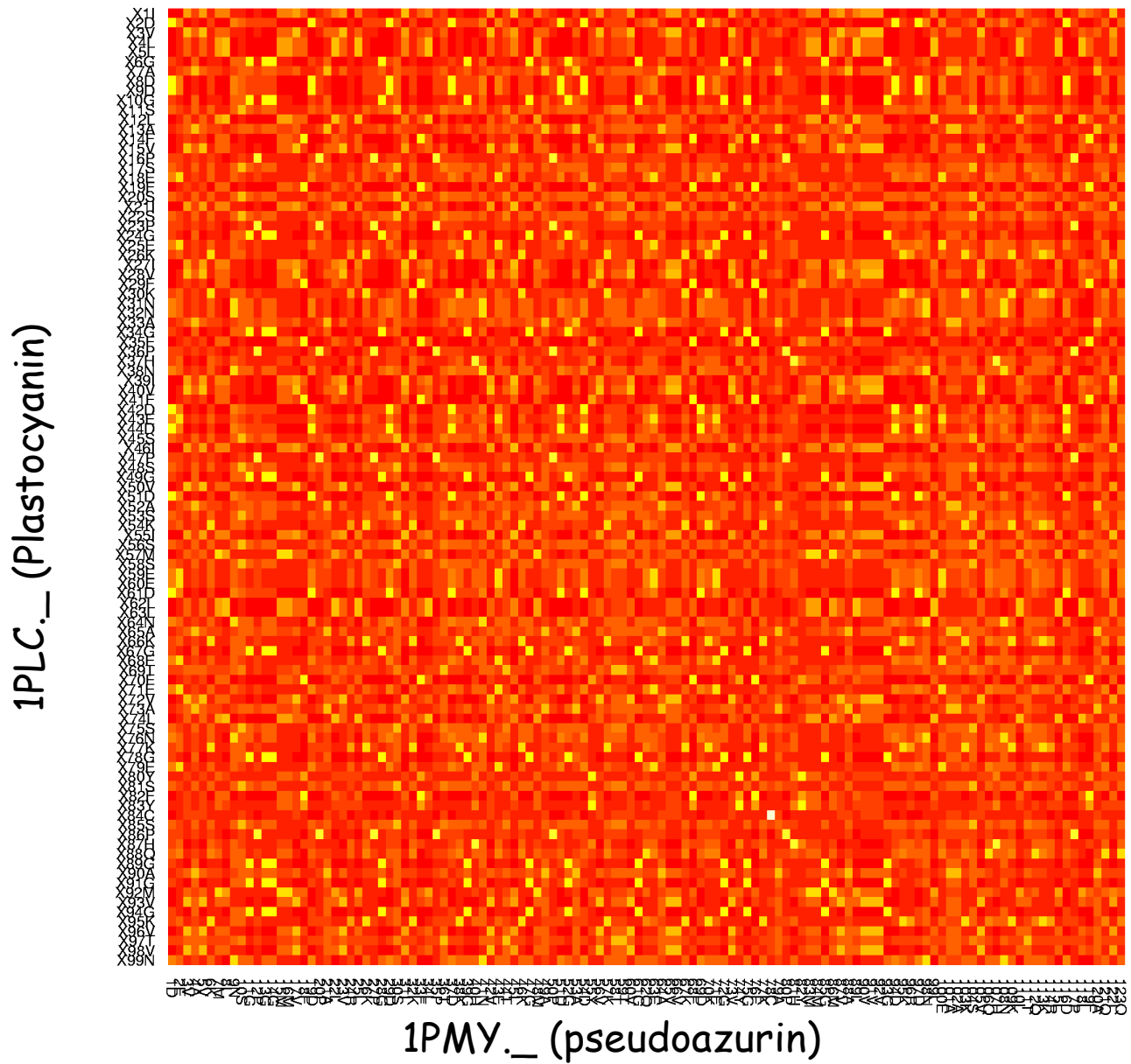
1PLB._ (Plastocyanin)

Dot-plot with BLOSUM colors



Relationship can
be detected
using BLASTP

Color dot-plot of low-similarity sequences



Relationship
CANNOT be
detected using
BLASTP

Part 2

THOUGHTS ABOUT HOW TO SOLVE THE PROBLEM

Idea catalog

- We would like to build a **scoring model** for pairwise alignments that more closely resembles what happens in **real sequence evolution**
 - Highly conserved sites/regions should have a high weight
 - Non-conserved regions should have a low weight (be allowed to vary without counting too much against the alignment score)
- **IMPORTANT:** Different protein families are under different selection pressure, so our model needs to account for this

Protein families

- Tools we can use, to identify the selective pressure on protein families:
 - Data sets of truly related proteins
 - Multiple alignment
 - Logo plots
 - Weight matrices

Protein family data sets

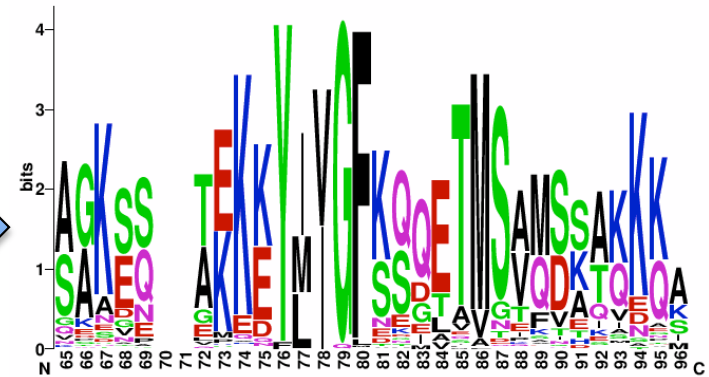
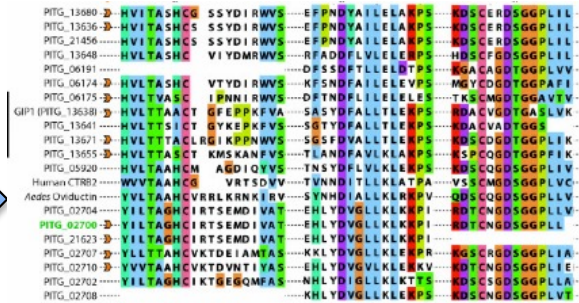
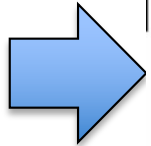
- How we can build such data sets:
 - Already known collections (literature, curated data sets)
 - Limitation: What have other people looked at before
 - “Text based” search in protein data bases (e.g. UniProt)
 - Limitation: Coverage, how well are the sequences described
 - BlastP (!)
 - Limitation: We only expect to find sequences of moderate to high similarity

