PSI-BLAST

Fishing in the (sequence) twilight zone

Introduction to Bioinformatics, 2022
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(With some borrowed concepts / slides from Morten Nielsen, Bent Petersen, Anders Gorm Pedersen and Henrik Nielsen)

Part 1

THE PROBLEM WITH PAIRWISE ALIGNMENTS

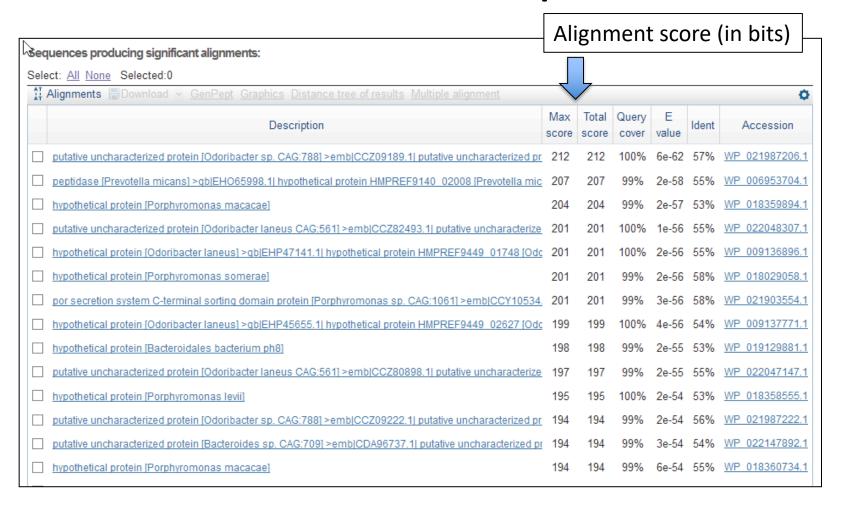
Reminder: how BLAST works

Use pairwise alignments to search databases for similar sequences

Query sequence

Database

BLASTP output



(Example from the BLAST exercise: At the protein level it was quite evident, that the unknown sequence was a serine peptidase)

BLASTP alignment

Alignment score

```
>ref|WP 006953704.1| peptidase [Prevotella micans]
Length=922
           207 bits ((526))
                            Expect = 2e-58, Method: Compositional matrix adjust.
 Score =
 Identities = 117/211 (55%), Positives = 145/211 (69%), Gaps = 14/211 (7%)
Query
            GHGTHVAGTVAAVNNNGIGVAGVAGGNGSTNSGARLMSTQIFNSDGDYTNSETLVYRAIV
            GHGTHVAGTVAA NNNG+GVAG+AGG+GSTNSG RL+S OIF
Sbict
       279 GHGTHVAGTVAARNNNGLGVAGIAGGDGSTNSGVRLLSCQIFRKSKEEGSAEA----AIK
                                                                           334
Query
       62
           YGADNGAVISONSWGSOSL-TIKELOKA---AIDYFIDYAGMDETGEIOT-GPMRGGIFI
                                                                           116
            Y ADNGAVI+O SWG S
                                 +KEL K+
                                           AIDYFI +AG D G ++ PM+GG+ I
Sbict
       335 YAADNGAVIAOCSWGYASKENVKELPKSLKEAIDYFITFAGCDAHGAORSDSPMKGGVMI
                                                                           394
Query
                                                                           176
       117 AAAGNDNVSTPNMPSAYERVLAVASMGPDFTKASYSTFGTWTDITAPGGDIDKFDLSEYG
             AAGN+N++
                         P+AYE+V++VAS
                                        +F KASYS + W
                                                       I+APGGD D F L + G
Sbjct
       395
           FAAGNENMNFKEFPAAYEKVISVASTAWNFOKASYSNYADWVSISAPGGDODAFGL-KAG
                                                                           453
Query
       177
           VLSTYADNY----YAYGEGTSMACPHVAGAA
                                             203
            VLST
                         Y Y +GTSMACPHV+G A
Sbict
       454 VLSTMPKKIASSGYGYMQGTSMACPHVSGIA
                                             484
```

(Example from the BLAST exercise: At the protein level it was quite evident, that the unknown sequence was a serine peptidase)

Alignment matrix: BLOSUM62

	A	R	N	D	С	Q	E	G	н	I	L	ĸ	M	F	P	s	т	W	Y	v	В	z	x	*
A	4	-1	-2	-2	0	-1	-1	0	-2	-1	-1	-1	-1	-2	-1	1	0	-3	-2	0	-2	-1	0	-4
R	-1	5	0	-2	-3	1	0	-2	0	-3	-2	2	-1	-3	-2	-1	-1	-3	-2	-3	-1	0	-1	-4
N	-2	0	6	1	-3	0	0	0	1	-3	-3	0	-2	-3	-2	1	0	-4	-2	-3	3	0	-1	-4
D	-2	-2	1	6	-3	0	2	-1	-1	-3	-4	-1	-3	-3	-1	0	-1	-4	-3	-3	4	1	-1	-4
С	0	-3	-3	-3	9	-3	-4	-3	-3	-1	-1	-3	-1	-2	-3	-1	-1	-2	-2	-1	-3	-3	-2	-4
Q	-1	1	0	0	-3	5	2	-2	0	-3	-2	1	0	-3	-1	0	-1	-2	-1	-2	0	3	-1	-4
E	-1	0	0	2	-4	2	5	-2	0	-3	-3	1	-2	-3	-1	0	-1	-3	-2	-2	1	4	-1	-4
G	0	-2	0	-1	-3	-2	-2	6	-2	-4	-4	-2	-3	-3	-2	0	-2	-2	-3	-3	-1	-2	-1	-4
H	-2	0	1	-1	-3	0	0	-2	8	-3	-3	-1	-2	-1	-2	-1	-2	-2	2	-3	0	0	-1	-4
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4	2	-3	1	0	-3	-2	-1	-3	-1	3	-3	-3	-1	-4
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4	-2	2	0	-3	-2	-1	-2	-1	1	-4	-3	-1	-4
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5	-1	-3	-1	0	-1	-3	-2	-2	0	1	-1	-4
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5	0	-2	-1	-1	-1	-1	1	-3	-1	-1	-4
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6	-4	-2	-2	1	3	-1	-3	-3	-1	-4
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7	-1	-1	-4	-3	-2	-2	-1	-2	-4
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4	1	-3	-2	-2	0	0	0	-4
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5	-2	-2	0	-1	-1	0	-4
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11	2	-3	-4	-3	-2	-4
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	-1	-3	-2	-1	-4
v	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4	-3	-2	-1	-4
В	-2	-1	3	4	-3	0	1	-1	0	-3	-4	0	-3	-3	-2	0	-1	-4	-3	-3	4	1	-1	-4
Z	-1	0	0	1	-3	3	4	-2	0	-3	-3	1	-1	-3	-1	0	-1	-3	-2	-2	1	4	-1	-4
Х	0	-1	-1	-1	-2	-1	-1	-1	-1	-1	-1	-1	-1	-1	-2	0	0	-2	-1	-1	-1	-1	-1	-4
*	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	1