Signal across multiple sequences

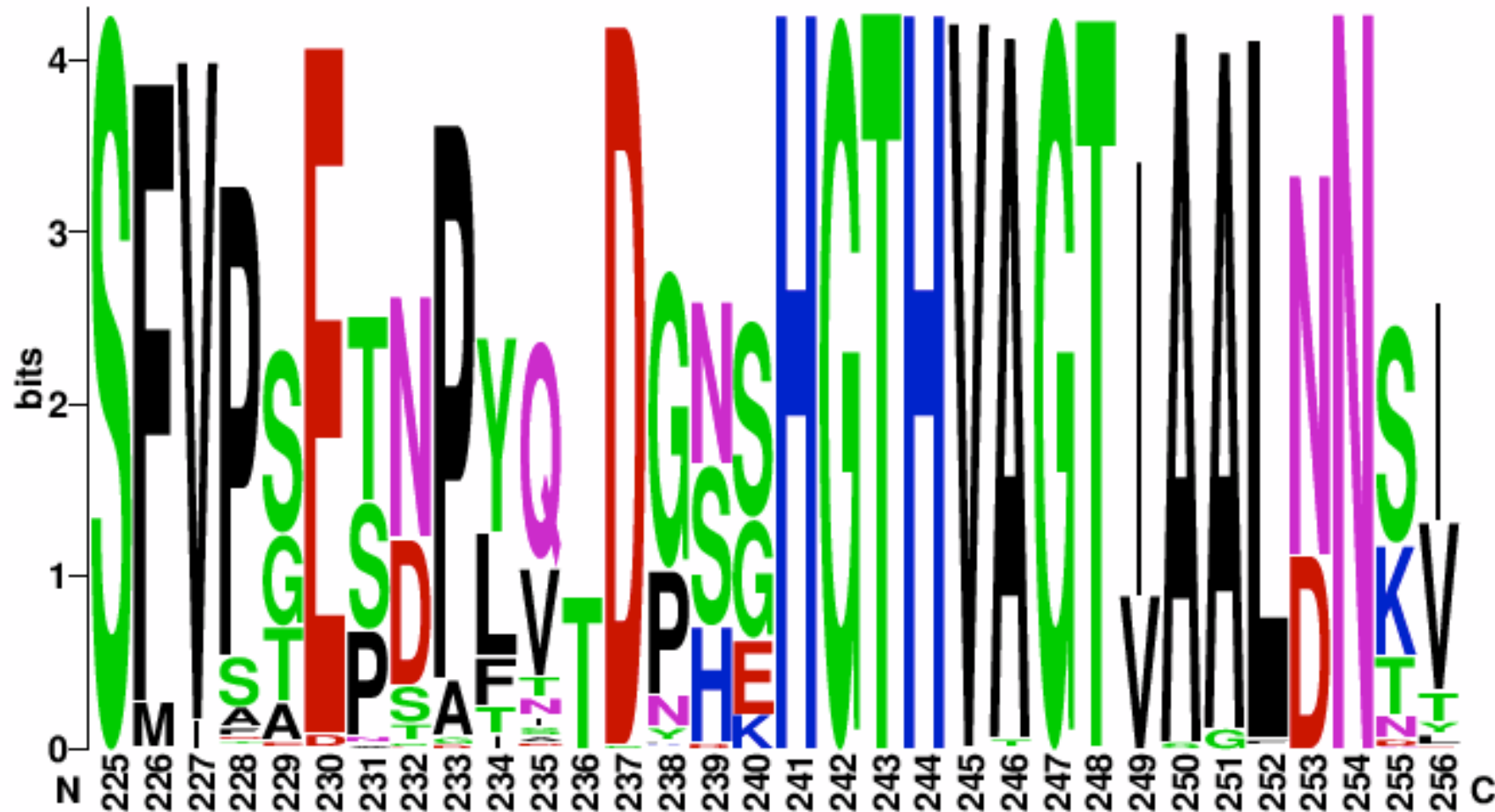
Multiple seq alignment (“MSA”)
(e.g. using MAFFT)

LOGO plot
(e.g. using WebLogo)

Seq 1
Seq 2
Seq 3
..
Seq n



LOGO example



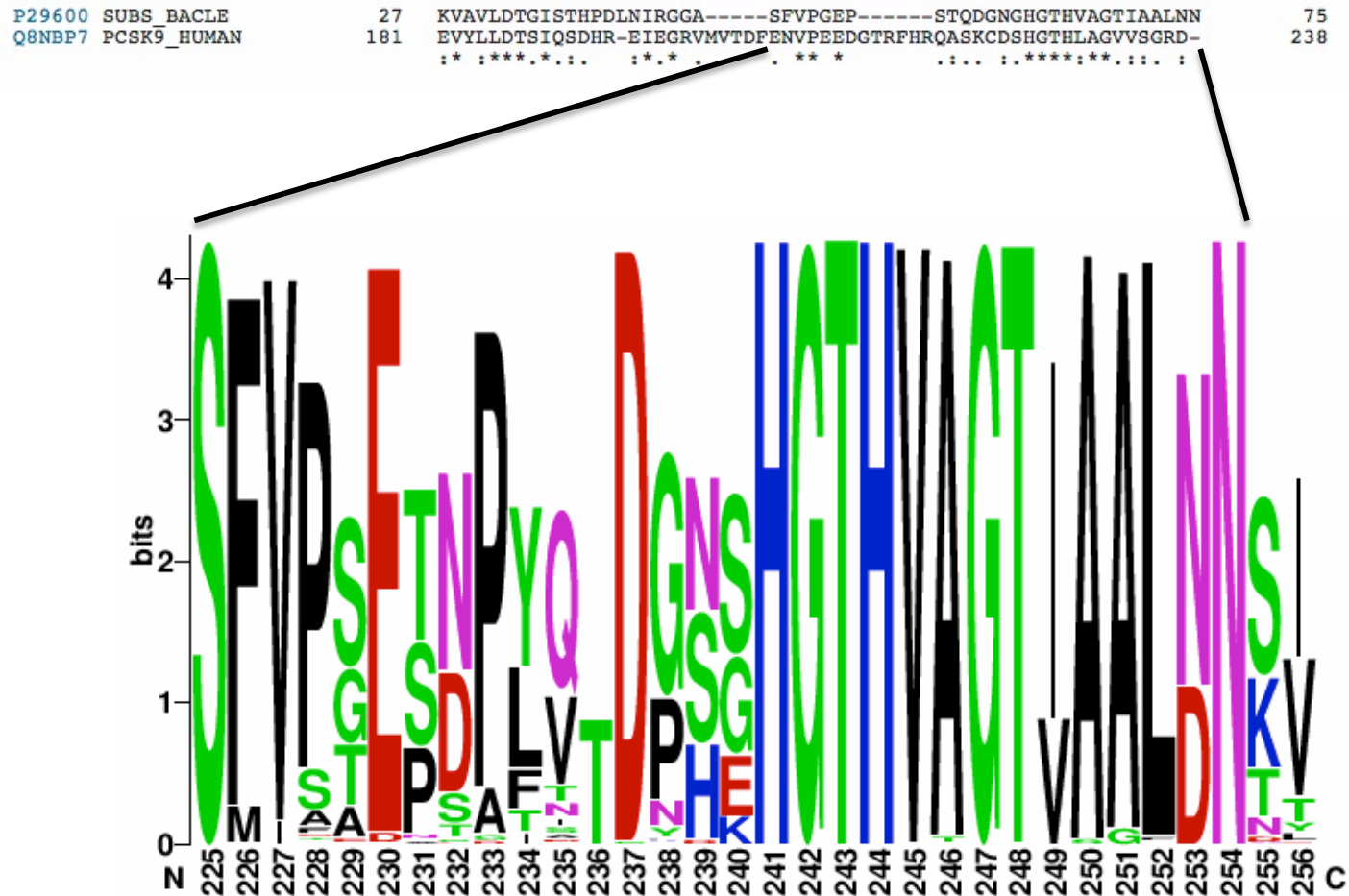
Small section of a LOGO from 1500 aligned bacterial serine proteases

Going back to pairwise alignments

P29600	SUBS_BACLE	1	-----	0
Q8NBP7	PCSK9_HUMAN	1	MGTVSSRRSWWPLPLLLLLLLLLLGPAGARAQEDEDGDYEELVLALRSEEDGLAEAPEHGT	60
P29600	SUBS_BACLE	1	-----	0
Q8NBP7	PCSK9_HUMAN	61	TATFHRCAKDPWRLPGTYVVVVLKEETHLSQSERTARRLQAQAARRGYLTKILHVFHGLLP	120
P29600	SUBS_BACLE	1	-----AQSVPWGISRVQAPAAHNRGL----TGSGV	26
Q8NBP7	PCSK9_HUMAN	121	GFLVKMSGDLELALKLPVVDYIEEDSSVFAQSIIPWNLERITPPRYRADEYQPPDGGSLV ***:***.:*: * : ** *	180
P29600	SUBS_BACLE	27	KVAVLDTGISTHPDLNIRGGA----SFVPGEPEP-----STQDGNHGHTHVAGTIAALNN	75
Q8NBP7	PCSK9_HUMAN	181	EVYLLDTSIQSDHR-EIEGRVMVTDIENVPEDGTRFHRQASKCDSHGTHLAGVVSGRD- :*:***.*.:. :*. * . . ** * :*****:***.:. :	238
P29600	SUBS_BACLE	76	SIGVLGVAPSAELYAVKVLGASGSGSVSSIAQGLEWAGNNGMHVANLSLGS--P---SPS	130
Q8NBP7	PCSK9_HUMAN	239	----AGVAKGASMRSLRVLNLCQKGTVSGTLIGLEFIRKSQLVQPVGPLVLLPLAGGYS *** .*: .: :***. .*.***. ***: .: : * * . *	294
P29600	SUBS_BACLE	131	ATLEQAVNSATSRGVLVVAASGNSGAGSISY-PARYANAMAVGATDQNNNRASF----SQ	185
Q8NBP7	PCSK9_HUMAN	295	RVLNAACQRLARAGVVLVTAAGNFRDDACLYSPASAPEVITVGATNAQDQPVTLGTGTN . *: * : : ***:***:*** .: * ** .: :*****: : : .: : : :	354
P29600	SUBS_BACLE	186	YGAGLDIVAPGVNVQSTY--PGSTYASLNGTSMATPHVAGAAALVKQKNPSWSNVQIRNH	243
Q8NBP7	PCSK9_HUMAN	355	FGRCVDLFAPEGEDIIGASSDCSTCFVSQSGTSQAAAHVAGIAAMMLSAPELTLAELRQR :* :*:*** : : .: .: .: * .*** *: ***** : : . : * . : : : : : :	414
P29600	SUBS_BACLE	244	LKNTATSLG-ST-----NLYGSGLVNAEAATR-----	269
Q8NBP7	PCSK9_HUMAN	415	LIHFSAKDVINEAWFPEDQRVLTPLNVAALPPSTHGAGWQLFCRTVWSAHSQPTRMATAV * : : : . . . : .: .*** * :	474
P29600	SUBS_BACLE	270	-----	269
Q8NBP7	PCSK9_HUMAN	475	ARCAPDEELLSCSSFSRSGKRRGERMEAQGGKLVCRAHNAFGGEGVYAIARCLLPQANC	534
P29600	SUBS_BACLE	270	-----	269
Q8NBP7	PCSK9_HUMAN	535	SVHTAPPAEASMGTRVHCHQQGHVLTGCSSHWEVEDLGTHTKPPVLRPRGQPNQCVGHREA	594
P29600	SUBS_BACLE	270	-----	269
Q8NBP7	PCSK9_HUMAN	595	SIHASCCHAPGLECKVKEHGI PAPQEQTVACEEGWTLTGCSALPGTSHVLGAYAVDNTC	654
P29600	SUBS_BACLE	270	-----	269
Q8NBP7	PCSK9_HUMAN	655	VVRSRDVSTTGSTSEGAVTAVAICCRSRHLAQAASQELQ	692

Alignment: Bacterial serine peptidase (“Savinase”) vs. human PCSK9

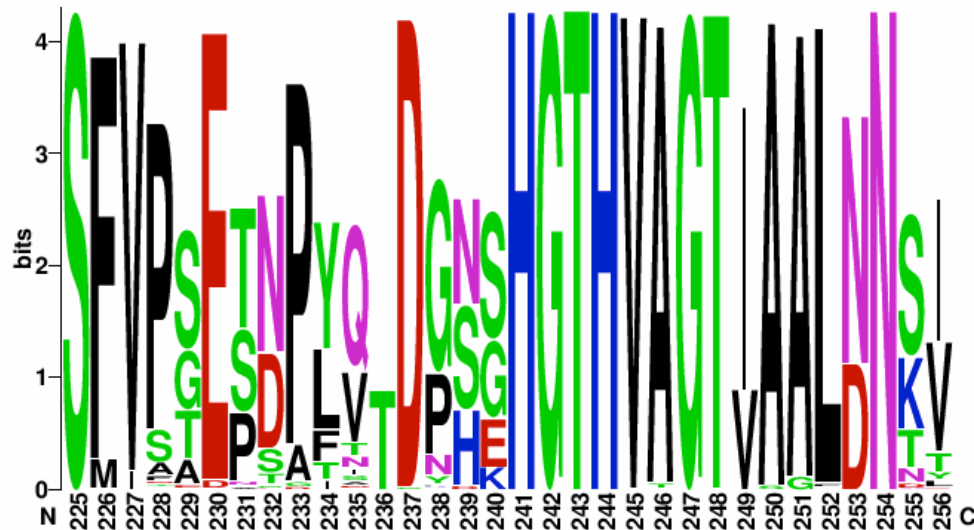
Going back to pairwise alignments



Goal: combine observations from large data set (1500 sequences) into the scoring scheme for the pairwise alignment

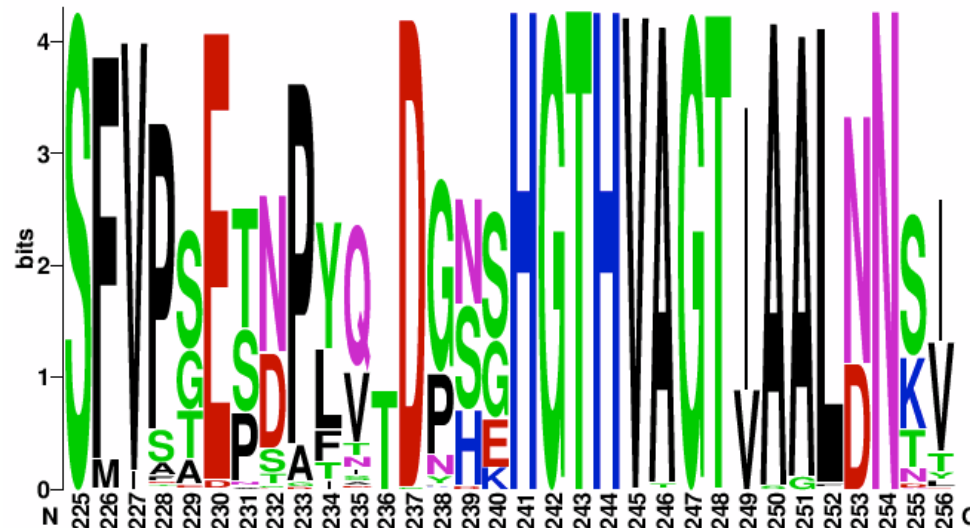
Naïve approach

- A naïve approach that would actually work:
 - When calculating the alignment score, look at how much information is in the LOGO plot (from the large data set) at the corresponding position.
 - Then scale the score from the BLOSUM62 matrix according to this.
 - That would mean that highly conserved regions would count more and variable regions would count less in the alignment score.