Not all positions are biological equal

Conserved region:

Is likely important for the function of the enzyme

Query	2	GHGTHVAGTVAAVNNNGIGVAGVAGGNGSTNSGARLMSTQIFNSDGDYTNSETLVYRAIV	61
		GHGTHVAGTVAA NNNG+GVAG+AGG+GSTNSG RL+S QIF + ++E AI	
Sbjct	279	GHGTHVAGTVAARNNNGLGVAGIAGGDGSTNSGVRLLSCQIFRKSKEEGSAEAAIK	334
Query	62	YGADNGAVISONSWGSQSL-TIKELQKAAIDYFIDYAGMDETGEIQT-GPMRGGIFI	116
		Y ADNGAVI+() SWG S +KEL K+ AIDYFI +AG D G ++ PM+GG+ I	
Sbjct	335	YAADNGAVIAOCSWGYASKENVKELPKSLKEAIDYFITFAGCDAHGAQRSDSPMKGGVMI	394
Query	117	AAAGNDNVSTPNMPSAYERVLAVASMGPDFTKASYSTFGTWTDITAPGGDIDKFDLSEYG	176
		AAGN+N++ P+AYE+V++VAS +F KASYS + W I+APGGD D F L + G	
Sbjct	395	FAAGNENMNFKEFPAAYEKVISVASTAWNFQKASYSNYADWVSISAPGGDQDAFGL-KAG	453
		\	
Query	177	VLSTYADNYYAYGEGTSMACPHVAGAA 203	
		VLST Y Y +GTSMACPHV+G A	
Sbjct	454	VLSTMPKKIASSGYGYMQGTSMACPHVSGIA 484	
		\	

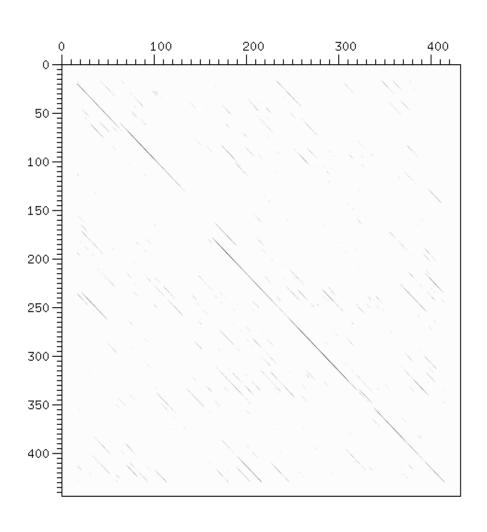
Variable region:

Is likely not that important for the function of the enzyme

Scoring of pairwise alignments

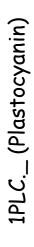
- In a normal pairwise alignment the same scores (the same matrix) is used for all positions
- As we saw before the selection pressure on the different parts of the sequence is not equal, and ideally we should take this into account
- IMPORTANT: if the sequences is of high enough similarity, this is usually not a big issue

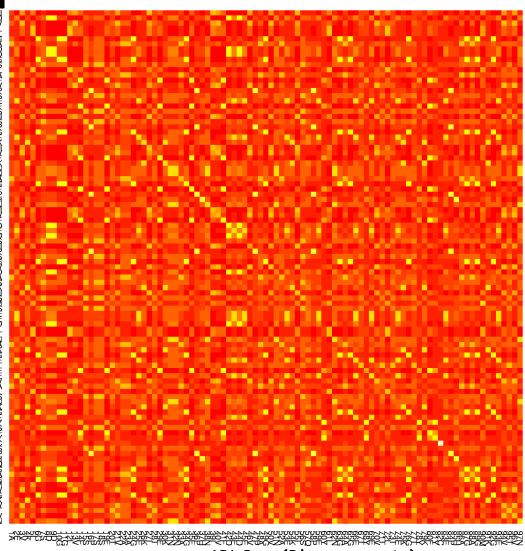
New method: Dot-plot



- Place two sequences along axes of plot
- 2. Place dot at grid points where two sequences have identical residues
- 3. Diagonals correspond to conserved regions

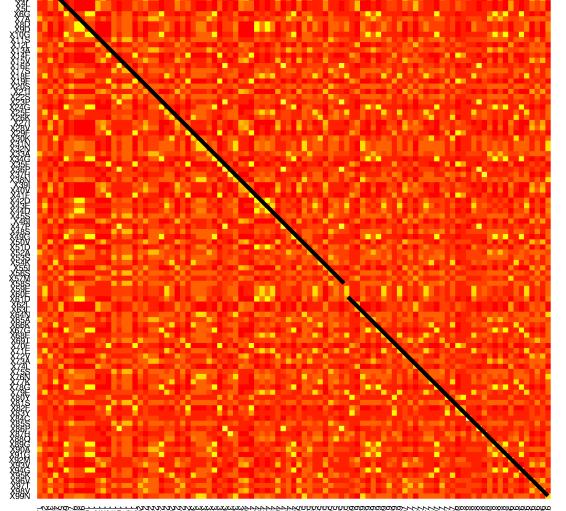
Dot-plot with BLOSUM colors





1PLB._ (Plastocyanin)

Dot-plot with BLOSUM colors



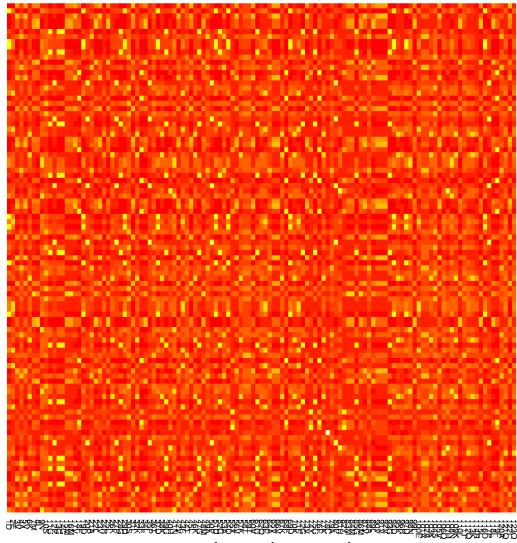
1PLC._ (Plastocyanin)

Relationship can be detected using BLASTP

1PLB._ (Plastocyanin)

Color dot-plot of **low-similarity** sequences





Relationship CANNOT be detected using BLASTP

1PMY._ (pseudoazurin)

Part 2

THOUGHTS ABOUT HOW TO SOLVE THE PROBLEM

Idea catalog

- We would like to build a scoring model for pairwise alignments that more closely resembles what happens in real sequence evolution
 - Highly conserved sites/regions should have a high weight
 - Non-conserved regions should have a low weight (be allowed to vary without counting too much against the alignment score)
- IMPORTANT: Different protein families are under different selection pressure, so our model needs to account for this

Protein families

- Tools we can use, to identify the selective pressure on protein families:
 - Data sets of truly related proteins
 - Multiple alignment
 - Logo plots
 - Weight matrices

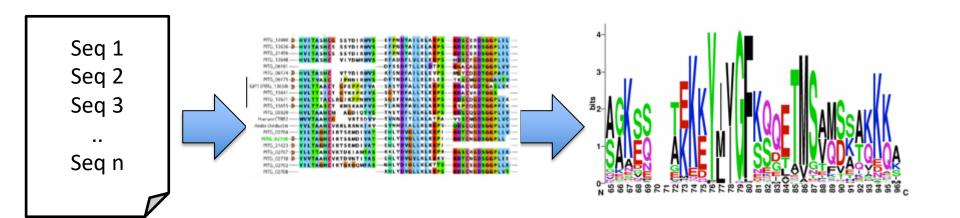
Protein family data sets

- How we can build such data sets:
 - Already known collections (literature, curated data sets)
 - Limitation: What have other people looked at before
 - "Text based" search in protein data bases (e.g. UniProt)
 - Limitation: Coverage, how well are the sequences described
 - BlastP (!)
 - Limitation: We only expect to find sequences of moderate to high similarity

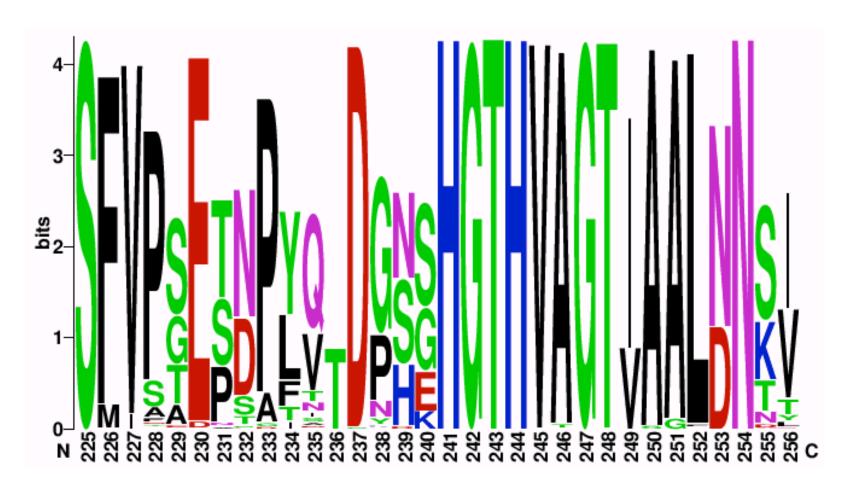
Signal across multiple sequences

Multiple seq alignment ("MSA") (e.g. using MAFFT)

LOGO plot (e.g using WebLogo)



LOGO example



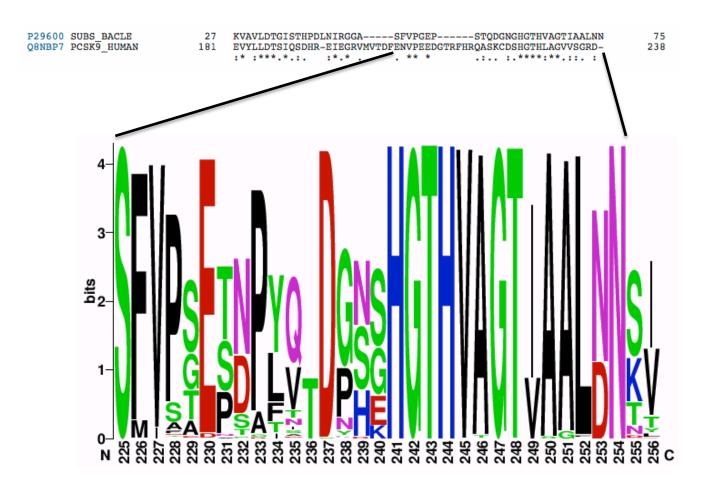
Small section of a LOGO from 1500 aligned bacterial serine proteases