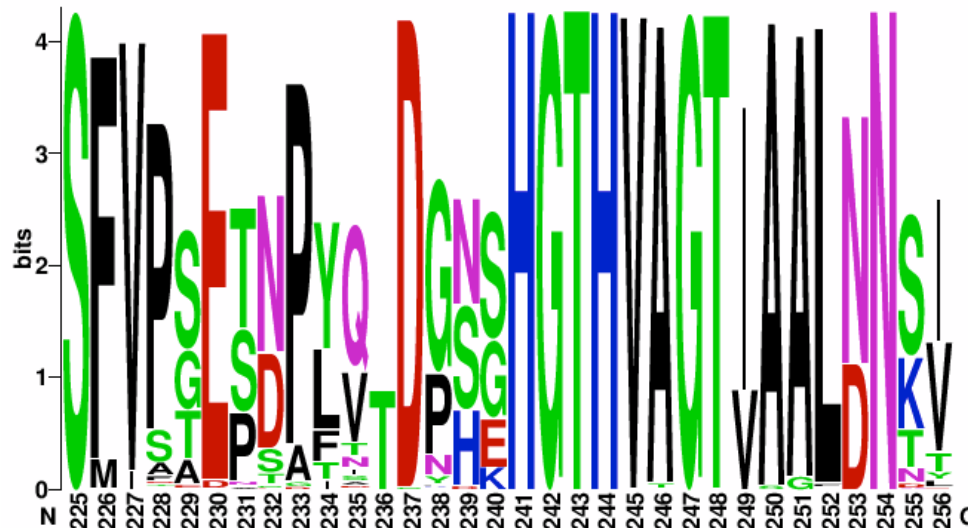
But we can actually do better

- Some things the naïve approach do not cover:
 - From the LOGO plot, a clear preference for certain amino acids at certain positions is seen.
 - We would like to build this into the model.



Weight matrices to the rescue

- Weight matrices:
 - Built from large data sets of aligned sequences.
 - Is essentially $\log_2(\text{observed}/\text{expected})$ AA frequencies (the pseudo-frequencies is a trick to cope with small data sets).
 - A score for how well new sequences match the pattern in the matrix can easily be calculated.



How to construct a WM

- A weight matrix is given as

$$W_{ij} = \log_2(p_{ij}/q_j)$$

Notice the LOG transform

- where i is a position in the motif, and j an amino acid. q_j is the background frequency for amino acid j .
- if $p_{ij} = 0$, we cannot apply the logarithm, so we have to add pseudocounts.


	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	0.6	0.4	-3.5	-2.4	-0.4	-1.9	-2.7	0.3	-1.1	1.0	0.3	0.0	1.4	1.2	-2.7	1.4	-1.2	-2.0	1.1	0.7
2	-1.6	-6.6	-6.5	-5.4	-2.5	-4.0	-4.7	-3.7	-6.3	1.0	5.1	-3.7	<u>3.1</u>	-4.2	-4.3	-4.2	-0.2	-5.9	-3.8	<u>0.4</u>
3	0.2	-1.3	0.1	1.5	0.0	-1.8	-3.3	0.4	0.5	-1.0	0.3	-2.5	1.2	1.0	-0.1	-0.3	-0.5	3.4	1.6	0.0
4	-0.1	-0.1	-2.0	2.0	-1.6	0.5	0.8	2.0	-3.3	0.1	-1.7	-1.0	-2.2	-1.6	1.7	-0.6	-0.2	1.3	-6.8	-0.7
5	-1.6	-0.1	0.1	-2.2	-1.2	0.4	-0.5	1.9	1.2	-2.2	-0.5	-1.3	-2.2	1.7	1.2	-2.5	-0.1	1.7	1.5	1.0
6	-0.7	-1.4	-1.0	-2.3	1.1	-1.3	-1.4	-0.2	-1.0	1.8	0.8	-1.9	0.2	1.0	-0.4	-0.6	0.4	-0.5	-0.0	2.1
7	1.1	-3.8	-0.2	-1.3	1.3	-0.3	-1.3	-1.4	2.1	0.6	0.7	-5.0	1.1	0.9	1.3	-0.5	-0.9	2.9	-0.4	0.5
8	-2.2	1.0	-0.8	-2.9	-1.4	0.4	0.1	-0.4	0.2	-0.0	1.1	-0.5	-0.5	0.7	-0.3	0.8	0.8	-0.7	1.3	-1.1
9	-0.2	-3.5	-6.1	-4.5	0.7	-0.8	-2.5	-4.0	-2.6	0.9	2.8	-3.0	-1.8	-1.4	-6.2	-1.9	-1.6	-4.9	-1.6	4.5

- W is a $L \times 20$ matrix, L is motif length
- $W_{ij} > 0$, Amino acid is seen **more** often than expected from random
- $W_{ij} < 0$, Amino acid is seen **less** often than expected from random

Scoring a sequence

- Score sequences to weight matrix by looking up and adding L values from the matrix

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	0.6	0.4	-3.5	-2.4	-0.4	-1.9	-2.7	0.3	-1.1	1.0	0.3	0.0	1.4	1.2	-2.7	1.4	-1.2	-2.0	1.1	0.7
2	-1.6	-6.8	-6.5	-5.4	-2.5	-4.0	-4.7	-3.7	-6.3	1.0	5.1	-3.7	3.1	-4.2	-4.3	-4.2	-0.2	-5.9	-3.8	0.4
3	0.2	-1.3	0.1	1.5	0.0	-1.8	-3.3	0.4	0.5	-1.0	0.3	-2.5	1.2	1.0	-0.1	-0.3	-0.5	3.4	1.6	0.0
4	-0.1	-0.1	-2.0	2.0	-1.6	0.5	0.8	2.0	-3.3	0.1	-1.7	-1.0	-2.2	-1.6	1.7	-0.6	-0.2	1.3	-6.8	-0.7
5	-1.6	-0.1	0.1	-2.2	-1.2	0.4	-0.5	1.9	1.2	-2.2	-0.5	-1.3	-2.2	1.7	1.2	-2.5	-0.1	1.7	1.5	1.0
6	-0.7	-1.4	-1.0	-2.3	1.1	-1.3	-1.4	-0.2	-1.0	1.8	0.8	-1.9	0.2	1.0	-0.4	-0.6	0.4	-0.5	-0.0	2.1
7	1.1	-3.8	-0.2	-1.3	1.3	-0.3	-1.3	-1.4	2.1	0.6	0.7	-5.0	1.1	0.9	1.3	-0.5	-0.5	2.9	-0.4	0.5
8	-2.2	1.0	-0.8	-2.9	-1.4	0.4	0.1	-0.4	0.2	-0.0	1.1	-0.5	-0.5	0.7	-0.3	0.8	0.8	-0.7	1.3	-1.1
9	-0.2	-3.5	-6.1	-4.5	0.7	-0.8	-2.5	-4.0	-2.6	0.9	2.8	-3.0	-1.8	-1.4	-6.2	-1.9	-1.6	-4.9	-1.6	4.5


RLLDDTPEV **11.9**
GLLGNVSTV
ALAKAAAAL

Which peptide is most likely to bind?