Going back to pairwise alignments

P29600 SUBS_BACLE Q8NBP7 PCSK9_HUMAN	1	MGTVSSRRSWWPLPLLLLLLLLGPAGARAQEDEDGDYEELVLALRSEEDGLAEAPEHGT	0 60
P29600 SUBS_BACLE Q8NBP7 PCSK9_HUMAN	61	TATFHRCAKDPWRLPGTYVVVLKEETHLSQSERTARRLQAQAARRGYLTKILHVFHGLLP	0 120
P29600 SUBS_BACLE Q8NBP7 PCSK9_HUMAN	1 121	AQSVPWGISRVQAPAAHNRGLTGSGV GFLVKMSGDLLELALKLPHVDYIEEDSSVFAQSIPWNLERITPPRYRADEYQPPDGGSLV ***:**.:*: * : * * *	26 180
P29600 SUBS_BACLE	27	KVAVLDTGISTHPDLNIRGGA SFVPGEPSTQDGNGHGTHVAGTIAALNN EVYLLDTSIQSDHR-EIEGRVMVTDI ENVPEEDGTRFHRQASKCDSHGTHLAGVVSGRD- :* :***.:: : : : : : : : : : : : : : : :	75
Q8NBP7 PCSK9_HUMAN	181		238
P29600 SUBS_BACLE	76	SIGVLGVAPSAELYAVKVLGASGSGSVSSIAQGLEWAGNNGMHVANLSLGSPSPSAGVAKGASMRSLRVLNCQGKGTVSGTLIGLEFIRKSQLVQPVGPLVVLLPLAGGYS *** .*: :::***.*: * * **	130
Q8NBP7 PCSK9_HUMAN	239		294
P29600 SUBS_BACLE Q8NBP7 PCSK9_HUMAN	131 295	ATLEQAVNSATSRGVLVVAASGNSGAGSISY-PARYANAMAVGATDQNNNRASFSQ RVLNAACQRLARAGVVLVTAAGNFRDDACLYSPASAPEVITVGATNAQDQPVTLGTLGTN .*: *:: **::*:* :: **::::::::::::::::::	185 354
P29600 SUBS_BACLE Q8NBP7 PCSK9_HUMAN	186 355	YGAGLDIVAPGVNVQSTYPGSTYASLNGTSMATPHVAGAAALVKQKNPSWSNVQIRNH FGRCVDLFAPGEDIIGASSDCSTCFVSQSGTSQAAAHVAGTAAMMLSAEPELTLAELRQR :* :*:.** :: :: :: :: :::::::::::::::::	243 414
P29600 SUBS_BACLE	244	LKNTATSLG-STNLYGSGLVNAEAATR	269
Q8NBP7 PCSK9_HUMAN	415		474
P29600 SUBS_BACLE	270	ARCAPDEELLSCSSFSRSGKRRGERMEAQGGKLVCRAHNAFGGEGVYAIARCCLLPQANC	269
Q8NBP7 PCSK9_HUMAN	475		534
P29600 SUBS_BACLE	270	SVHTAPPAEASMGTRVHCHQQGHVLTGCSSHWEVEDLGTHKPPVLRPRGQPNQCVGHREA	269
Q8NBP7 PCSK9_HUMAN	535		594
P29600 SUBS_BACLE	270	SIHASCCHAPGLECKVKEHGIPAPQEQVTVACEEGWTLTGCSALPGTSHVLGAYAVDNTC	269
Q8NBP7 PCSK9_HUMAN	595		654
P29600 SUBS_BACLE	270	VVRSRDVSTTGSTSEGAVTAVAICCRSRHLAQASQELQ	269
Q8NBP7 PCSK9_HUMAN	655		692

Alignment: Bacterial serine peptidase ("Savinase") vs. human PCSK9

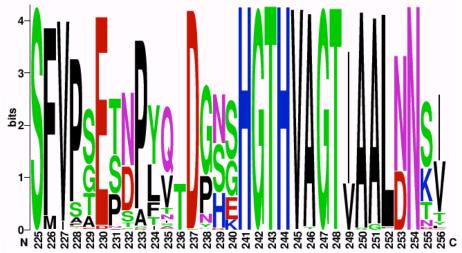
Going back to pairwise alignments



Goal: combine observations from large data set (1500 sequences) into the scoring scheme for the pairwise alignment

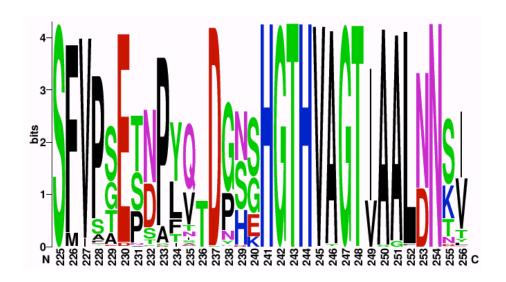
Naïve approach

- A naïve approach that would actually work:
 - When calculating the alignment score, look at how much information is in the LOGO plot (from the large data set) at the corresponding position.
 - Then scale the score from the BLOSUM62 matrix according to this.
 - That would mean that highly conserved regions would count more and variable regions would count less in the alignment score.



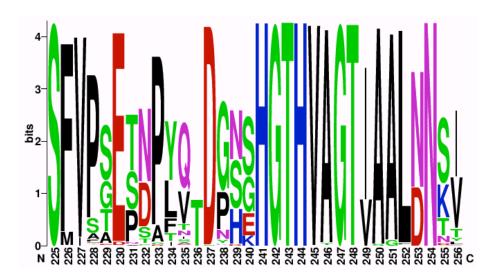
But we can actually do better

- Some things the naïve approach do not cover:
 - From the LOGO plot, a clear preference for certain amino acids at certain positions is seen.
 - We would like to build this into the model.



Weight matrices to the rescue

- Weight matrices:
 - Built from large data sets of aligned sequences.
 - Is essentially log2(observed/expected) AA frequencies (the pseudo-frequencies is a trick to cope with small data sets).
 - A score for how well new sequences match the pattern in the matrix can easily be calculated.



how to construct a WM

A weight matrix is given as

Notice the LOG transform

```
W_{ij} = \log_2(p_{ij}/q_j)
```

— where i is a position in the motif, and j an amino acid. q_j is the background frequency for amino acid j.

```
      A
      R
      N
      D
      C
      Q
      E
      G
      H
      I
      L
      K
      M
      F
      P
      S
      T
      W
      Y
      V

      1
      0.6
      0.4
      -3.5
      -2.4
      -0.4
      -1.9
      -2.7
      0.3
      -1.1
      1.0
      0.3
      0.0
      1.4
      1.2
      -2.7
      1.4
      -1.2
      -2.0
      1.1
      0.7

      2
      -1.6
      -6.5
      -5.4
      -2.5
      -4.0
      -4.7
      -3.7
      -6.3
      1.0
      5.1
      -3.7
      4.2
      -4.3
      -4.2
      -0.2
      -5.9
      -3.8
      0.4
      0.5
      -1.0
      0.3
      -2.5
      1.2
      1.2
      -4.3
      -4.2
      -0.2
      -5.9
      -3.8
      0.4
      0.5
      1.0
      0.3
      -2.5
      1.2
      1.0
      -0.1
      -0.3
      -0.5
      3.4
      1.6
      0.4
      0.0
      0.4
      0.0
      0.3
      -2.5
      1.6
      1.7
      -0.6
      -0.2
      1.3
      -6.8
      -0.7
      1.5
      1.0
      -0.1
      -0.1
      -0.2
      1.0
      -0.1
      -0.2
      1
```

- W is a L x 20 matrix, L is motif length
- Wij > 0, Amino acid is seen more often than expected from random
- Wij < 0, Amino acid is seen **less** often than expected from random

scoring a sequence

 Score sequences to weight matrix by looking up and adding L values from the matrix

```
A P N D C Q E G H I L K M F P S T W Y V 1 0.6 0.4 -3.5 -2.4 -0.4 -1.9 -2.7 0.3 -1.1 1.0 0.0 0.0 1.4 1.2 -2.7 1.4 -1.2 -2.0 1.1 0.7 2 -1.6 -6.0 -6.5 -5.4 -2.5 -4.0 -4.7 -3.7 -6.3 1.0 5 1.0 0.0 1.4 1.2 -2.7 1.4 -1.2 -2.0 1.1 0.7 3 0.2 -1.3 0.1 1.5 0.0 -1.8 -3.3 0.4 0.5 -1.0 0.3 -2.5 1.2 1.0 -0.1 -0.3 -0.5 3.4 1.6 0.0 4 -0.1 -0.1 -2.0 2-0 -1.6 0.5 0.8 2.0 -3.3 0.1 -1.7 -1.0 -2.2 -1.6 1.7 -0.6 -0.2 1.3 -6.8 -0.7 5 -1.6 -0.1 0.1 2.2 -1.2 0.4 -0.5 1.9 1.2 -2.2 -0.5 -1.3 -2.2 1.7 1.2 -2.5 -0.1 1.7 1.5 1.0 6 -0.7 -1.4 -1.0 -2.3 1.1 -1.3 -1.4 -0.2 -1.0 1.8 0.8 -1.9 0.2 1.0 -0.4 -0.6 0.4 -0.5 -0.0 2.1 7 1.1 -3.8 -0.2 -1.3 1.3 -0.3 -1.3 -1.4 2.1 0.6 0.7 -5.0 1.1 0.9 1.3 -0.5 -0.5 0.8 0.8 -0.7 1.3 -1.1 9 -0.2 -3.5 -6.1 -4.5 0.7 -0.8 -2.9 -1.4 0.4 0.2 -0.0 1.1 -0.5 -0.5 0.7 -0.3 0.8 0.8 -0.7 1.3 -1.1 9 -0.2 -3.5 -6.1 -4.5 0.7 -0.8 -2.9 -1.4 0.4 -2.6 0.9 2.8 -3.0 -1.8 -1.4 -6.2 -1.9 -1.6 -4.9 -1.6 4.5
```



RLLDDTPEV 11.9 GLLGNVSTV ALAKAAAAL Which peptide is most likely to bind?
Which peptide second?

scoring a sequence

 Score sequences to weight matrix by looking up and adding L values from the matrix

```
        A
        R
        N
        D
        C
        Q
        E
        G
        H
        I
        L
        K
        M
        F
        P
        S
        T
        W
        Y
        V

        1
        0.6
        0.4
        -3.5
        -2.4
        -0.4
        -1.9
        -2.7
        0.3
        -1.1
        1.0
        0.3
        0.0
        1.4
        1.2
        -2.7
        1.4
        -1.2
        -2.0
        1.1
        0.7

        2
        -1.6
        -6.6
        -6.5
        -5.4
        -2.5
        -4.0
        -4.7
        -3.7
        -6.3
        1.0
        5.1
        -3.7
        3.1
        -4.2
        -4.3
        -4.2
        -0.2
        -5.9
        -3.8
        0.4

        3
        0.2
        -1.3
        0.1
        1.5
        0.0
        -1.8
        -3.3
        0.4
        0.5
        -1.0
        0.3
        -2.5
        1.2
        1.0
        -0.1
        -0.3
        -0.5
        3.4
        1.6
        0.0

        4
        -0.1
        -0.1
        -2.2
        -1.2
        0.4
        -0.5
        1.9
        1.2
        -2.2
        -0.
```

RLLDDTPEV

11.9 84nM

GLLGNVSTV

14.7 23nM

ALAKAAAAL

4.3 309nM

Which peptide is most likely to

bind?

Which peptide second?

Where have we seen this before?

Estimation of the BLOSUM 62 matrix

- Use the BLOCKS database (ungapped alignments of especially conserved regions of multiple alignments)
- For each alignment in the BLOCKS database the sequences are grouped into clusters with at least 62% identical residues (for BLOSUM 62)
- All pairs of sequences are compared between clusters, and the observed pair frequencies are noted

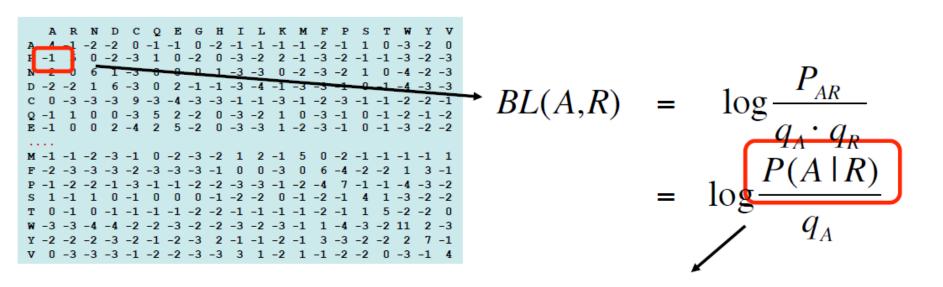


```
FIBRONECTIN 2; BLOCK
            GNSAGEPCVFPFIFLGKQYSTCTREGRGDGHLWCATT
            GNADGAPCHFPFTFEGRSYTACTTDGRSDGMAWCSTT
            LTVTGEPCHFPFQYHRQLYHKCTHKGRPGPQPWCATT
HGFA HUMAN
            LTEDGRPCRFPFRYGGRMLHACTSEGSAHRKW
            GNANGATCAFPFKFENKWYADCTSAGRSDGWLWCGTT
MPRI MOUSE
            ETDDGEPCVFPFIYKGKSYDECVLEGRAKLWCSKTAN
PB1 PIG
SFP1 BOVIN
            ELPEDEECVFPFVYRNRKHFDCTVHGSLFPWCSLDAD
SFP3 BOVIN
SFP4 BOVIN
            AVFEGPACAFPFTYKGKKYYMCTRKNSVLLWCSLDTE
SP1 HORSE
            AATDYAKCAFPFVYRGOTYDRCTTDGSLFRISWCSVT
COG2 CHICK
            GNSEGAPCVFPFIFLGNKYDSCTSAGRNDGKLWCAST
COG2 HUMAN
            GNSEGAPCVFPFTFLGNKYESCTSAGRSDGKMWCATT
COG2 MOUSE
COG2 RABIT
COG2 RAT
            GNSEGAPCVFPFTFLGNKYESCTSAGRNDGKVWCATT
COG9 BOVIN
            GNADGKPCVFPFTFQGRTYSACTSDGRSDGYRWCATT
COG9 HUMAN
            GNADGKPCQFPFIFQGQSYSACTTDGRSDGYRWCATT
COG9 MOUSE
            GNGEGKPCVFPFIFEGRSYSACTTKGRSDGYR
COG9 RAT
FINC BOVIN
FINC HUMAN
            GNSNGALCHFPFLYNNHNYTDCTSEGRRDNMKWCGTT
MPRI HUMAN
            ETDDGVPCVFPFIFNGKSYEECIIESRAKLWCSTTAD
PA2R BOVIN
            GNAHGTPCMFPFQYNQQWHHECTREGREDNLLWCATT
PA2R RABIT
            GNAHGTPCMFPFQYNHQWHHECTREGRQDDSLWCATT
```