Which peptide second?

Scoring a sequence

- Score sequences to weight matrix by looking up and adding L values from the matrix

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	0.6	0.4	-3.5	-2.4	-0.4	-1.9	-2.7	0.3	-1.1	1.0	0.3	0.0	1.4	1.2	-2.7	1.4	-1.2	-2.0	1.1	0.7
2	-1.6	-6.6	-6.5	-5.4	-2.5	-4.0	-4.7	-3.7	-6.3	1.0	5.1	-3.7	3.1	-4.2	-4.3	-4.2	-0.2	-5.9	-3.8	0.4
3	0.2	-1.3	0.1	1.5	0.0	-1.8	-3.3	0.4	0.5	-1.0	0.3	-2.5	1.2	1.0	-0.1	-0.3	-0.5	3.4	1.6	0.0
4	-0.1	-0.1	-2.0	2.0	-1.6	0.5	0.8	2.0	-3.3	0.1	-1.7	-1.0	-2.2	-1.6	1.7	-0.6	-0.2	1.3	-6.8	-0.7
5	-1.6	-0.1	0.1	-2.2	-1.2	0.4	-0.5	1.9	1.2	-2.2	-0.5	-1.3	-2.2	1.7	1.2	-2.5	-0.1	1.7	1.5	1.0
6	-0.7	-1.4	-1.0	-2.3	1.1	-1.3	-1.4	-0.2	-1.0	1.8	0.8	-1.9	0.2	1.0	-0.4	-0.6	0.4	-0.5	-0.0	2.1
7	1.1	-3.8	-0.2	-1.3	1.3	-0.3	-1.3	-1.4	2.1	0.6	0.7	-5.0	1.1	0.9	1.3	-0.5	-0.9	2.9	-0.4	0.5
8	-2.2	1.0	-0.8	-2.9	-1.4	0.4	0.1	-0.4	0.2	-0.0	1.1	-0.5	-0.5	0.7	-0.3	0.8	0.8	-0.7	1.3	-1.1
9	-0.2	-3.5	-6.1	-4.5	0.7	-0.8	-2.5	-4.0	-2.6	0.9	2.8	-3.0	-1.8	-1.4	-6.2	-1.9	-1.6	-4.9	-1.6	4.5

RLLDDTPEV

11.9 84nM

GLLGNVSTV

14.7 23nM

ALAKAAAAL

4.3 309nM

Which peptide is most likely to bind?

Which peptide second?

Where have we seen this before?

Estimation of the BLOSUM 62 matrix

- Use the BLOCKS database (ungapped alignments of especially conserved regions of multiple alignments)
- For each alignment in the BLOCKS database the sequences are grouped into clusters with at least 62% identical residues (for BLOSUM 62)
- All pairs of sequences are compared *between* clusters, and the **observed pair frequencies** are noted



```

ID    FIBRONECTIN_2; BLOCK
COG9_CANFA  GNSAGEPCVFPFIFLQKQYSTCTREGRGDGHLWCATT
COG9_RABIT  GNADGAPCHFPPFTEGRSYTACTTDGRSDGMAWCSTT
FA12_HUMAN  LTVTGEPCHFPPQYHRQLYHKCTHKGRPGQPWCATT
HGFA_HUMAN  LTEDGRPCRFPFRYGGRLHACTSEGSABRKCATH
MANR_HUMAN  GNANGATCAFPPKFNKQYADCTSAGRSDGWLWCSTT
MPRI_MOUSE  ETDDGEPVFPFIYKGSYDECVLGRAKLWCSTAN
PB1_PIG     AITSDDKCVFPFIYKGNLYFDCTLHDSTYYWCSVTY
SFP1_BOVIN  ELPEDEECVFPFVYRNRKHFDCVHGSFLPWCSLDAD
SFP3_BOVIN  AETKDNKCVFPFIYGNKQYFDCTLHGSFLPWCSLDAD
SFP4_BOVIN  AVFEGPACAFPPFYKGGKYYMCTRKNSVLLWCSLDTE
SP1_HORSE   AATDYAKCAFPPVYRQTYDRCTTDGSLFRISWCSVT
COG2_CHICK  GNSEGAPCVFPFIFLGNKYDSCTSAGRNDGKLCWCAST
COG2_HUMAN  GNSEGAPCVFPFIFLGNKYESCTSAGRSDGKMWCAT
COG2_MOUSE  GNSEGAPCVFPFIFLGNKYESCTSAGRNDGKVCWCAT
COG2_RABIT  GNSEGAPCVFPFIFLGNKYESCTSAGRSDGKMWCATS
COG2_RAT    GNSEGAPCVFPFIFLGNKYESCTSAGRNDGKVCWCAT
COG9_BOVIN  GNADGKPCVFPFTFQGRYSACTSDGRSDGYRWCATT
COG9_HUMAN  GNADGKPCQFPFIFQGSYSACTTDGRSDGYRWCATT
COG9_MOUSE  GNGEGKPCVFPFIFEGRSYSACTTKGRSDGYRWCATT
COG9_RAT    GNGDGKPCVFPFIFEGHSYSACTTKGRSDGYRWCATT
FINC_BOVIN  GNSNGALCHFPPFLYNNHNYTDCTSEGRRDNMKWCGTT
FINC_HUMAN  GNSNGALCHFPPFLYNNHNYTDCTSEGRRDNMKWCGTT
FINC_RAT    GNSNGALCHFPPFLYNNRNSDCTSEGRRDNMKWCGTT
MPRI_BOVIN  ETEDGEPCVFPFVFNKGSYEECVVESRRLWCATTAN
MPRI_HUMAN  ETDDGVPCVFPFIFNGKSYEECIIESRAKLWCSTTAD
PA2R_BOVIN  GNAHGTPCMFPFQYNQOHHHECTREGREDNLLWCATT
PA2R_RABIT  GNAHGTPCMFPFQYNHQQHHHECTREGRODDSLWCATT
    
```

BLOSUM score = $\log_2(\text{observed pair freq}/\text{expected pair freq})$

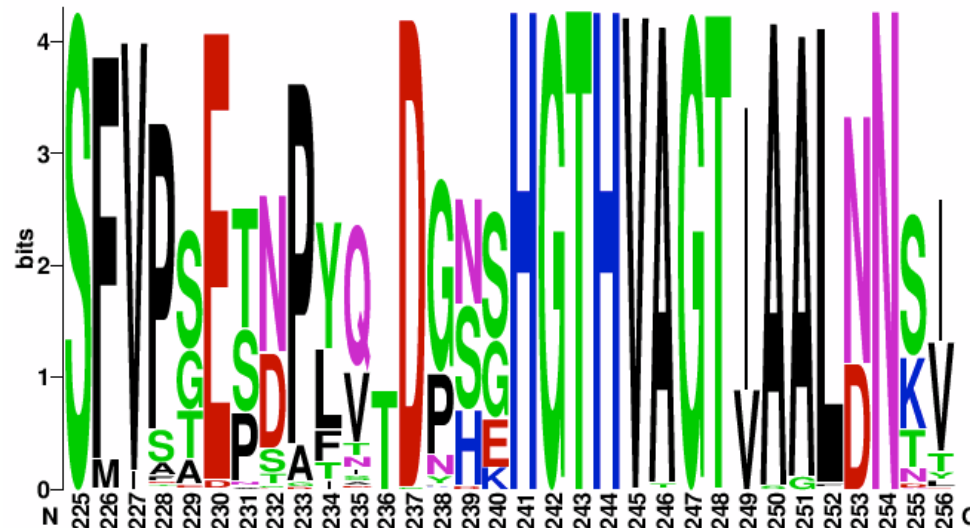
IMPORTANT: This means that BLOSUM is **not** position specific – it is a kind of an averaged across all alignment positions.

Idea: merge BLOSUM and WMs

- Pairwise alignment:
 - Alignment score = $\text{sum}(\text{BLOSUM}(\text{for each AA pair}))$
 - + penalty for gaps
 - IMPORTANT: 2 sequences
- Weight matrix:
 - WM score = $\text{sum}(\text{WM_score}(\text{for each AA, for each position}))$
 - IMPORTANT: single sequence

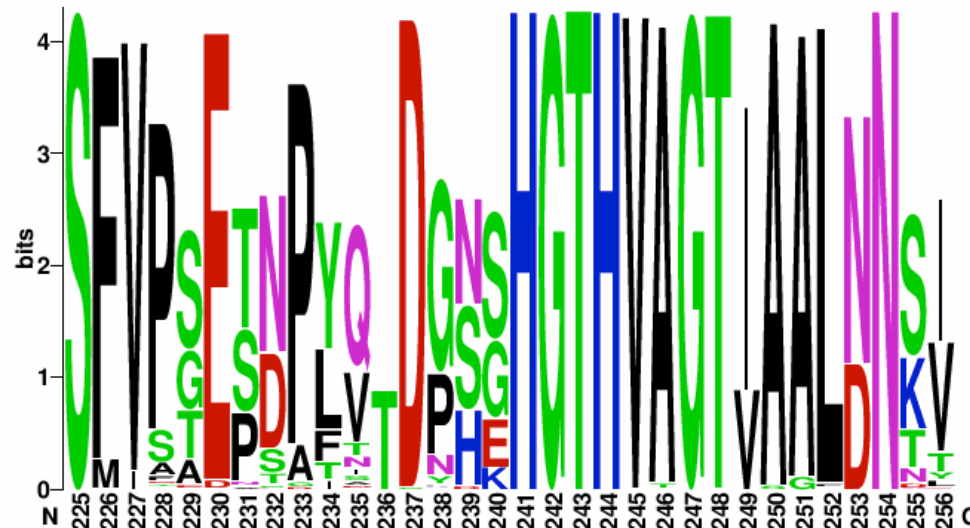
Idea: merge BLOSUM and WMs

- “New BLOSUM”:
 - Use protein family data set to estimate AA pair frequencies **per position**.
 - We need to apply the **pseudo-count** approach to account for AAs we do not observe.



Idea: merge BLOSUM and WMs

- “New alignment”:
 - Look up alignment score per position
 - Sum up score + penalize for gaps the usual way

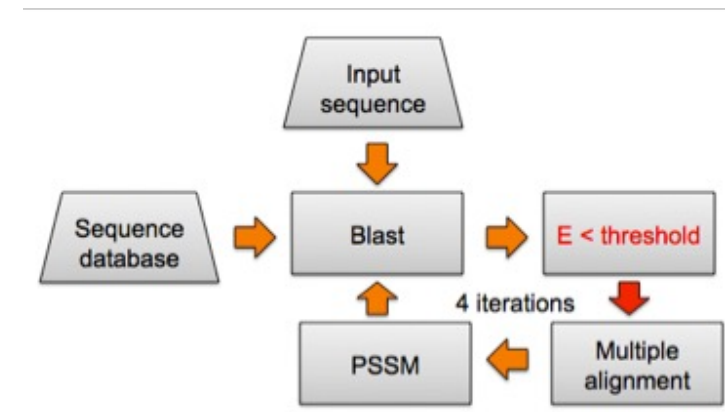
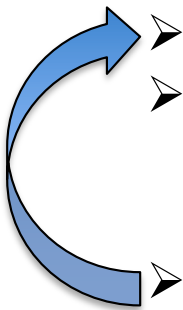


Part 3

HOW PSI-BLAST ACTUALLY WORKS

PSI-BLAST

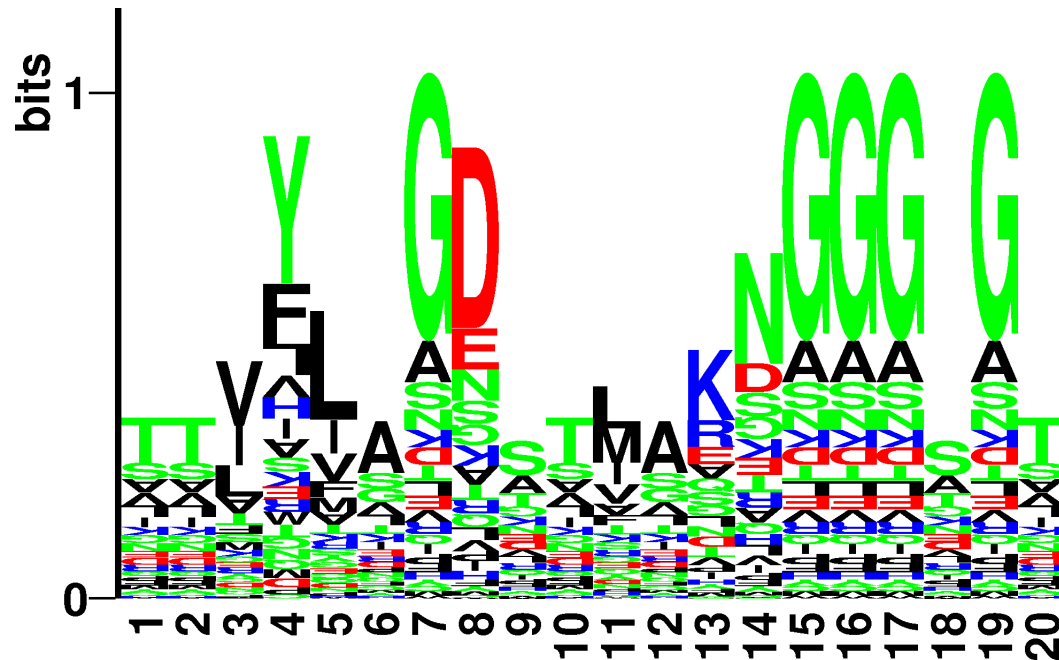
- **Position-Specific Iterative BLAST**
- Start with one sequence (as with BLASTP)
- Build protein family model on the fly:
 - **Step 0:** Start with an alignment model build purely on BLOSUM 62*
 - **Step 1:** Find set of related sequences
 - **Step 2:** Build refined **position specific** alignment model based on the identified related sequences
 - **Step 3:** Re-iterate step 1-2 until model does not improve anymore (in practice 3-4 iterations)