BLOSUM score = log2(observed pair freq/expected pair freq)

IMPORTANT: This means that BLOSUM is **not** position specific – it is a kind of an average across all alignment positions.

The two views on BLOSUM



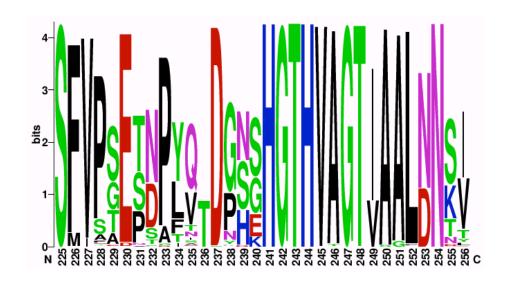
Idea: merge BLOSUM and WMs

- Pair wise alignment:
 - Alignment score = sum(BLOSUM(for each AA pair))
 - + penalty for gaps
 - IMPORTANT: 2 sequences

- Weight matrix:
 - WM score = sum(WM_score(for each AA, for each position))
 - IMPORTANT: single sequence

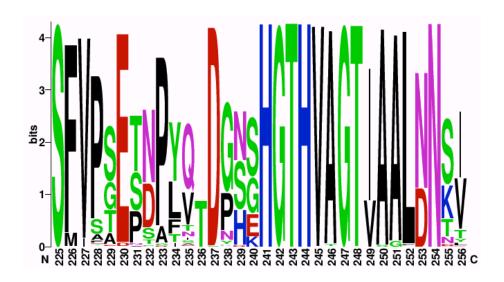
Idea: merge BLOSUM and WMs

- "New BLOSUM":
 - Use protein family data set to estimate AA pair frequencies per position.
 - We need to apply the **pseudo-count** approach to account for AAs we do not observe.



Idea: merge BLOSUM and WMs

- "New alignment":
 - Look up alignment score per position
 - Sum up score + penalize for gaps the usual way

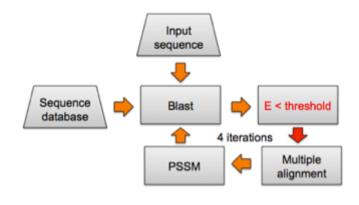


Part 3

HOW PSI-BLAST ACTUALLY WORKS

PSI-BLAST

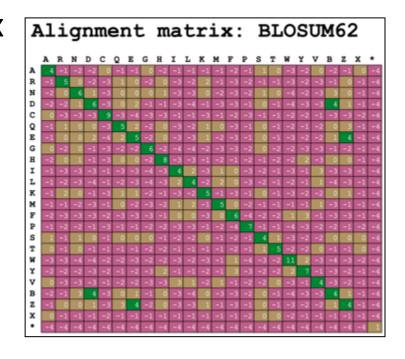
- Position-Specific Iterative BLAST
- Start with one sequence (as with BLASTP)
- Build protein family model on the fly:
 - > Step 0: Start with an alignment model build purely on BLOSUM 62*
 - **Step 1:** Find set of related sequences
 - **Step 2:** Build refined **position specific** alignment model based on the identified related sequences
 - **Step 3:** Re-**iterate** step 1-2 until model does not improve anymore (in practice 3-4 iterations)



^{*}The NCBI server actually "cheats" a bit here and just run BLASTP in step 0 for speed reasons

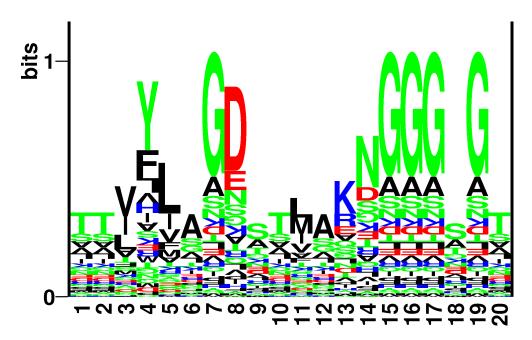
PSSM

- PSSM (pronounced "PoSSoM"):
 - Position-Specific Scoring Matrix
- Start by creating a n*20 matrix
 - n = length of input sequence
- For each AA in the input sequence look up the corresponding row in the BLOSUM62 matrix and copy in the values



PSSM visualization

- Trick:
 - The PSSM can be visualized as a LOGO plot
 - Here's what it can look like initially (after the trivial seeding with BLOSUM62):