*The NCBI server actually “cheats” a bit here and just run BLASTP in step 0 for speed reasons

PSSM visualization

- Trick:
 - The PSSM can be visualized as a LOGO plot
 - Here's what it can look like initially (after the trivial seeding with BLOSUM62):



PSSM adjusted after each iteration

Seed: Savinase (p29600) – database: NR

<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>1</u>	A	4	3	-3	-3	-2	-2	-3	-3	-2	-2	2	0	-3	-1	-1	-2	-1	-2	-2	-2
<u>2</u>	Q	-2	-3	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	-1	-2	-2	-2	-1	0	-3	-3	-1	-2	2	5	-1	0	0	-2	-1	-2	0	0
<u>4</u>	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	-2	-4	-3	-3	-4	-3
<u>5</u>	P	-2	-3	-4	-5	-4	-4	-5	-5	8	-4	-1	-2	-4	0	-2	-3	-2	-3	1	-1
<u>6</u>	W	-4	-2	-4	-3	-4	-3	0	11	-3	-4	-4	-4	5	-5	-4	-3	-4	-4	-3	-3
<u>7</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-4	2	-3	-3	-3	-4	-2	-3
<u>8</u>	I	-2	-5	6	1	3	1	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
<u>9</u>	S	0	-2	-3	-3	-2	-1	-3	-3	4	-3	2	1	-1	0	1	0	0	-1	2	2
<u>10</u>	R	-1	-3	-3	-1	-3	-2	-1	-3	-2	-3	0	-2	0	0	3	5	2	3	0	0
<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>11</u>	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	-1	-3	-4	-4	-5	-4	-4	-4	-4
<u>12</u>	Q	-2	0	-4	-3	-3	-2	-3	-3	-2	-4	0	-2	-1	3	4	3	3	0	0	1
<u>13</u>	A	5	1	-1	-3	0	-2	-3	-4	-2	-2	1	0	-3	-3	-2	-3	-2	-3	-3	-2
<u>14</u>	P	-2	-3	-2	-2	-3	-3	-4	-5	6	-4	-1	1	-4	0	0	-2	-2	-3	3	-1
<u>15</u>	A	2	-2	-1	-1	0	0	-2	-3	-2	-2	0	1	-1	0	1	-1	2	0	1	1
<u>16</u>	A	3	-2	0	0	3	-1	-2	-3	-2	-2	0	0	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	H	-3	-4	-3	-2	-3	-2	-2	7	-4	-4	-2	-3	1	0	4	7	-2	-2	-3	-1
<u>18</u>	N	1	-1	-3	-2	-2	-1	-3	-3	-2	-3	2	0	-1	3	1	-1	1	0	1	2
<u>19</u>	R	0	-2	-2	-1	-2	-1	-2	1	-2	-3	2	0	0	1	3	-1	0	2	0	1
<u>20</u>	G	0	6	-5	-5	-4	-1	-4	-4	-3	-4	-1	-2	-4	1	-3	-3	-3	-3	-1	-2

After iteration 2

PSSM adjusted after each iteration

Seed: Savinase (p29600) – database: NR

<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>1</u>	A	3	4	-3	-3	-2	-2	-3	-3	-2	-2	2	-1	-3	-1	-2	-2	-1	-2	-2	-2
<u>2</u>	Q	-2	-4	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	0	-2	-1	-2	0	-2	-2	-4	-1	-2	2	5	-2	-1	0	-2	-1	-2	0	1
<u>4</u>	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	0	-3	-2	-3	-3	-2
<u>5</u>	P	-2	-3	-4	-4	-4	-4	-5	-5	8	-4	-1	-2	-4	-1	-2	-3	-2	-3	2	-2
<u>6</u>	W	-4	-4	-4	-3	-4	-3	0	11	-2	-4	-4	-4	5	-5	-4	-3	-4	-4	-3	-4
<u>7</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-4	0	-3	-3	-3	-4	-3	-3
<u>8</u>	I	-2	-5	6	0	3	2	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
<u>9</u>	S	-1	-2	-3	-2	-2	-2	-4	-4	4	-3	1	1	-3	0	1	-2	0	-1	2	2
<u>10</u>	R	-1	-3	-3	-2	-3	-2	-1	-3	-2	-4	0	-2	-1	0	3	5	2	3	0	0
<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>11</u>	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	0	-2	-4	-4	-4	-4	-4	-4	-4
<u>12</u>	Q	-2	-1	-4	-3	-3	-2	-3	-3	-2	-4	0	-1	-1	3	4	2	3	0	0	1
<u>13</u>	A	5	1	-1	-3	-1	-2	-3	-4	-2	-2	1	0	-3	-2	-2	-3	-2	-3	-3	-2
<u>14</u>	P	-1	-3	-2	-2	-3	-3	-4	-4	6	-3	-1	1	-3	0	1	-2	-2	-2	3	-1
<u>15</u>	A	1	-2	-1	-1	0	-1	-2	-3	-2	-2	0	1	-1	0	1	-1	1	1	1	1
<u>16</u>	A	4	-2	0	0	3	-1	-2	-3	-2	-2	0	-1	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	H	-3	-3	-3	-3	-3	-2	-2	8	-4	-4	-2	-3	1	0	4	7	-2	-2	-2	0
<u>18</u>	N	1	0	-3	-3	-2	-2	-3	-3	-2	-3	2	0	-2	3	2	-1	1	0	1	1
<u>19</u>	R	0	-2	-2	0	-2	-1	-2	1	-2	-3	2	0	0	1	3	-1	0	2	-1	0
<u>20</u>	G	0	6	-4	-4	-4	-1	-4	-4	-3	-4	0	-1	-4	1	-2	-3	-2	-3	-2	-2

After iteration 3

PSSM adjusted after each iteration

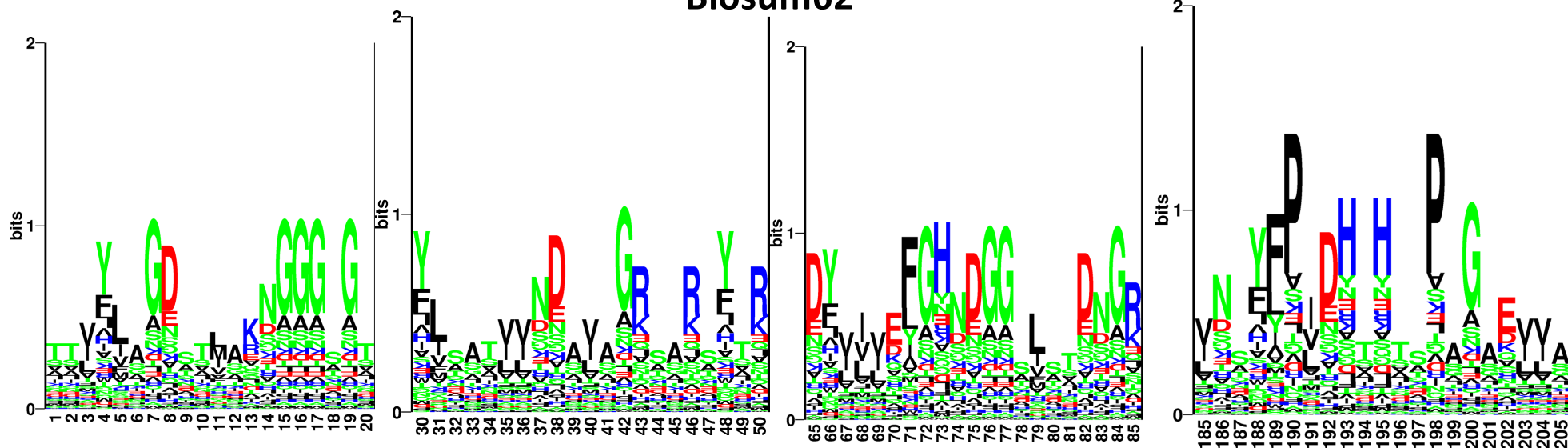
Seed: Savinase (p29600) – database: NR

<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>1</u>	A	3	3	-3	-3	-2	-2	-3	-3	-2	-2	2	-1	-3	-1	-2	-2	-1	-2	-2	-2
<u>2</u>	Q	-3	-4	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	-1	-2	-1	-2	0	-2	-2	-4	-1	-2	2	5	-2	0	0	-2	-1	-1	0	1
<u>4</u>	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	0	-3	-2	-3	-3	-1
<u>5</u>	P	-2	-2	-4	-5	-4	-4	-5	-5	8	-4	-1	-2	-4	-1	-2	-3	-2	-3	3	-2
<u>6</u>	W	-4	-5	-4	-3	-4	-3	0	12	-2	-4	-4	-4	5	-5	-4	-3	-5	-4	-3	-4
<u>7</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-5	-1	-3	-3	-3	-4	-3	-4
<u>8</u>	I	-2	-5	6	0	3	2	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
<u>9</u>	S	-1	-2	-3	-2	-3	-2	-4	-4	4	-3	1	1	-3	0	1	-2	0	-1	1	3
<u>10</u>	R	-1	-3	-3	-2	-3	-2	-1	-3	-2	-4	-1	-1	-1	0	3	5	2	3	0	0
<u>P</u>	<u>C</u>	<u>A</u>	<u>G</u>	<u>I</u>	<u>L</u>	<u>V</u>	<u>M</u>	<u>F</u>	<u>W</u>	<u>P</u>	<u>C</u>	<u>S</u>	<u>T</u>	<u>Y</u>	<u>N</u>	<u>Q</u>	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
<u>11</u>	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	0	-2	-4	-4	-4	-4	-4	-4	-4
<u>12</u>	Q	-1	-1	-4	-3	-3	-2	-3	-3	-2	-4	0	-1	-1	3	4	2	3	1	0	1
<u>13</u>	A	5	1	-2	-3	-1	-2	-3	-4	-2	-2	1	0	-3	-2	-2	-3	-2	-3	-3	-2
<u>14</u>	P	-1	-3	-2	-1	-3	-2	-4	-4	6	-3	-1	1	-3	0	1	-2	-2	-2	3	-1
<u>15</u>	A	1	-2	-1	-1	0	-1	-2	-3	-2	-2	0	1	-1	0	1	-1	1	1	1	1
<u>16</u>	A	4	-2	0	0	3	-1	-2	-3	-2	-2	0	-1	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	H	-3	-4	-3	-3	-3	-2	-2	8	-4	-4	-2	-3	1	0	4	7	-2	-2	-3	0
<u>18</u>	N	1	0	-3	-3	-2	-2	-3	-3	-2	-3	2	0	-2	3	2	-1	1	0	1	1
<u>19</u>	R	0	-2	-2	-1	-2	-1	-2	1	-2	-3	2	0	0	1	2	0	0	2	-1	0
<u>20</u>	G	-1	6	-5	-4	-4	-1	-4	-4	-3	-4	-1	-1	-4	2	-3	-3	-2	-3	-2	-2

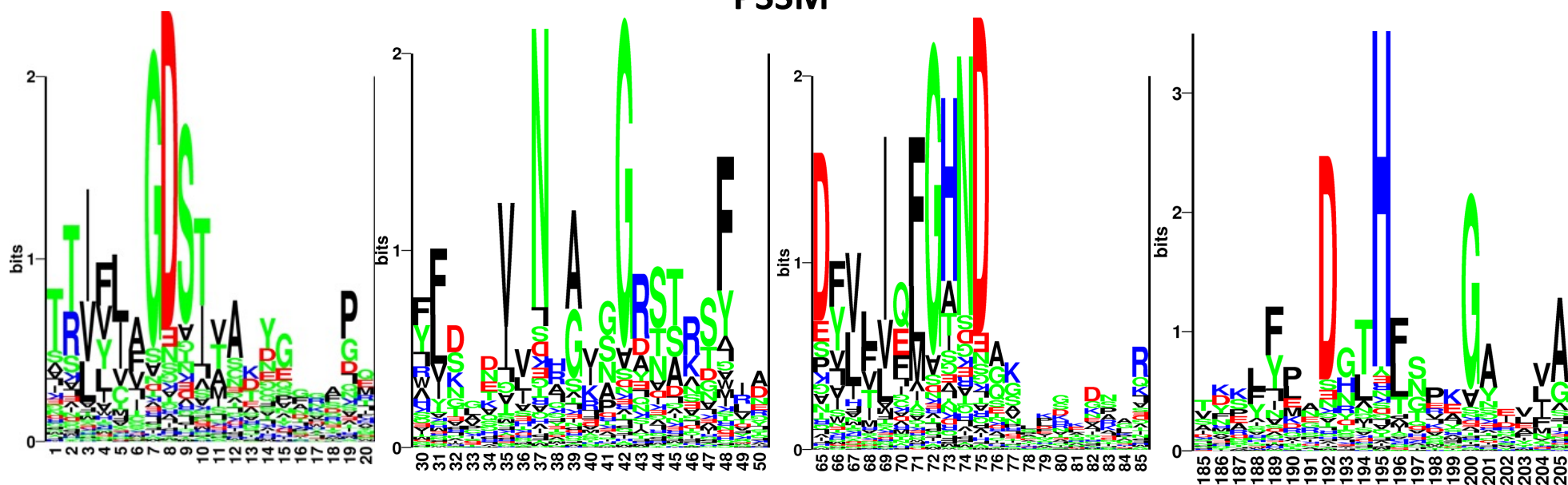
After iteration 4

Example (SGNH active site)

Blosum62



PSSM



Saving a PSSM for later use

- Very important:
 - The PSSM you have arrived at after all your iterations can be saved for later use
- Uses:
 - **Scenario 1:** Visualize your PSSM to assess the patterns picked up.
 - **Scenario 2:** Run your search again (perhaps ½ year later) without having to go through all the iterations.
 - **Scenario 3:** Search a **different database** using your PSSM
 - For example: train a rock solid PSSM for detecting prokaryotic serine peptidases on the big “NR” database, then save it and use it to hunt for human/mouse remote homologs.
 - You’ll HAVE TO do it this way, as it’s highly unlikely to find sufficiently good homologs to build the model in the restricted data set.

PSI-BLAST summary

- Is much better at finding remote homologs compared to BLASTP
 - If used correctly!
 - Remember to build your PSSM on the best possible data set, and potentially re-apply it in the actual data set you want to search
- Great for building data sets of related sequences
 - In the NCBI interface you can save all found sequences as a single pre-aligned multi FASTA file