PSSM adjusted after each iteration

Seed: Savinase (p29600) – database: NR

<u>P</u>	С	A	G	Ī	<u>L</u>	v	<u>M</u>	E	w	<u>P</u>	<u>c</u>	<u>s</u>	I	Y	N	Q	<u>H</u>	K	R	D	E
1	Α	4	3	-3	-3	-2	-2	-3	-3	-2	-2	2	0	-3	-1	-1	-2	-1	-2	-2	-2
2	Q	-2	-3	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	-1	-2	-2	-2	-1	0	-3	-3	-1	-2	2	5	-1	0	0	-2	-1	-2	0	0
4	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	-2	-4	-3	-3	-4	-3
<u>5</u>	Р	-2	-3	-4	-5	-4	-4	-5	-5	8	-4	-1	-2	-4	0	-2	-3	-2	-3	1	-1
<u>6</u>	W	-4	-2	-4	-3	-4	-3	0	11	-3	-4	-4	-4	5	-5	-4	-3	-4	-4	-3	-3
<u>Z</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-4	2	-3	-3	-3	-4	-2	-3
<u>8</u>	I	-2	-5	6	1	3	1	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
9	S	0	-2	-3	-3	-2	-1	-3	-3	4	-3	2	1	-1	0	1	0	0	-1	2	2
<u>10</u>	R	-1	-3	-3	-1	-3	-2	-1	-3	-2	-3	0	-2	0	0	3	5	2	3	0	0
<u>P</u>	С	<u>A</u>	<u>G</u>	Ī	<u>L</u>	<u>v</u>	<u>M</u>	<u>E</u>	<u>w</u>	<u>P</u>	<u>C</u>	<u>s</u>	I	<u>Y</u>	<u>N</u>	Q	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
11	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	-1	-3	-4	-4	-5	-4	-4	-4	-4
<u>12</u>	Q	-2	0	-4	-3	-3	-2	-3	-3	-2	-4	0	-2	-1	3	4	3	3	0	0	1
<u>13</u>	Α	5	1	-1	-3	0	-2	-3	-4	-2	-2	1	0	-3	-3	-2	-3	-2	-3	-3	-2
<u>14</u>	Р	-2	-3	-2	-2	-3	-3	-4	-5	6	-4	-1	1	-4	0	0	-2	-2	-3	3	-1
<u>15</u>	Α	2	-2	-1	-1	0	0	-2	-3	-2	-2	0	1	-1	0	1	-1	2	0	1	1
<u>16</u>	Α	3	-2	0	0	3	-1	-2	-3	-2	-2	0	0	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	Н	-3	-4	-3	-2	-3	-2	-2	7	-4	-4	-2	-3	1	0	4	7	-2	-2	-3	-1
<u>18</u>	N	1	-1	-3	-2	-2	-1	-3	-3	-2	-3	2	0	-1	3	1	-1	1	0	1	2
<u>19</u>	R	0	-2	-2	-1	-2	-1	-2	1	-2	-3	2	0	0	1	3	-1	0	2	0	1
<u>20</u>	G	0	6	-5	-5	-4	-1	-4	-4	-3	-4	-1	-2	-4	1	-3	-3	-3	-3	-1	-2

After iteration 2

PSSM adjusted after each iteration

Seed: Savinase (p29600) – database: NR

<u>P</u>	С	A	G	Ī	<u>L</u>	v	<u>M</u>	E	w	<u>P</u>	<u>c</u>	<u>s</u>	I	Y	N	Q	<u>H</u>	<u>K</u>	R	D	E
1	Α	3	4	-3	-3	-2	-2	-3	-3	-2	-2	2	-1	-3	-1	-2	-2	-1	-2	-2	-2
2	Q	-2	-4	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	0	-2	-1	-2	0	-2	-2	-4	-1	-2	2	5	-2	-1	0	-2	-1	-2	0	1
4	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	0	-3	-2	-3	-3	-2
<u>5</u>	P	-2	-3	-4	-4	-4	-4	-5	-5	8	-4	-1	-2	-4	-1	-2	-3	-2	-3	2	-2
<u>6</u>	W	-4	-4	-4	-3	-4	-3	0	11	-2	-4	-4	-4	5	-5	-4	-3	-4	-4	-3	-4
<u>Z</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-4	0	-3	-3	-3	-4	-3	-3
<u>8</u>	I	-2	-5	6	0	3	2	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
9	S	-1	-2	-3	-2	-2	-2	-4	-4	4	-3	1	1	-3	0	1	-2	0	-1	2	2
<u>10</u>	R	-1	-3	-3	-2	-3	-2	-1	-3	-2	-4	0	-2	-1	0	3	5	2	3	0	0
<u>P</u>	С	A	<u>G</u>	Ī	<u>L</u>	<u>v</u>	<u>M</u>	<u>E</u>	<u>w</u>	<u>P</u>	<u>c</u>	<u>s</u>	I	<u>Y</u>	<u>N</u>	Q	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
11	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	0	-2	-4	-4	-4	-4	-4	-4	-4
<u>12</u>	Q	-2	-1	-4	-3	-3	-2	-3	-3	-2	-4	0	-1	-1	3	4	2	3	0	0	1
<u>13</u>	Α	5	1	-1	-3	-1	-2	-3	-4	-2	-2	1	0	-3	-2	-2	-3	-2	-3	-3	-2
<u>14</u>	Р	-1	-3	-2	-2	-3	-3	-4	-4	6	-3	-1	1	-3	0	1	-2	-2	-2	3	-1
<u>15</u>	Α	1	-2	-1	-1	0	-1	-2	-3	-2	-2	0	1	-1	0	1	-1	1	1	1	1
<u>16</u>	Α	4	-2	0	0	3	-1	-2	-3	-2	-2	0	-1	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	Н	-3	-3	-3	-3	-3	-2	-2	8	-4	-4	-2	-3	1	0	4	7	-2	-2	-2	0
<u>18</u>	N	1	0	-3	-3	-2	-2	-3	-3	-2	-3	2	0	-2	3	2	-1	1	0	1	1
<u>19</u>	R	0	-2	-2	0	-2	-1	-2	1	-2	-3	2	0	0	1	3	-1	0	2	-1	0
<u>20</u>	G	0	6	-4	-4	-4	-1	-4	-4	-3	-4	0	-1	-4	1	-2	-3	-2	-3	-2	-2

After iteration 3

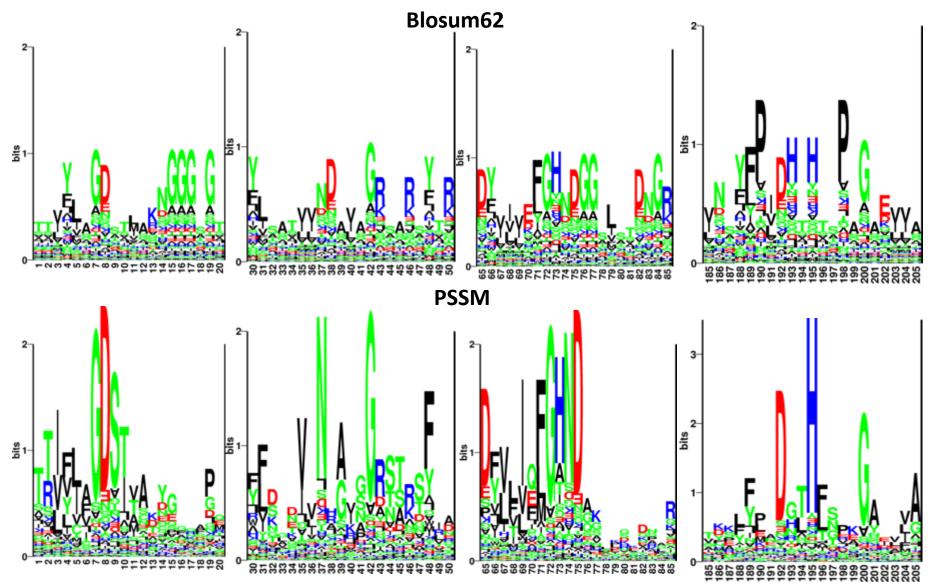
PSSM adjusted after each iteration

Seed: Savinase (p29600) – database: NR

<u>P</u>	С	A	G	Ī	<u>L</u>	v	<u>M</u>	E	w	<u>P</u>	<u>c</u>	<u>s</u>	I	Y	N	Q	Н	K	R	D	Ē
1	Α	3	3	-3	-3	-2	-2	-3	-3	-2	-2	2	-1	-3	-1	-2	-2	-1	-2	-2	-2
2	Q	-3	-4	-5	-4	-4	-2	-5	-4	-3	-5	-2	-2	-3	-2	8	-1	0	-1	-2	1
<u>3</u>	S	-1	-2	-1	-2	0	-2	-2	-4	-1	-2	2	5	-2	0	0	-2	-1	-1	0	1
4	V	-1	-4	3	0	4	2	-1	-3	-3	-2	-1	2	-2	-3	0	-3	-2	-3	-3	-1
<u>5</u>	Р	-2	-2	-4	-5	-4	-4	-5	-5	8	-4	-1	-2	-4	-1	-2	-3	-2	-3	3	-2
<u>6</u>	W	-4	-5	-4	-3	-4	-3	0	12	-2	-4	-4	-4	5	-5	-4	-3	-5	-4	-3	-4
<u>Z</u>	G	-1	7	-5	-5	-5	-4	-5	-4	-4	-4	-1	-3	-5	-1	-3	-3	-3	-4	-3	-4
<u>8</u>	I	-2	-5	6	0	3	2	0	-3	-4	-2	-3	-2	0	-4	-4	-4	-4	-4	-4	-4
9	S	-1	-2	-3	-2	-3	-2	-4	-4	4	-3	1	1	-3	0	1	-2	0	-1	1	3
<u>10</u>	R	-1	-3	-3	-2	-3	-2	-1	-3	-2	-4	-1	-1	-1	0	3	5	2	3	0	0
<u>P</u>	С	<u>A</u>	<u>G</u>	Ī	<u>L</u>	<u>v</u>	<u>M</u>	<u>E</u>	<u>w</u>	<u>P</u>	<u>C</u>	<u>s</u>	I	<u>Y</u>	<u>N</u>	Q	<u>H</u>	<u>K</u>	<u>R</u>	<u>D</u>	<u>E</u>
11	V	-2	-5	5	0	4	0	-2	-4	-4	-2	-3	0	-2	-4	-4	-4	-4	-4	-4	-4
<u>12</u>	Q	-1	-1	-4	-3	-3	-2	-3	-3	-2	-4	0	-1	-1	3	4	2	3	1	0	1
<u>13</u>	Α	5	1	-2	-3	-1	-2	-3	-4	-2	-2	1	0	-3	-2	-2	-3	-2	-3	-3	-2
<u>14</u>	Р	-1	-3	-2	-1	-3	-2	-4	-4	6	-3	-1	1	-3	0	1	-2	-2	-2	3	-1
<u>15</u>	Α	1	-2	-1	-1	0	-1	-2	-3	-2	-2	0	1	-1	0	1	-1	1	1	1	1
<u>16</u>	Α	4	-2	0	0	3	-1	-2	-3	-2	-2	0	-1	-1	-3	-2	-3	-2	-3	-2	-2
<u>17</u>	Н	-3	-4	-3	-3	-3	-2	-2	8	-4	-4	-2	-3	1	0	4	7	-2	-2	-3	0
<u>18</u>	N	1	0	-3	-3	-2	-2	-3	-3	-2	-3	2	0	-2	3	2	-1	1	0	1	1
<u>19</u>	R	0	-2	-2	-1	-2	-1	-2	1	-2	-3	2	0	0	1	2	0	0	2	-1	0
<u>20</u>	G	-1	6	-5	-4	-4	-1	-4	-4	-3	-4	-1	-1	-4	2	-3	-3	-2	-3	-2	-2

After iteration 4

Example. (SGNH active site)



Saving a PSSM for later use

Very important:

 The PSSM you have arrived at after all your iterations can be saved for later use

Uses:

- Scenario 1: Visualize your PSSM to assess the patterns picked up.
- Scenario 2: Run your search again (perhaps ½ year later) with out having to go through all the iterations.
- Scenario 3: Search a different database using your PSSM
 - For example: train a rock solid PSSM for detecting prokaryotic serine peptidases on the big "NR" database, then save it and use it to hunt for human/mouse remote homologs.
 - You'll HAVE TO do it this way, as it's highly unlikely to find sufficiently good homologs to build the model in the restricted data set.

PSI-BLAST summary

- Is much better at finding remote homologs compared to BLASTP
 - If used correctly!
 - Remember to build your PSSM on the best possible data set, and potentially re-apply it in the actual data set you want to search
- Great for building data sets of related sequences
 - In the NCBI interface you can save all found sequences as a single pre-aligned multi FASTA file